

# Structure of model biological membranes with neutron scattering: present developments and future perspectives

Giovanna Fragneto  
European Spallation Source ERIC

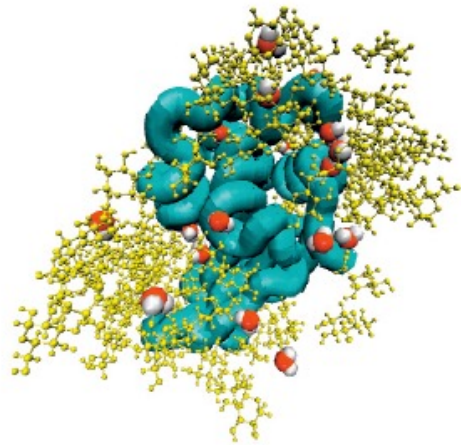


Uncharged, subatomic particles found in atomic nuclei  
Approx. mass of neutron  $1.67 \times 10^{-27}$  kg,  $v = 2.2$  km/s at RT  
energy  $\sim 0.025$  eV, wave-particle duality,  $\lambda = 0.18$  nm at RT

# Neutrons: An ideal probe at the atomic scale

- Like X-rays thermal neutrons possess the right wavelengths.
- In addition neutrons possess the ideal energies for spectroscopy of thermal fluctuations.

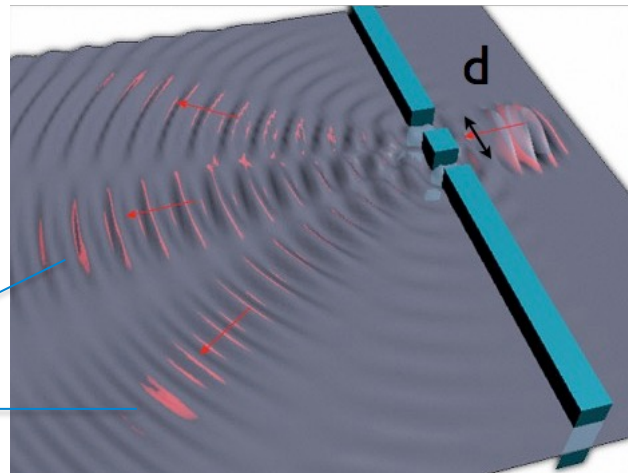
From 1000 nm



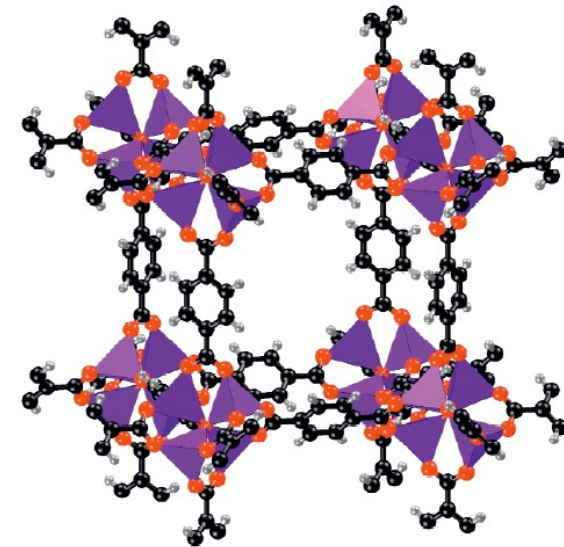
Cold Neutrons

$$\Delta\phi = \frac{\Delta y}{D} = \frac{\lambda}{d}$$

*The information is encoded in the change of direction and speed of the neutrons as they path through the material.*



Up to 0.001 nm



Hot Neutrons

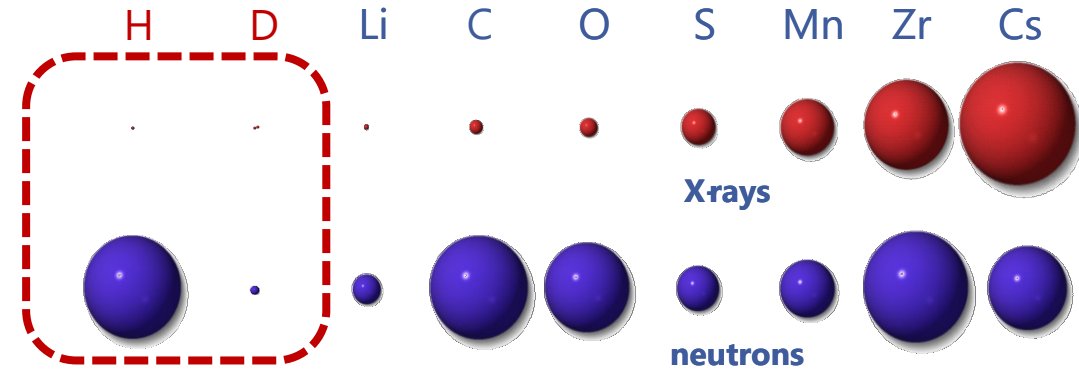


# Why use neutrons to study soft and biological material?

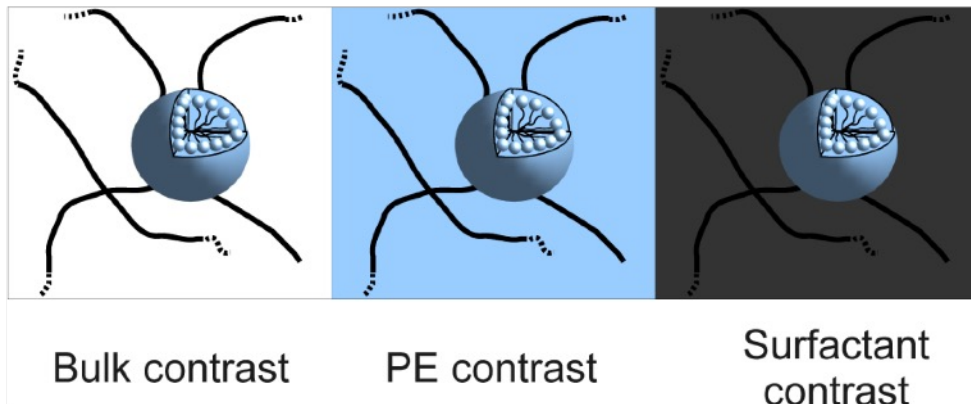
## Neutrons interact with nuclei



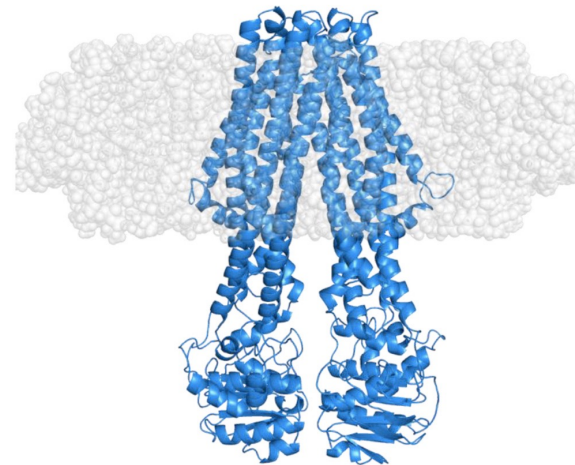
- are sensitive to light atoms, particularly hydrogen
- can exploit isotopic substitution, especially H/D
- 'see' materials differently to X-rays, complementary



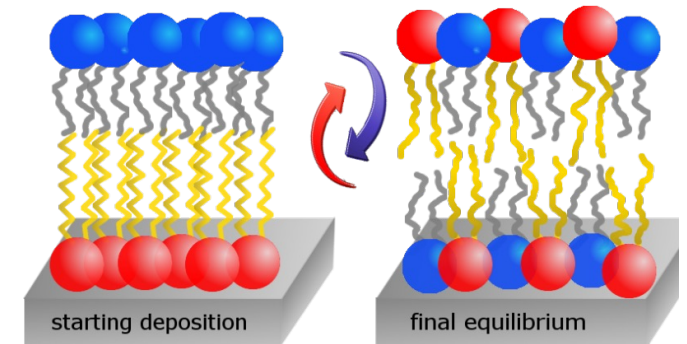
### Polyelectrolyte + surfactant complex



Hoffmann et al. *J. Chem. Phys.* 2015.



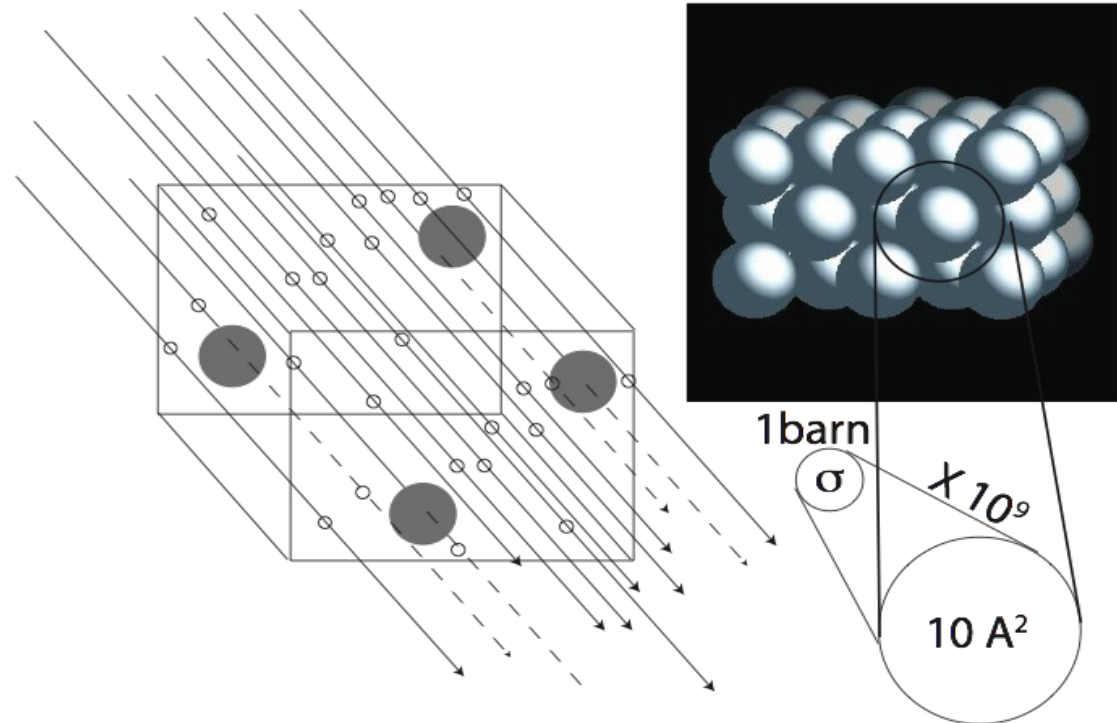
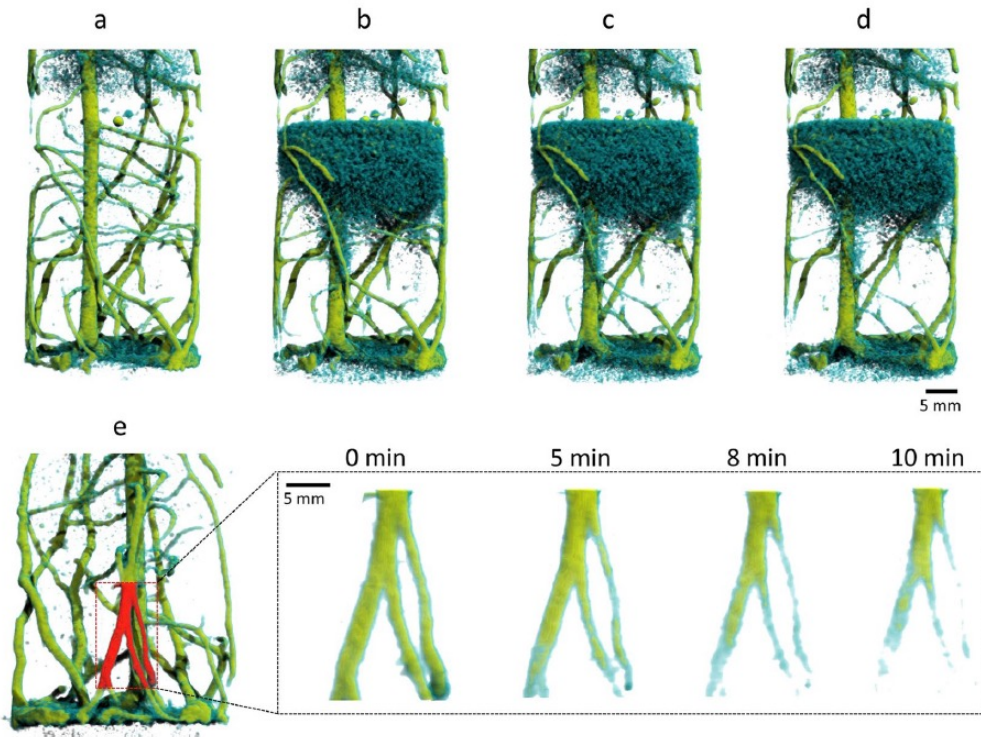
Josts et al. *Structure* 2018



Gerelli Y., et al., *Langmuir* 2012

# Neutrons are a neutral particle

- are highly penetrating → *buried interfaces*
- can be used as non-destructive probes
- can be used to study samples in extreme environments



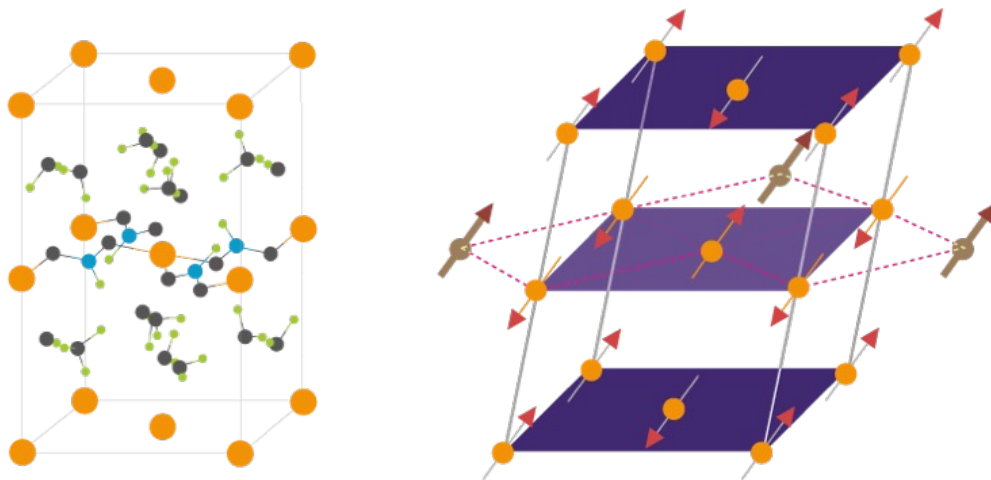




# Neutrons have a spin, therefore a sensitivity to magnetic properties

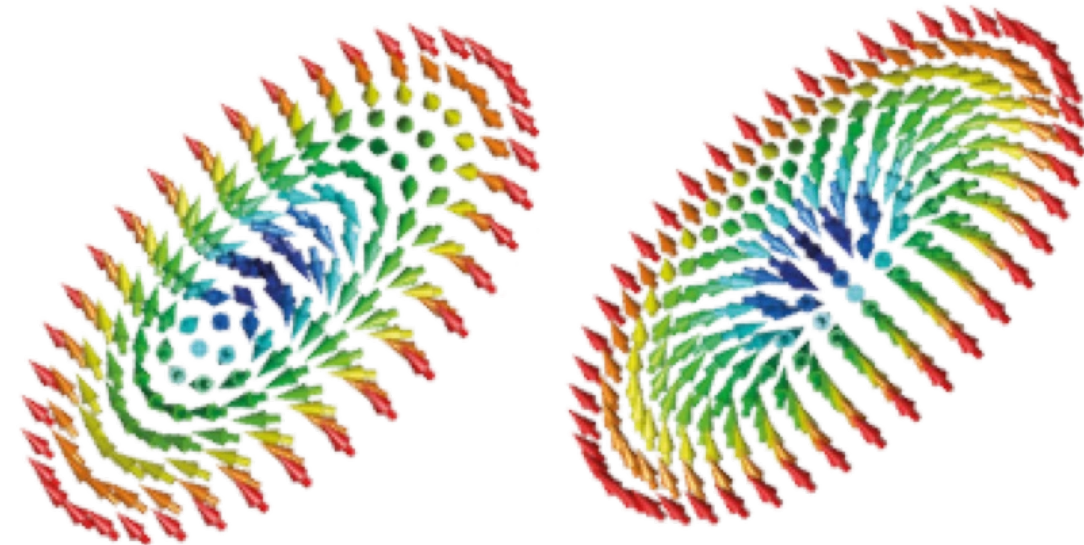
Beams of polarized neutrons (in which all the spins are aligned) allow the characterization of exotic materials with complex structure and behavior

- Copper
- Oxygen
- Carbon
- Hydrogen/deuterium
- Magnetic interactions

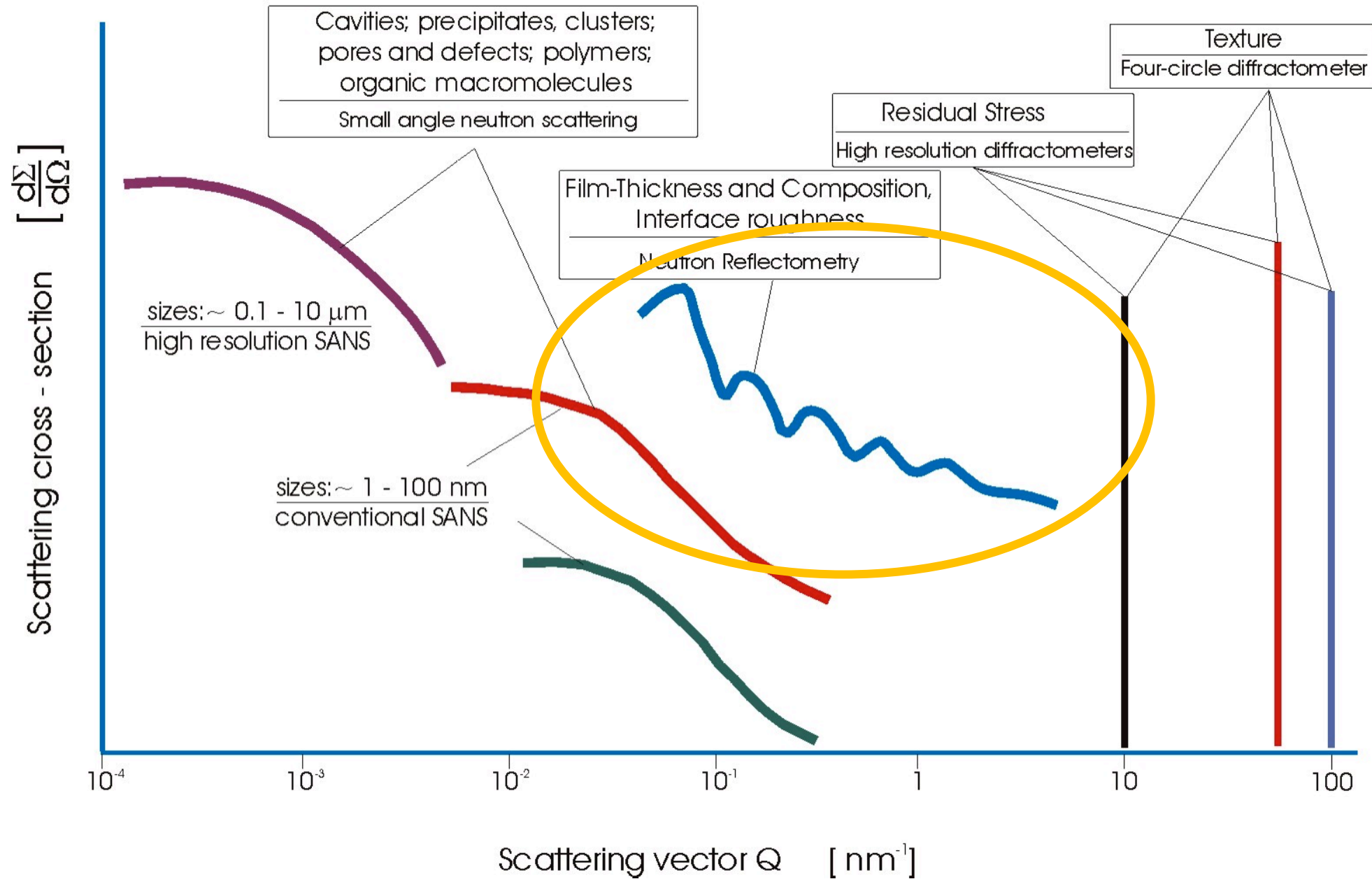


**Copper formate:  
crystal structure vs magnetic structure**

**Arrangement of spins in two different types of skyrmions**



# Schematic view of elastic neutron scattering spectra

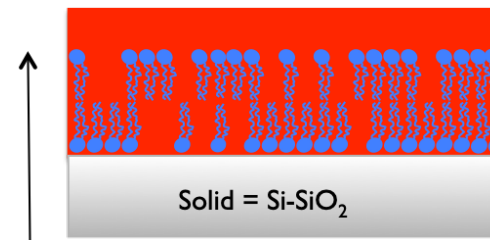
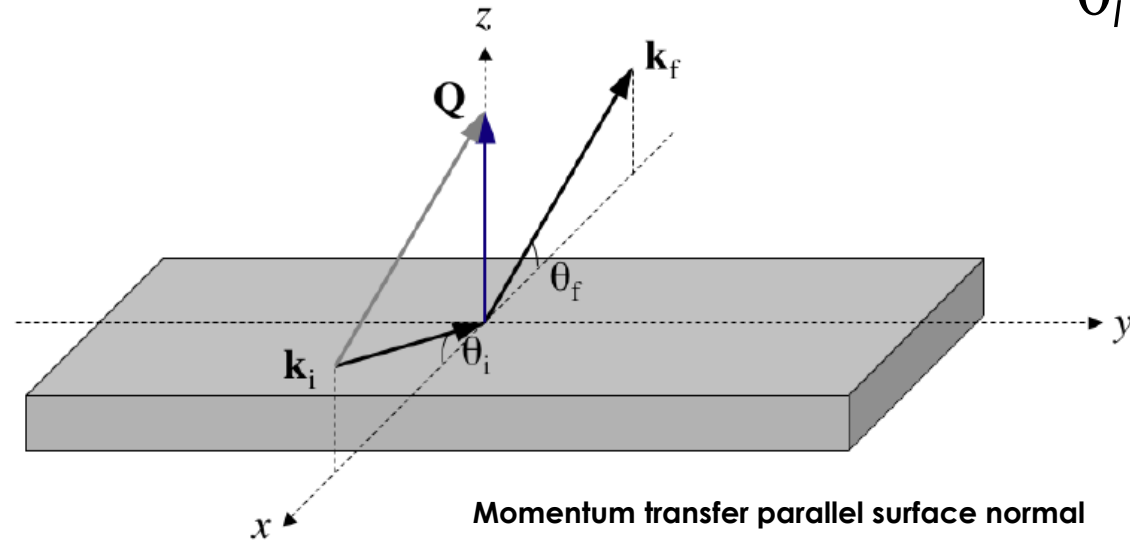
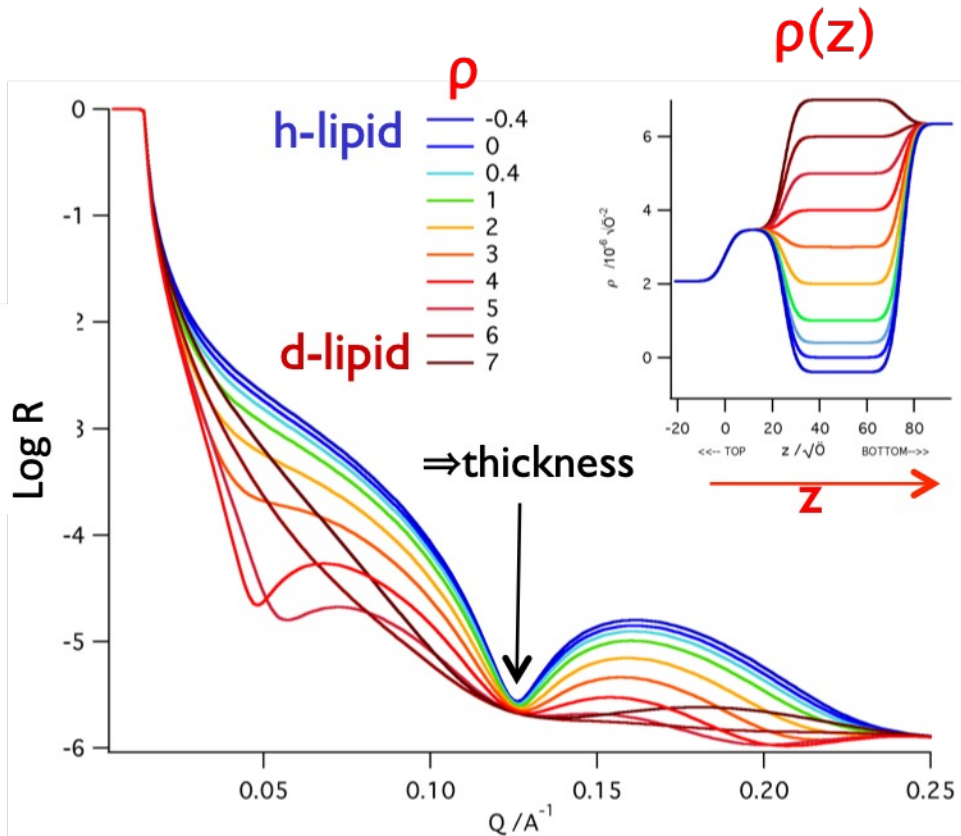




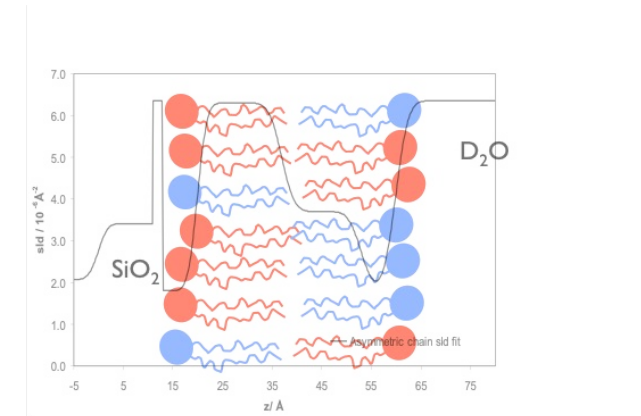
# Specular Reflectivity

- Thickness of layers at interfaces
- Roughness/interdiffusion
- Composition in the direction normal to the interface

$$\theta_i = \theta_f$$



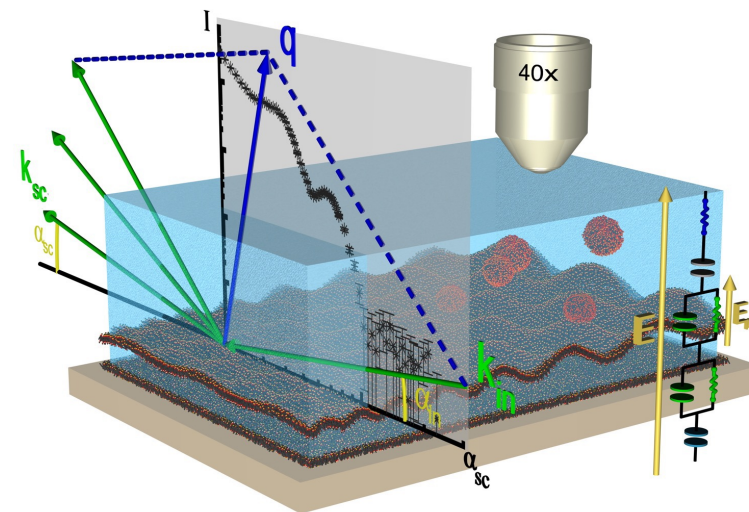
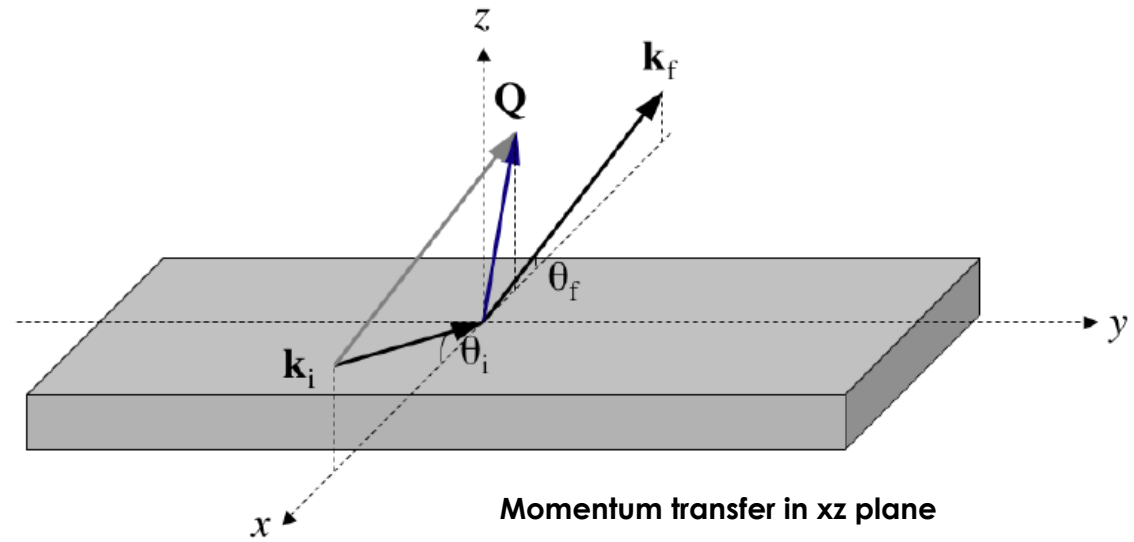
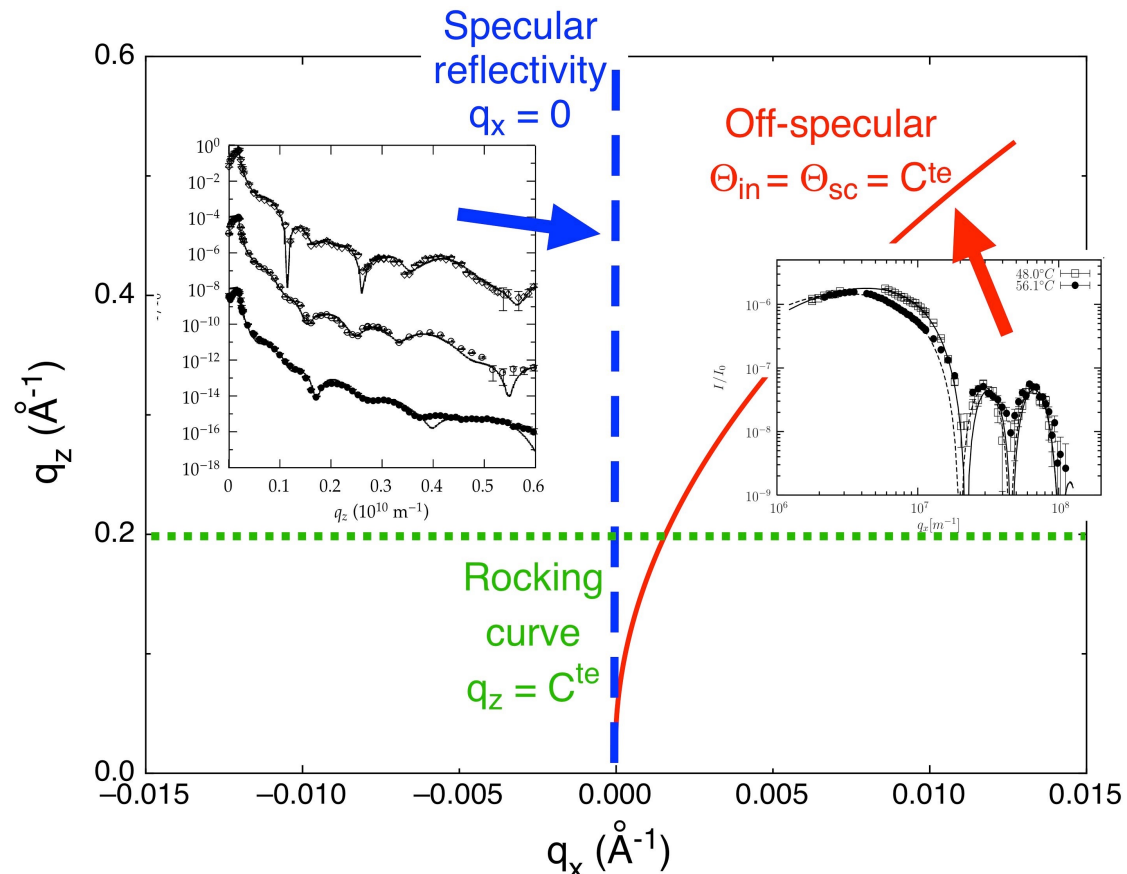
Model structure of lipid membrane  
= layered model of structure & composition



Scattering Length Density Profile

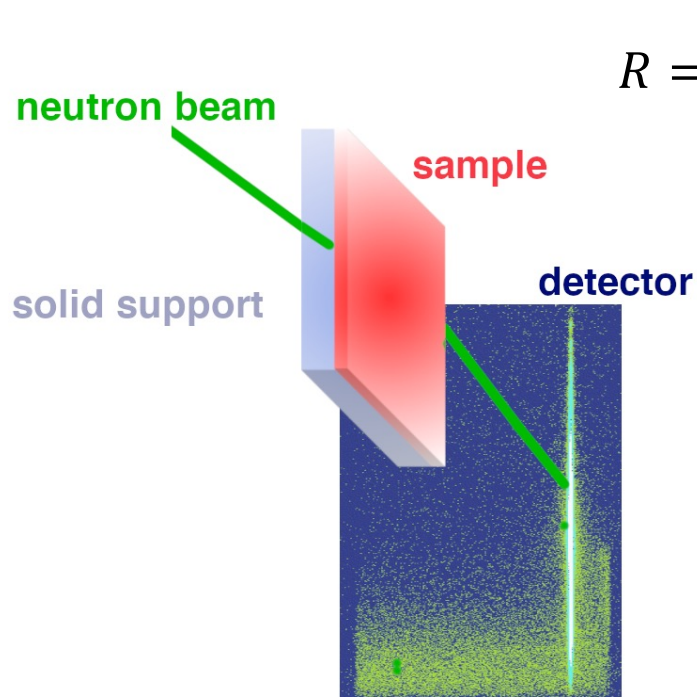
# Off-specular Reflectivity

In-plane features (height fluctuations, domains, holes ...) can be probed by **off-specular** measurements: for thin films synchrotron radiation is more suitable

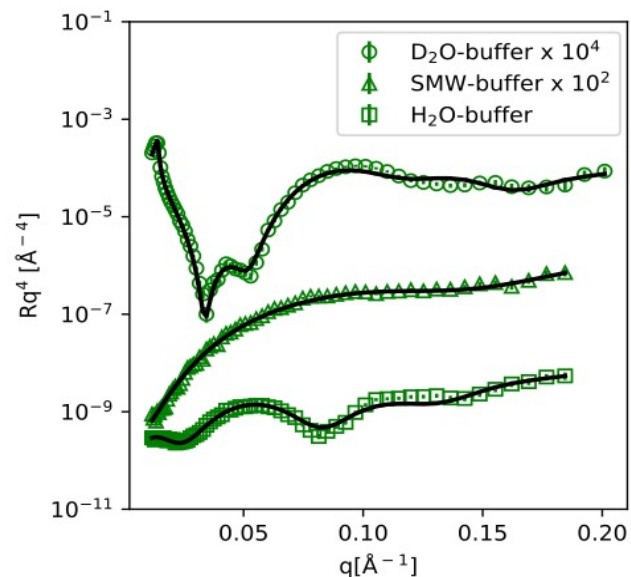




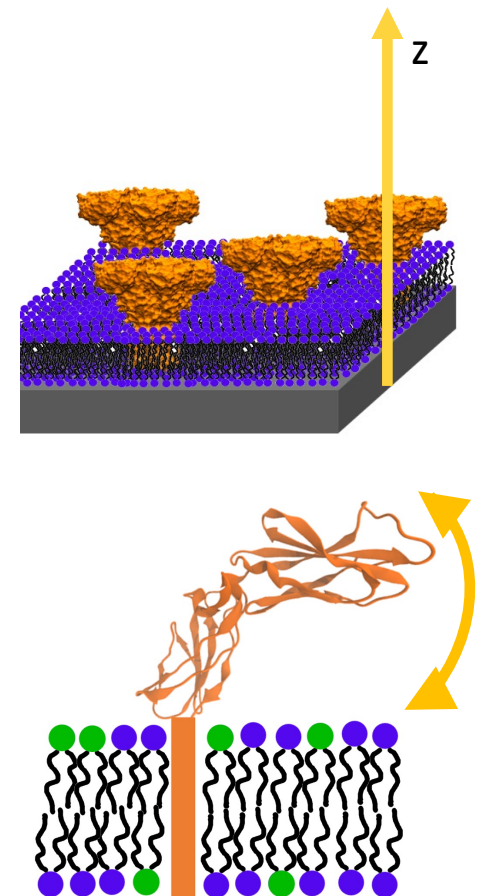
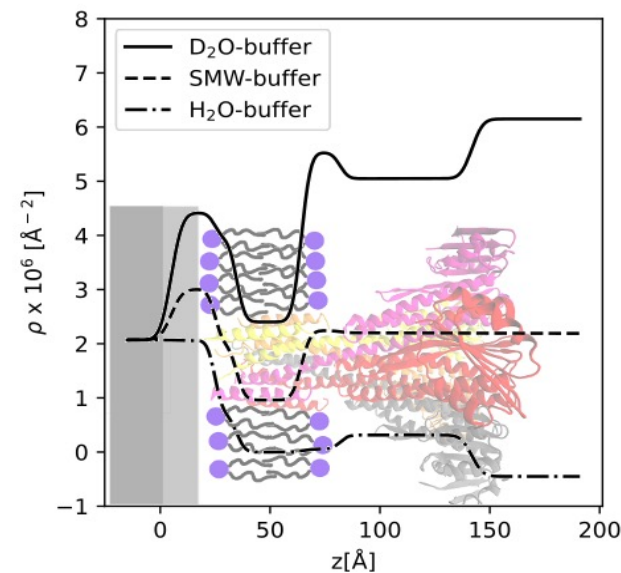
# Specular reflectivity measurements



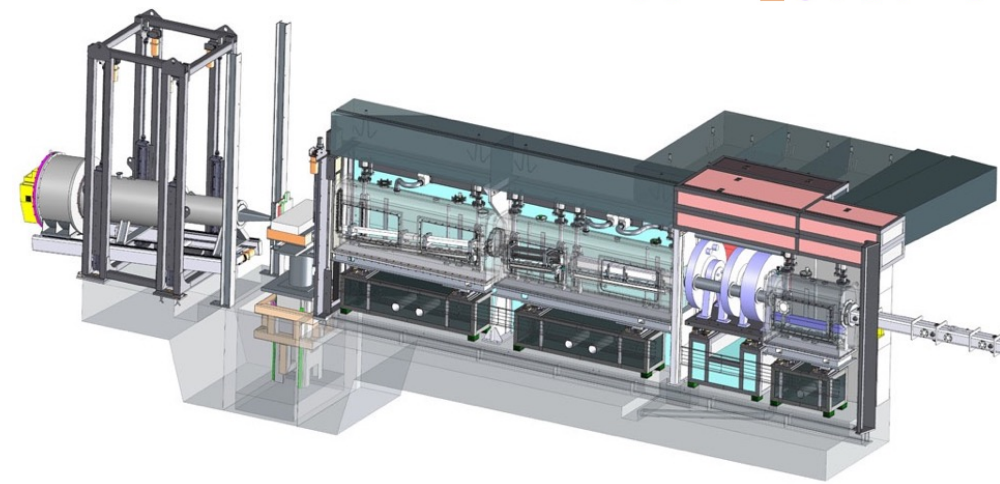
$$R = \frac{I_r}{I_0}$$



$$q = \frac{4\pi}{\lambda} \sin\theta$$



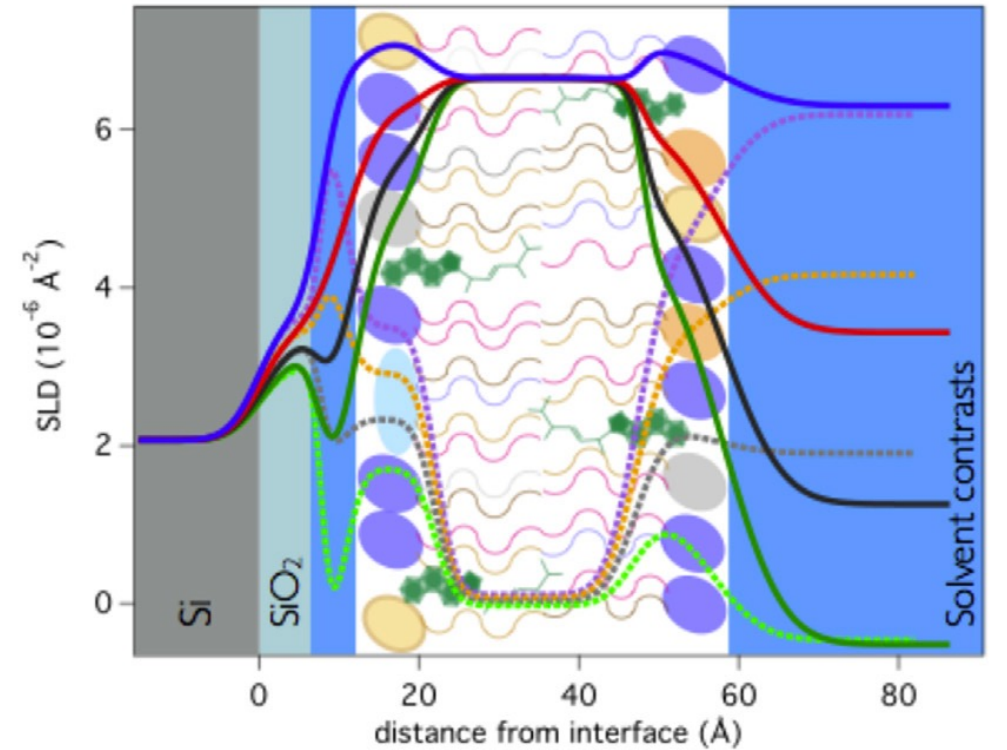
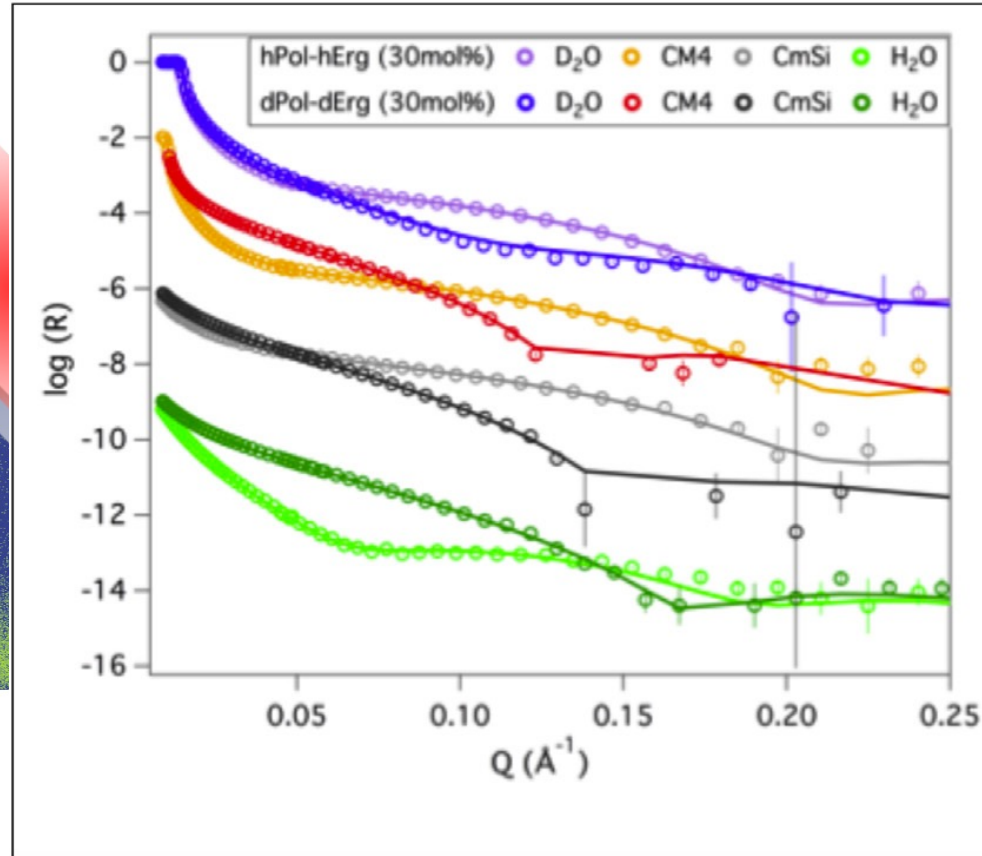
- Analysis done by model fitting
- Isotopic substitution and multiple contrast measurements used to improve resolution



# Specular reflectivity measurements

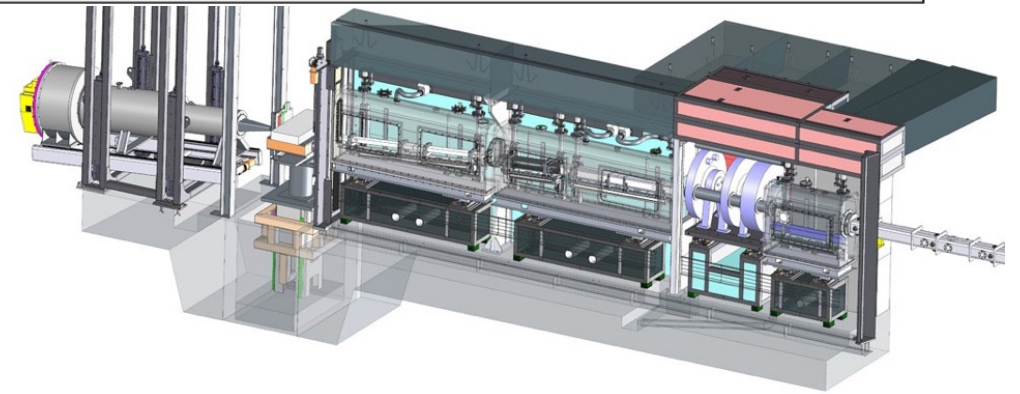
neutron beam

solid support



Current Opinion in Colloid & Interface Science

- Isotopic substitution and multiple contrast measurements used to improve resolution





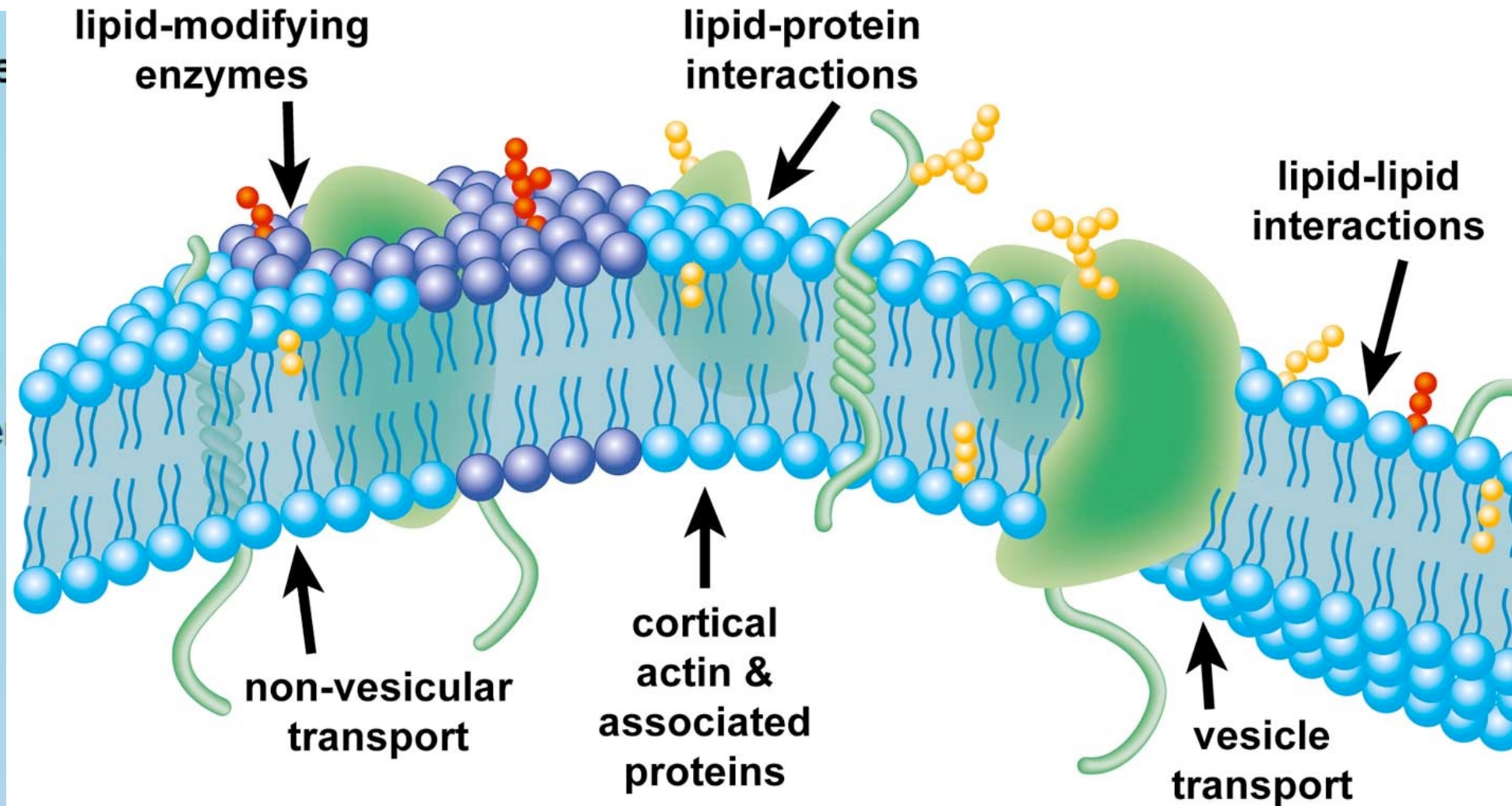
# Mechanisms at cell membranes

Complex assembly  
of lipids & proteins

Exhibit short range  
but not long range  
order

High degree of lateral  
heterogeneity

Transversely  
asymmetric

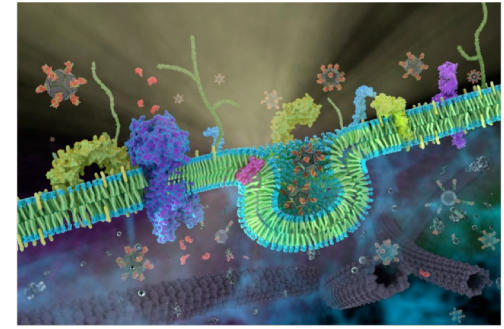


Lipid scaffold composed by a large variety of lipid species and levels of chain unsaturation, often difficult to synthesise chemically. Because of the complexity model membrane systems are used for fundamental studies.



developing model membrane systems since 1997...

**Total surface of membranes covers  
an area of 30,000 m<sup>2</sup> in our body**



**Function of membrane proteins** : dependent on membrane composition, lipid-protein interaction, lipid mediated protein-protein interaction

**Pharmacological interest** : Drug transport through membranes (dependent on physico-chemical membrane properties), anti-microbial peptides

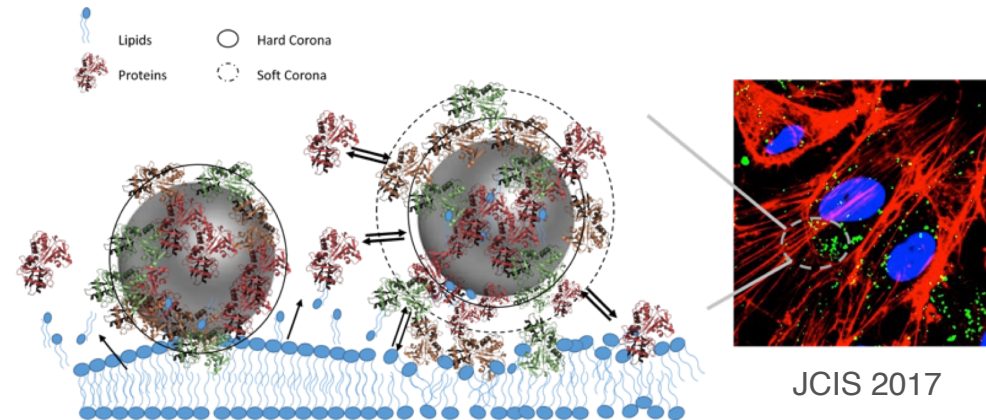
Membranes may play a *direct* role in **signal transduction**

**Diseases** associated with changes in lipid composition (diabetes, schizophrenia, Tay-Sachs syndrome, Alzheimer, Parkinson)

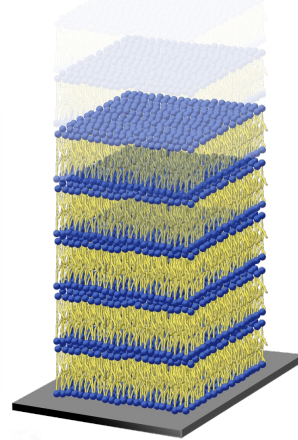
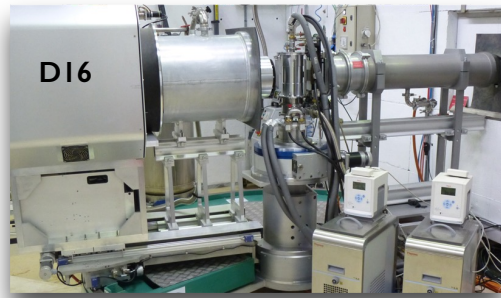
**Cell adhesion**

**Nano-biotechnology applications** (biosensors, bio-coatings)

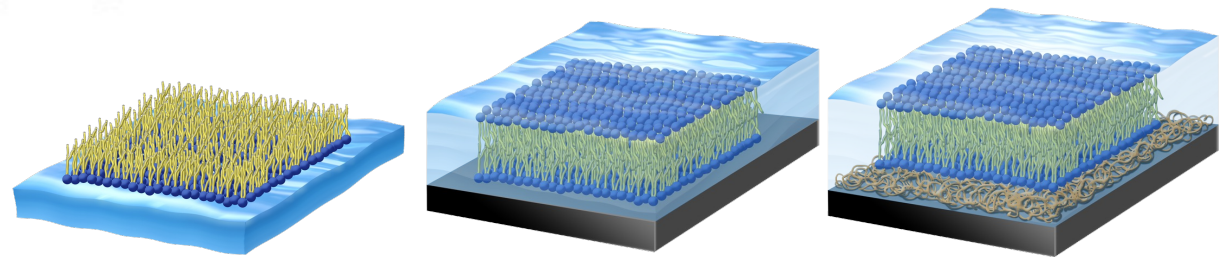
and .... **fascinating chemistry and physics!**



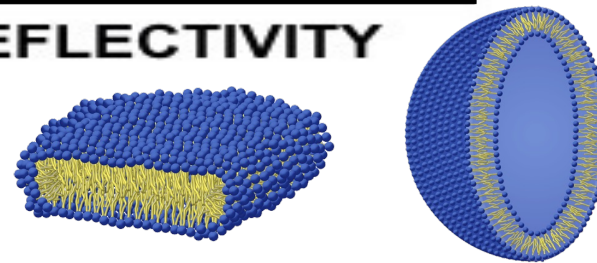
# Model membranes and scattering techniques



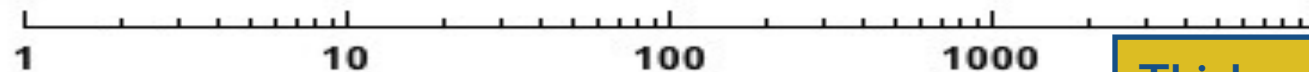
DIFFRACTION



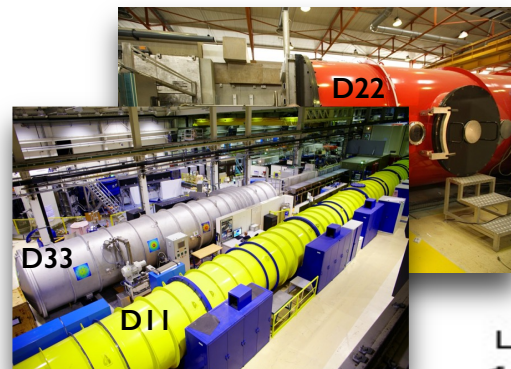
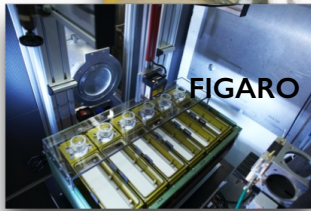
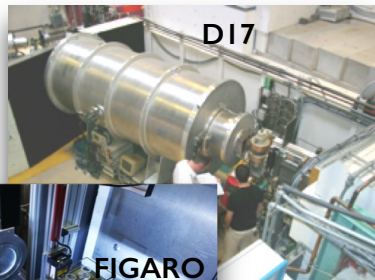
REFLECTIVITY



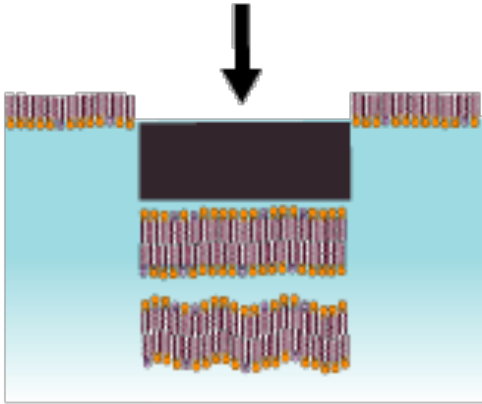
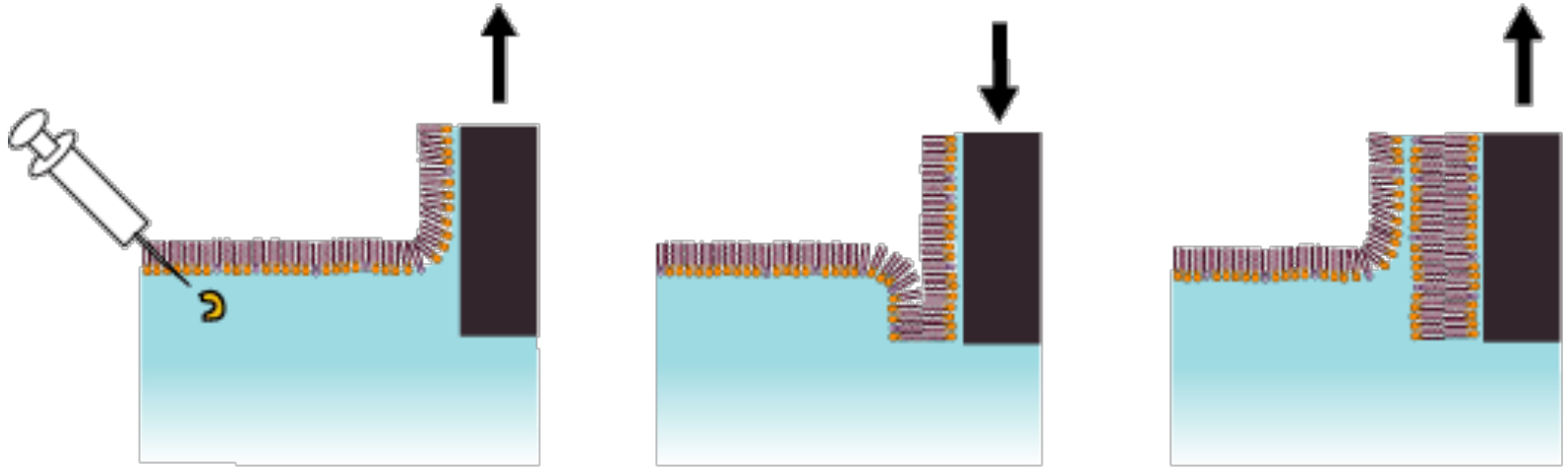
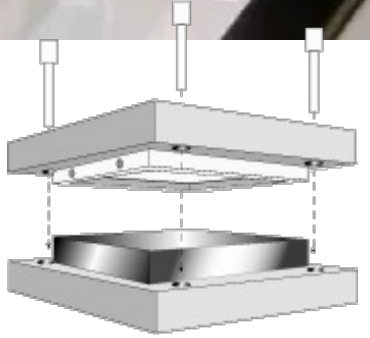
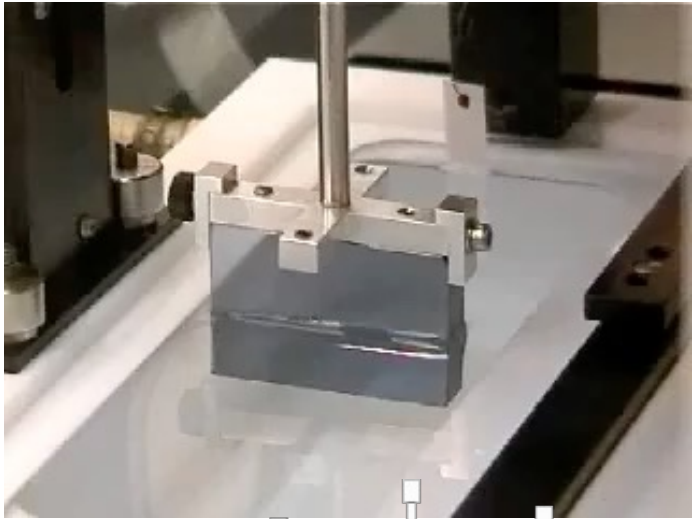
SANS



Thickness Å



# Floating bilayers prepared by Langmuir-Blodgett Langmuir-Schaefer techniques



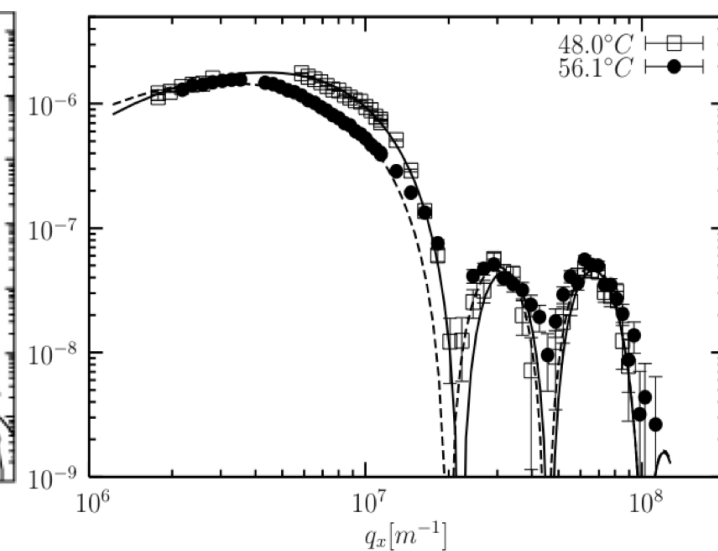
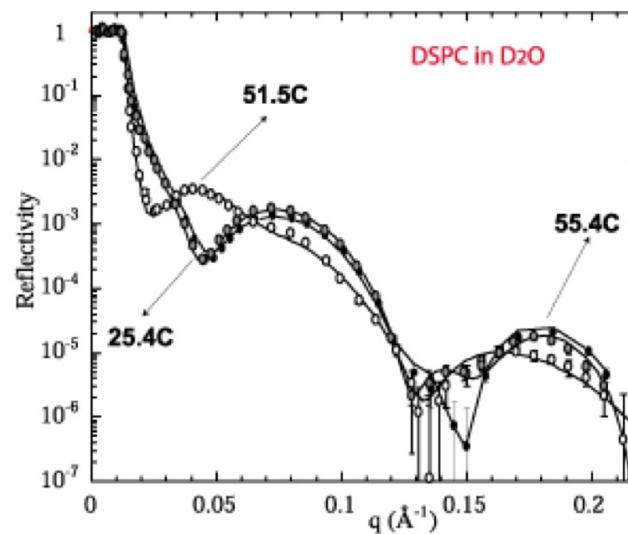
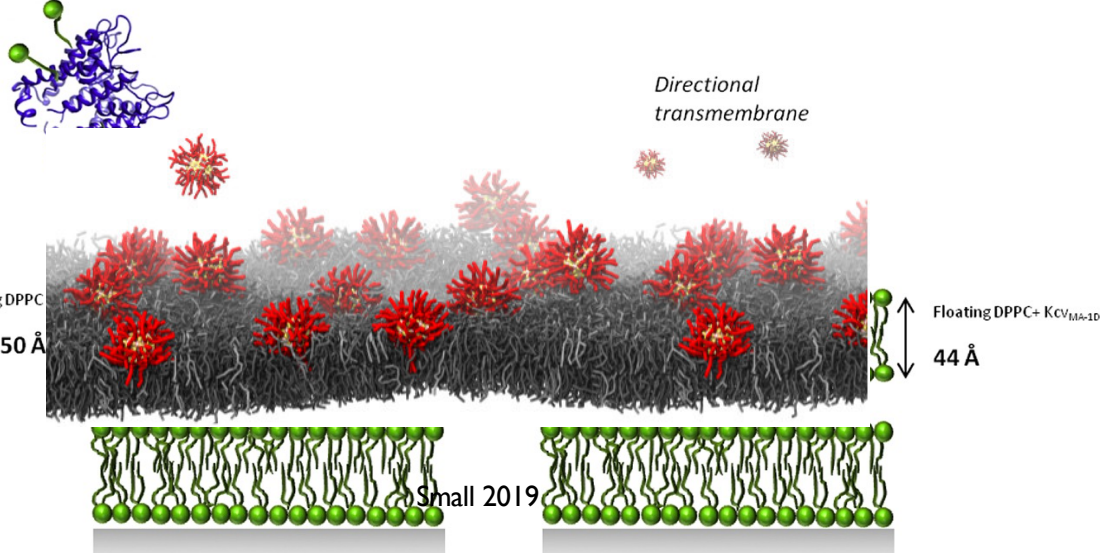
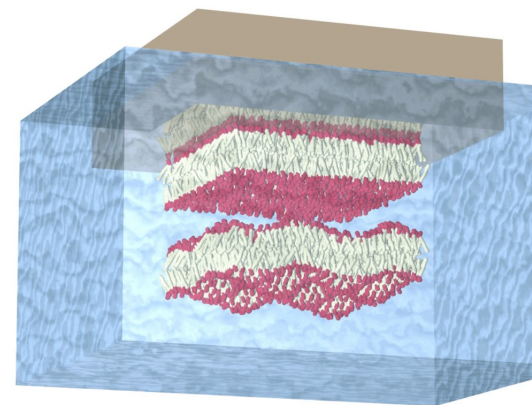
EPJB 1999



# 25 years of neutron and synchrotron radiation studies of structure and fluctuations of floating bilayers

EPJB 1999  
 EPL 2001  
 Langmuir 2001  
 Langmuir 2003  
 Langmuir 2005  
 Langmuir 2005  
 PNAS 2005  
 EPJE 2006  
 Soft Matter 2007  
 Langmuir 2009  
 BBA 2012  
 PNAS 2012  
 Langmuir 2012  
 EPJE 2013  
 Soft Matter 2015  
 PRL 2016  
 BBA 2018  
 Small 2019  
 J Chem Phys Lett 2019  
 J. Coll. Int. Sci. 2021

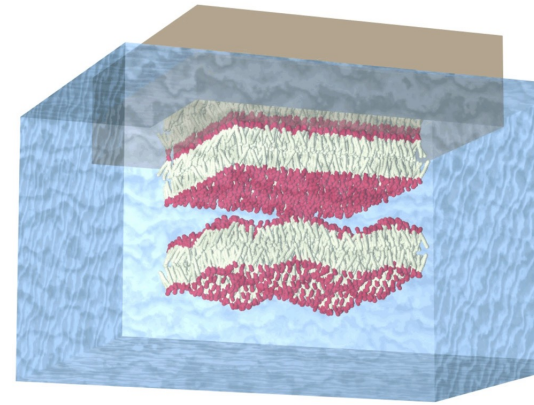
- Effect of temperature (giant swelling)
- Effect of charges
- Effect of AC current
- Interaction with gene delivery complexes
- Effect of domain forming molecules/asymmetry
- Lipid flip-flop
- Interaction with nano particles
- Transmembrane insertion



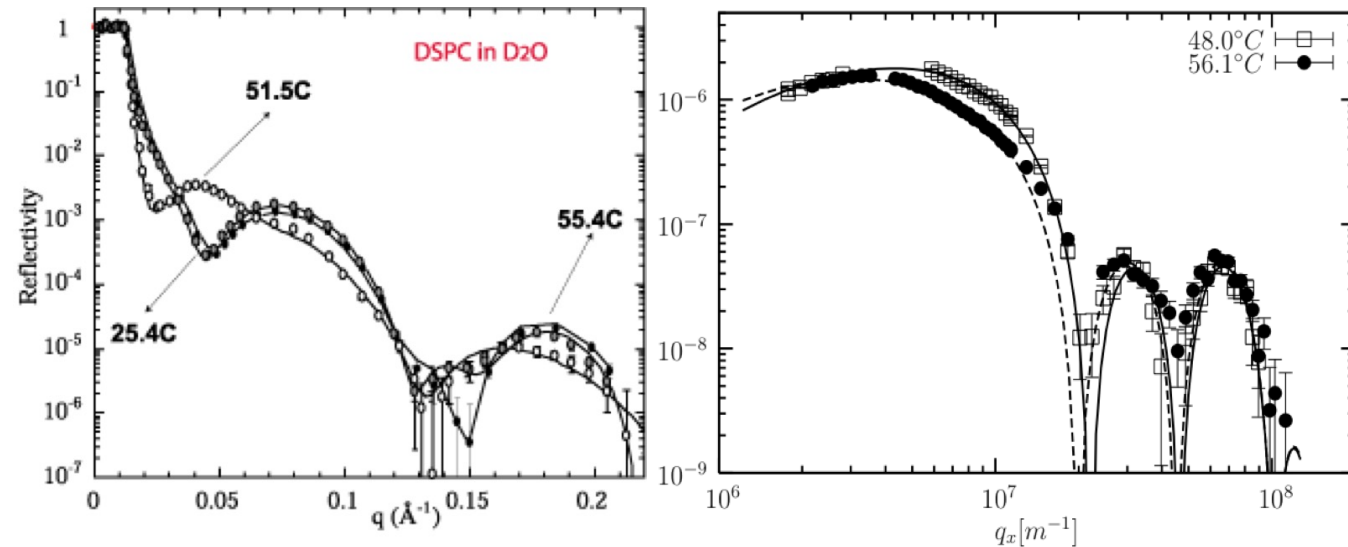
$$\langle z(q_{\parallel}) z(q_{\parallel}) \rangle = \frac{k_B T}{A + \gamma q_{\parallel}^2 + \kappa q_{\parallel}^4}$$

# 25 years of neutron and synchrotron radiation studies of structure and fluctuations of floating bilayers

- Effect of temperature (giant swelling)
- Effect of charges
- Effect of AC current
- Interaction with gene delivery complexes
- Effect of domain forming molecules/asymmetry
- Lipid flip-flop
- Interaction with nano particles
- **Transmembrane insertion and induced fluctuations**



EPJB 1999  
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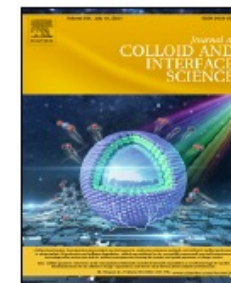


$$\langle z(q_{\parallel}) z(q_{\parallel}) \rangle = \frac{k_B T}{A + \gamma q_{\parallel}^2 + \kappa q_{\parallel}^4}$$



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# Journal of Colloid and Interface Science

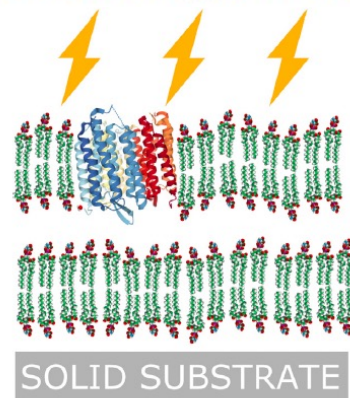
journal homepage: [www.elsevier.com/locate/jcis](http://www.elsevier.com/locate/jcis)

Regular Article

## Insertion and activation of functional Bacteriorhodopsin in a floating bilayer



Tetiana Mukhina<sup>a,b,1</sup>, Yuri Gerelli<sup>a,c,\*</sup>, Arnaud Hemmerle<sup>d</sup>, Alexandros Koutsioubas<sup>e</sup>, Kirill Kovalev<sup>f,g,e,h,i,j</sup>, Jean-Marie Teulon<sup>f</sup>, Jean-Luc Pellequer<sup>f</sup>, Jean Daillant<sup>d</sup>, Thierry Charitat<sup>b,\*</sup>, Giovanna Fragneto<sup>a,\*</sup>



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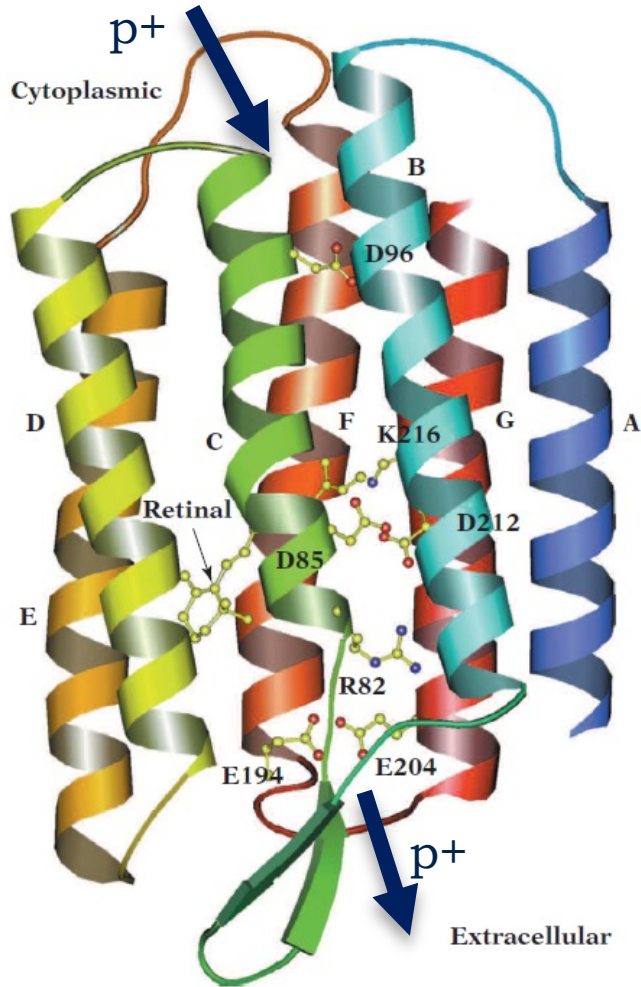
## ABSTRACT

The proton pump transmembrane protein bacteriorhodopsin was successfully incorporated into planar floating lipid bilayers in gel and fluid phases, by applying a detergent-mediated incorporation method. The method was optimized on single supported bilayers by using quartz crystal microbalance, atomic force and fluorescence microscopy techniques. Neutron and X-ray reflectometry were used on both single and floating bilayers with the aim of determining the structure and composition of this membrane-



# Bacteriorhodopsin (BR)

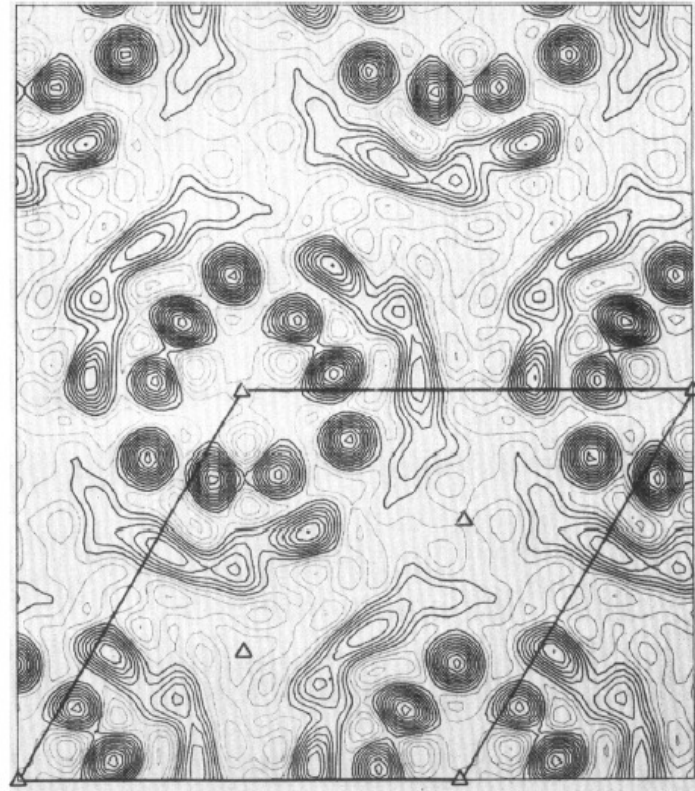
High-resolution structure of BR



Pebay-Peyroula, E., et al., *Biochim Biophys Acta*. 2000, 1460:119–132.

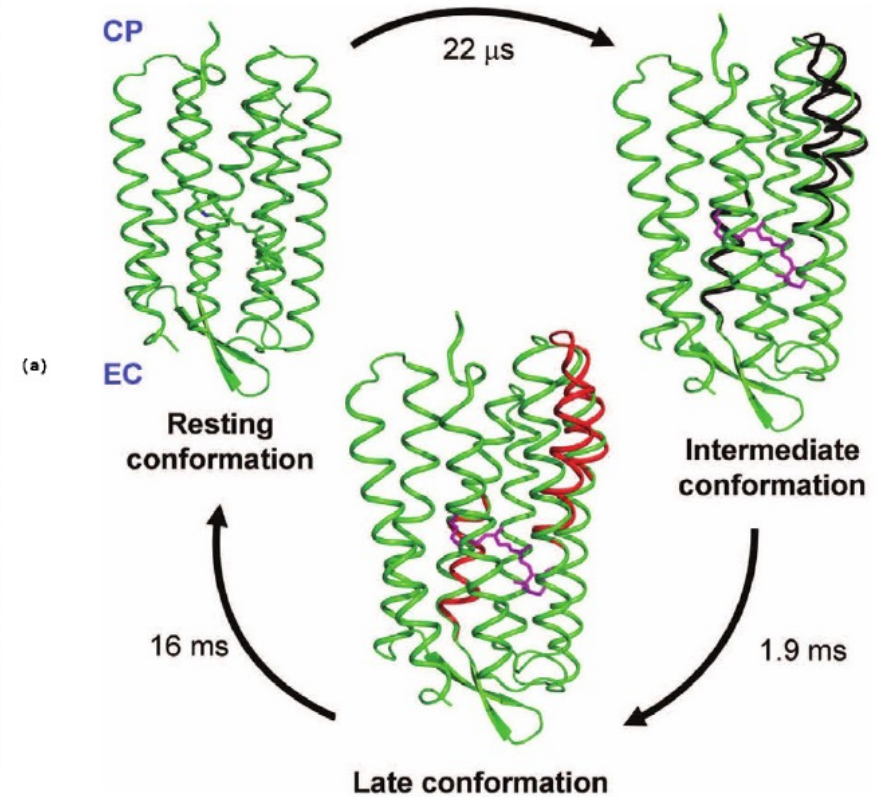
26~kDa transmembrane protein that acts as a light-driven proton pump in *Halobacterium salinarum*, converting light energy into a proton gradient.

Electron density profile of the 2D crystalline purple membrane



Unwin P. N. T. and R. Henderson *J. Mol. Biol.*, 1976, 94:425 - 440

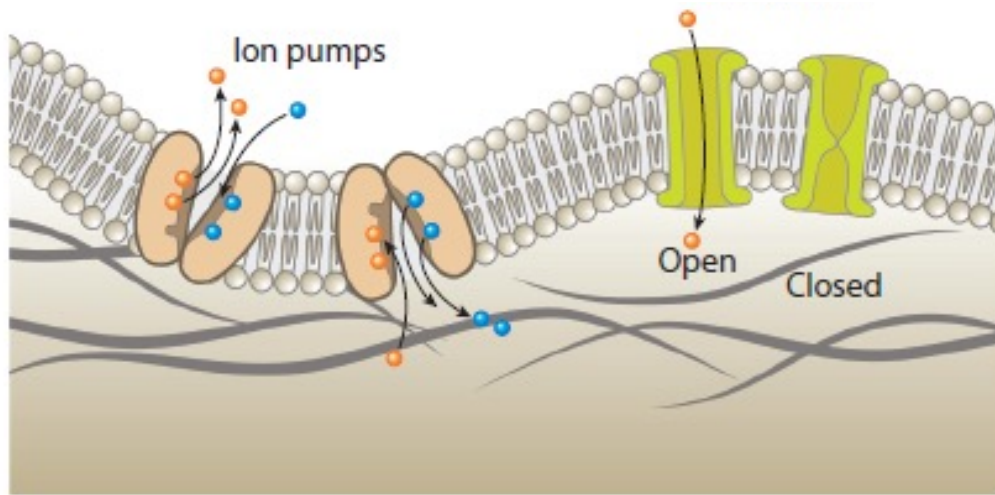
Conformational changes of BR during the photocycle



Andersson, M., et al., *Structure*. 2009, 17:1265–1275.

# Membrane Fluctuations

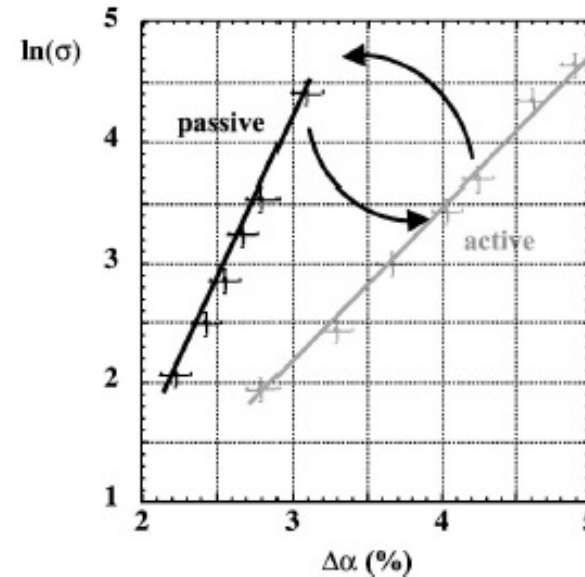
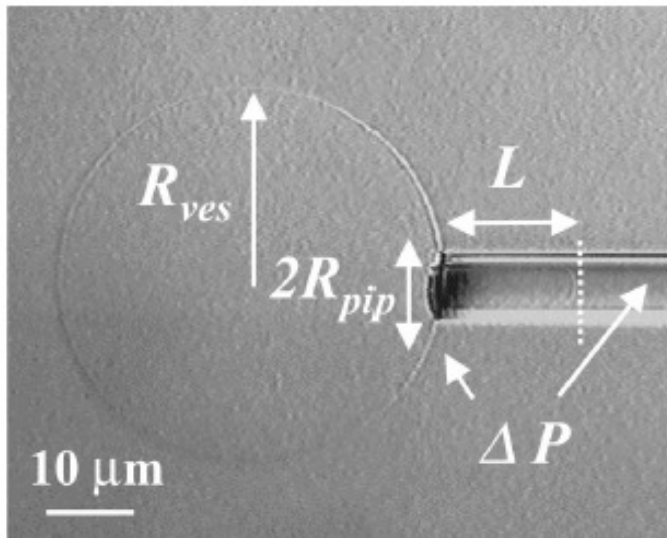
- Thermal fluctuations
- Active fluctuations - out of equilibrium system



Turlier H. Annu. Rev. Condens. Matter Phys. 2019. 10:213–32

19

**Micropipette experiments**  
**Videomicroscopy experiments**



$$\Delta\alpha = \alpha_0 - \alpha = \frac{k_B T_{eff}}{8\pi\kappa} \ln\left(\frac{\sigma}{\sigma_0}\right)$$

$$1.7 \leq \frac{T_{eff}}{T} \leq 2.7$$

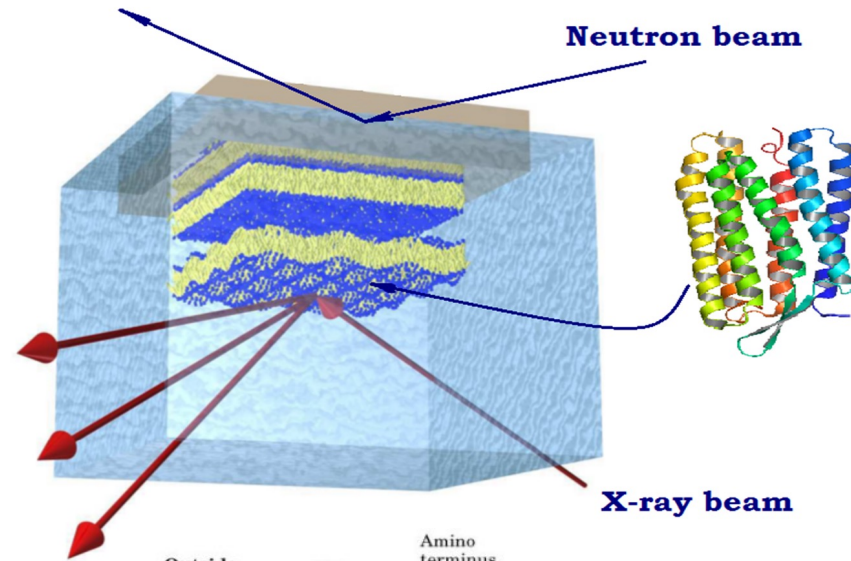
A larger excess area could be pulled out by micropipette aspiration when BR was active, indicating that its **proton pumping activity induces an amplification of the membrane shape fluctuations** and a strong decrease in the effective bending modulus of the membrane. **Information only at the micrometer scale.**



# Out-of-equilibrium fluctuations of phospholipid membranes induced by active transmembrane proteins

## *X-ray and synchrotron radiation Specular and Off- Specular*

- Structure with Å resolution
- Fluctuation spectrum
- Lateral inhomogeneities and defects
- Physical properties of the system

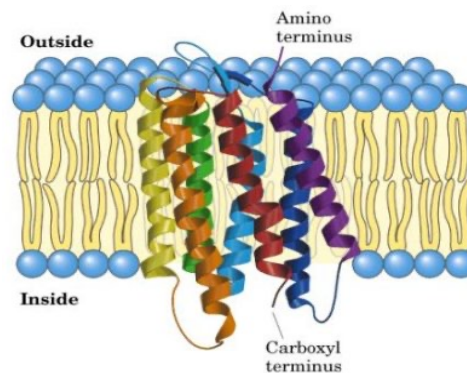


## *Neutron reflectometry*

- Structure with Å resolution
- Atomic composition
- Interface roughness
- Solvent content

## *Fluorescence microscopy*

- Lateral features of the system with  $\mu\text{m}$  resolution
- Sample fluorescence/bleaching



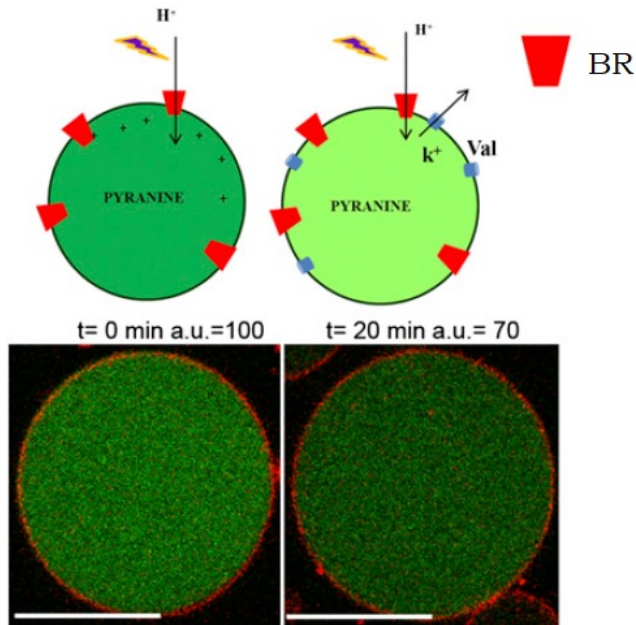
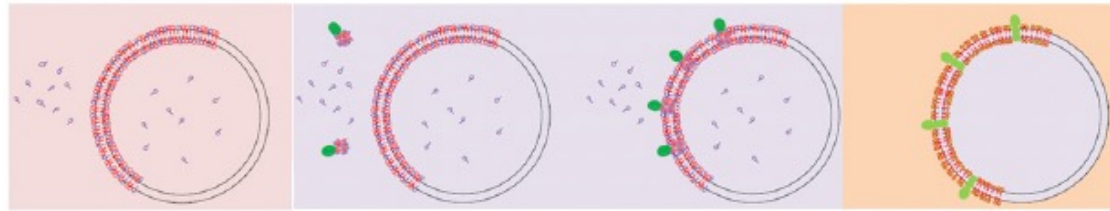
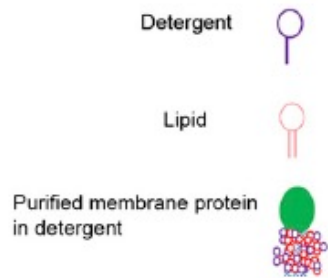
## *Atomic Force Microscopy AFM*

- Lateral and transversal features of the system with **nm** resolution
- Mechanical properties of the sample
- Force measurements



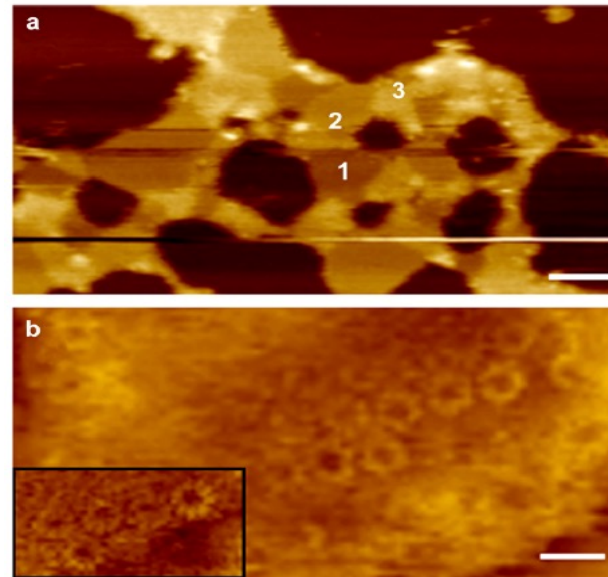


# Detergent-mediated protein incorporation



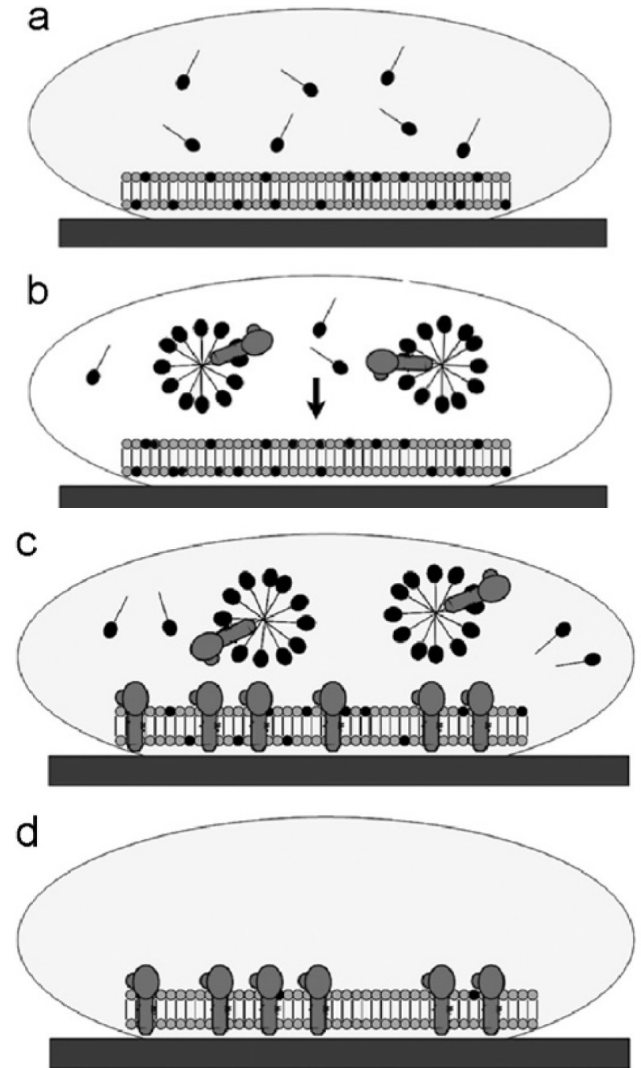
GUV with incorporated BR.

Dezi et al. PNAS , 2013, 110 (18) 7276-7281



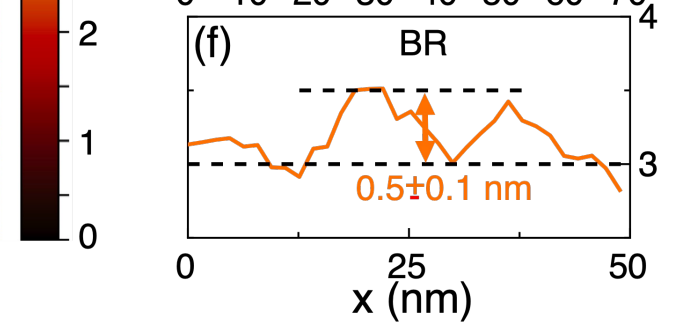
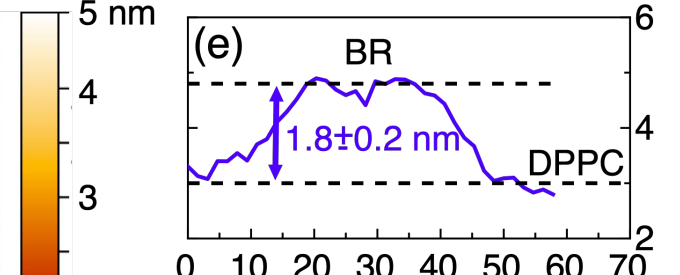
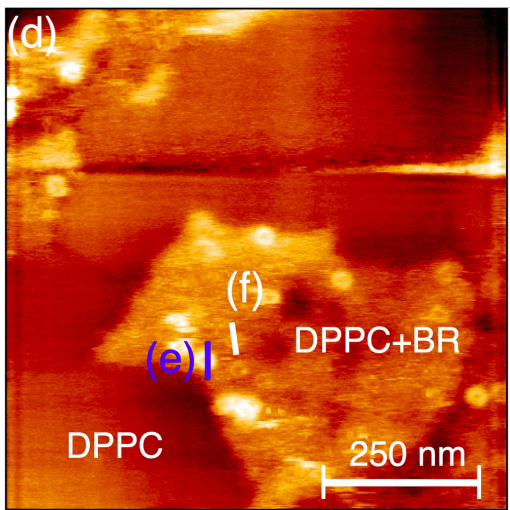
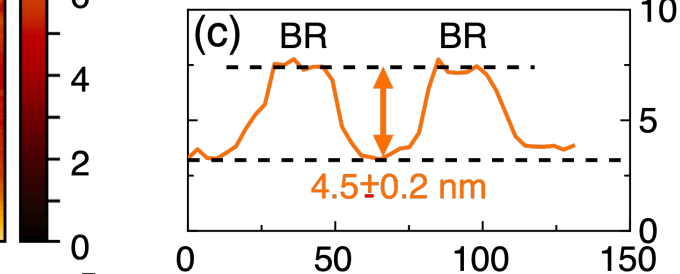
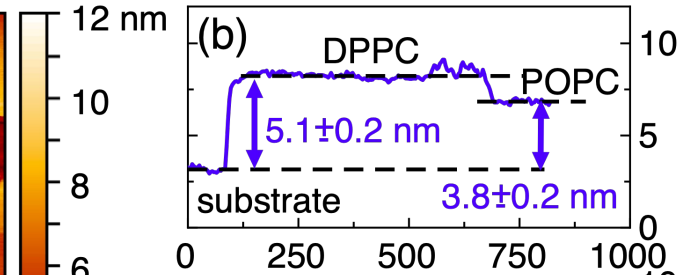
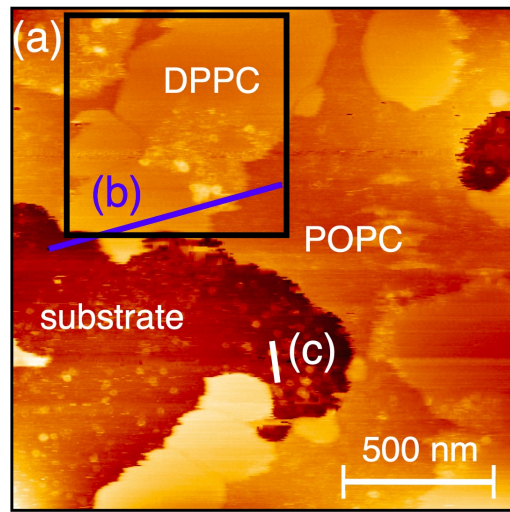
AFM imaging of LH2 protein incorporated into SLB.

Milhiet et al., Biophys. J. , 2016, 91(9) 3268–3275



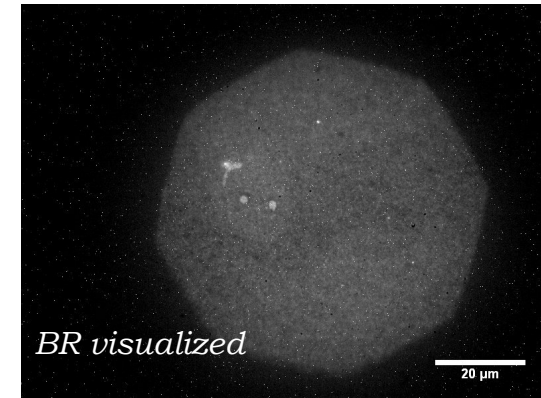
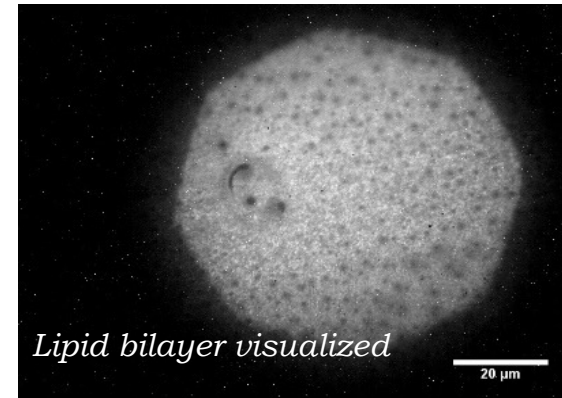
Berquand A. et al. Ultramicroscopy, 2007, 107(10-11), 928–933

Atomic force microscopy

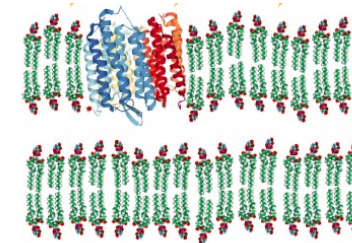
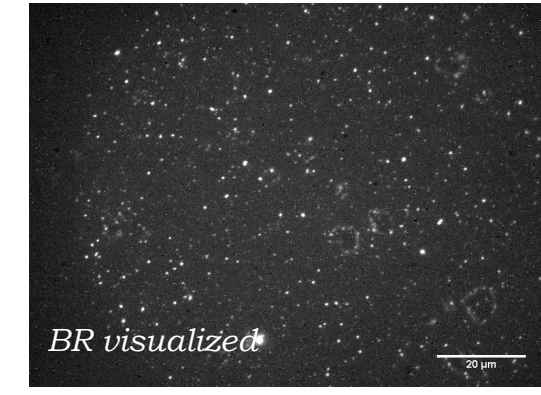
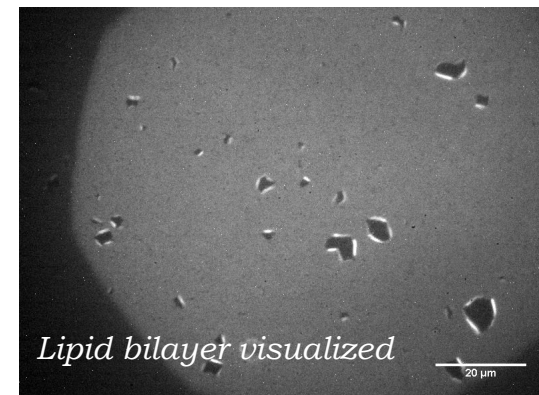


Phase separated gel-fluid  
single bilayer + BR,  
IBS, Grenoble

Fluorescence microscopy



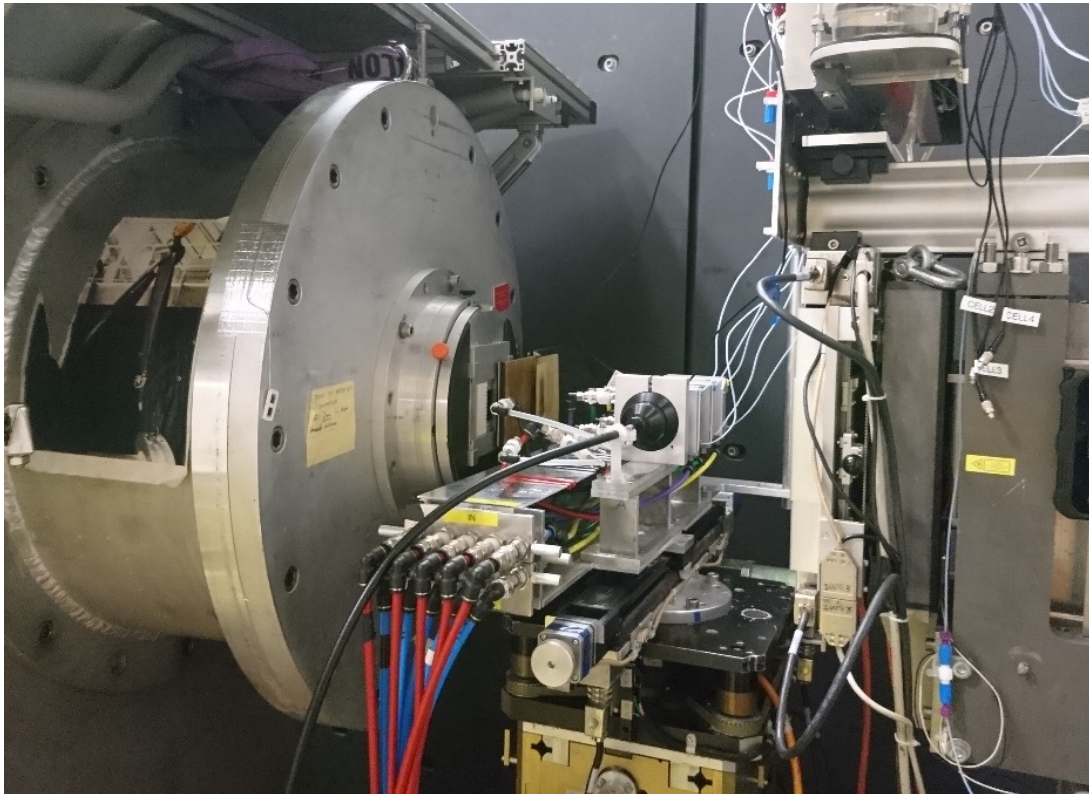
Gel single bilayer + BR



SOLID SUBSTRATE

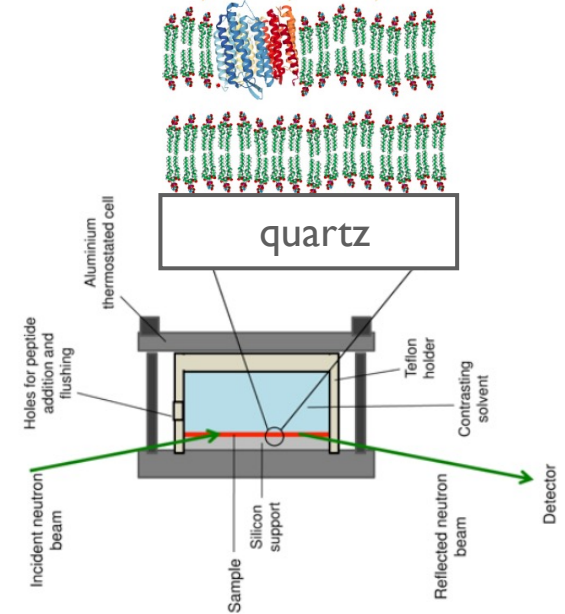
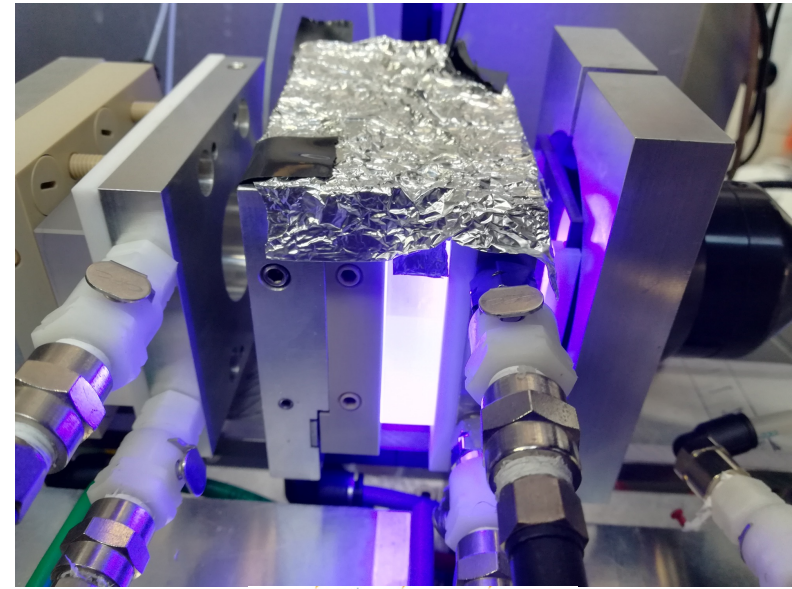
Gel double bilayer + BR,  
Institut Charles Sadron, Strasbourg





**D17 reflectometer ( ILL, Grenoble, France).**

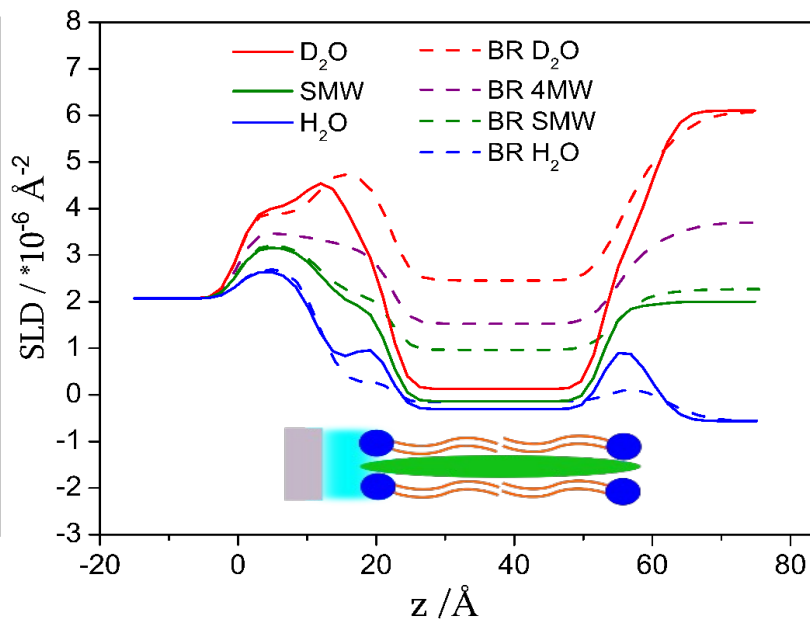
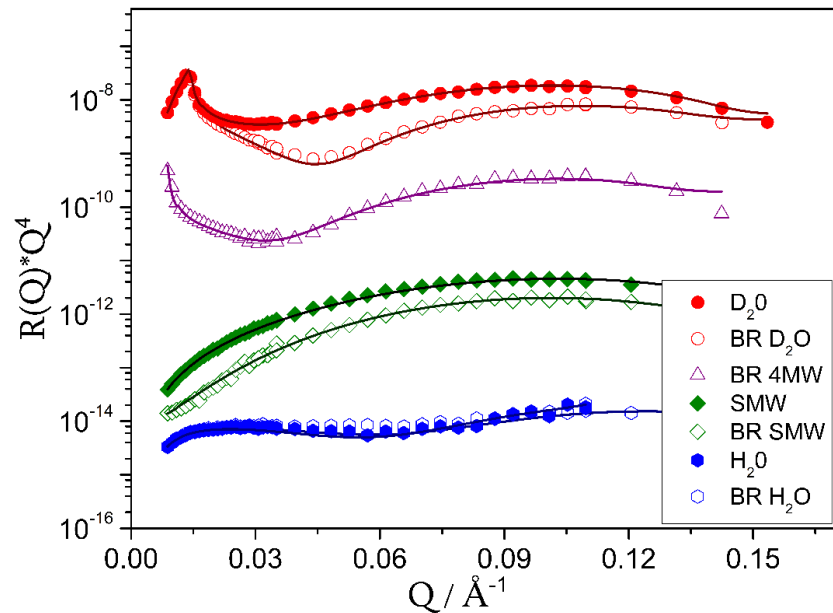
Time-of-flight mode





# Neutron Reflectometry

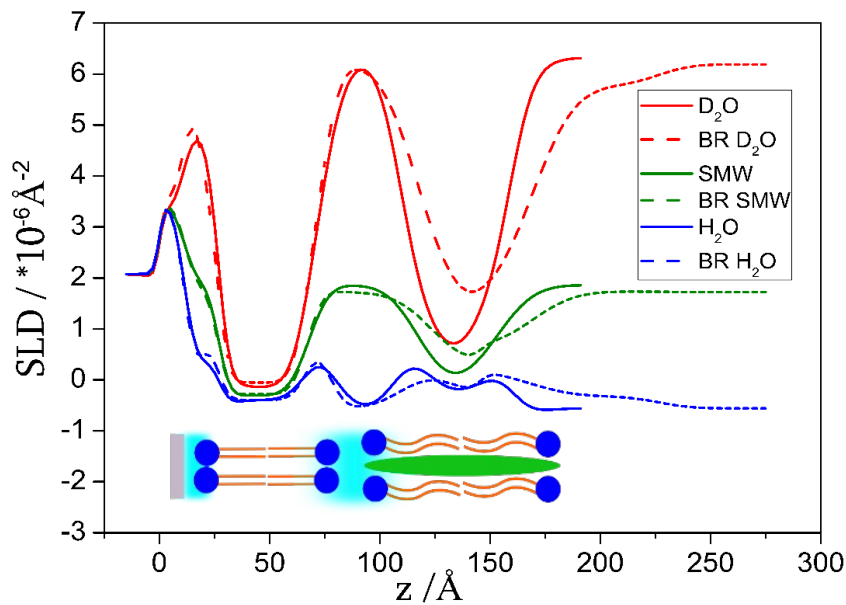
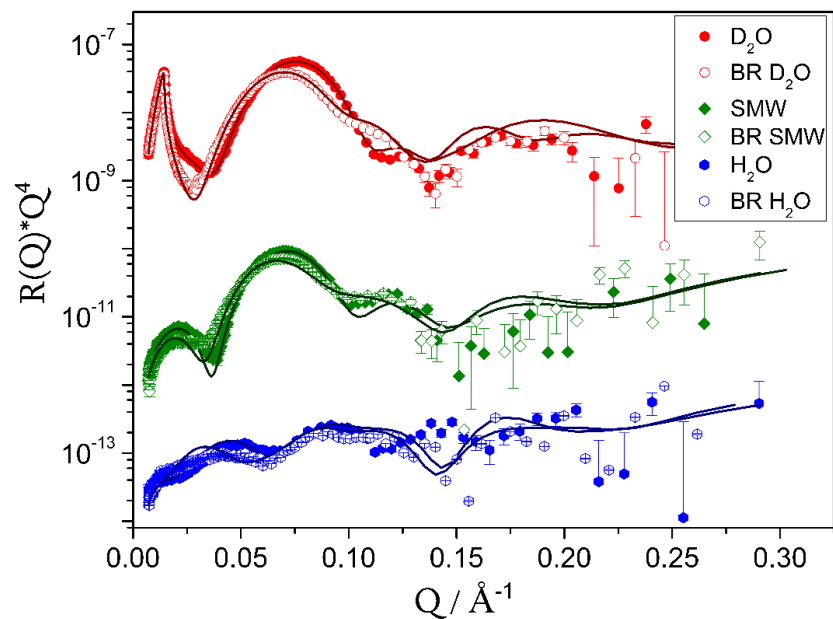
POPC



**Fluid single bilayer + BR**  
**MARIA reflectometer**  
*JCNS, Germany*

*18 % by volume of BR*

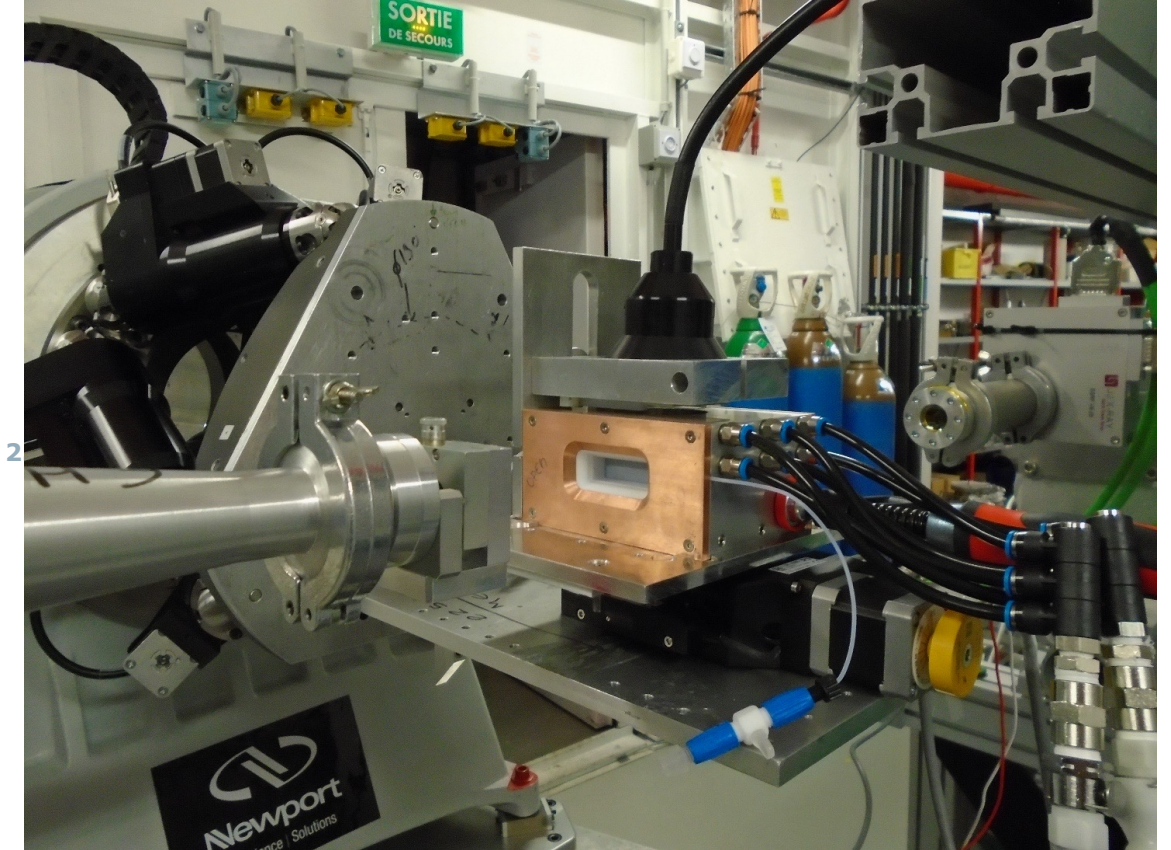
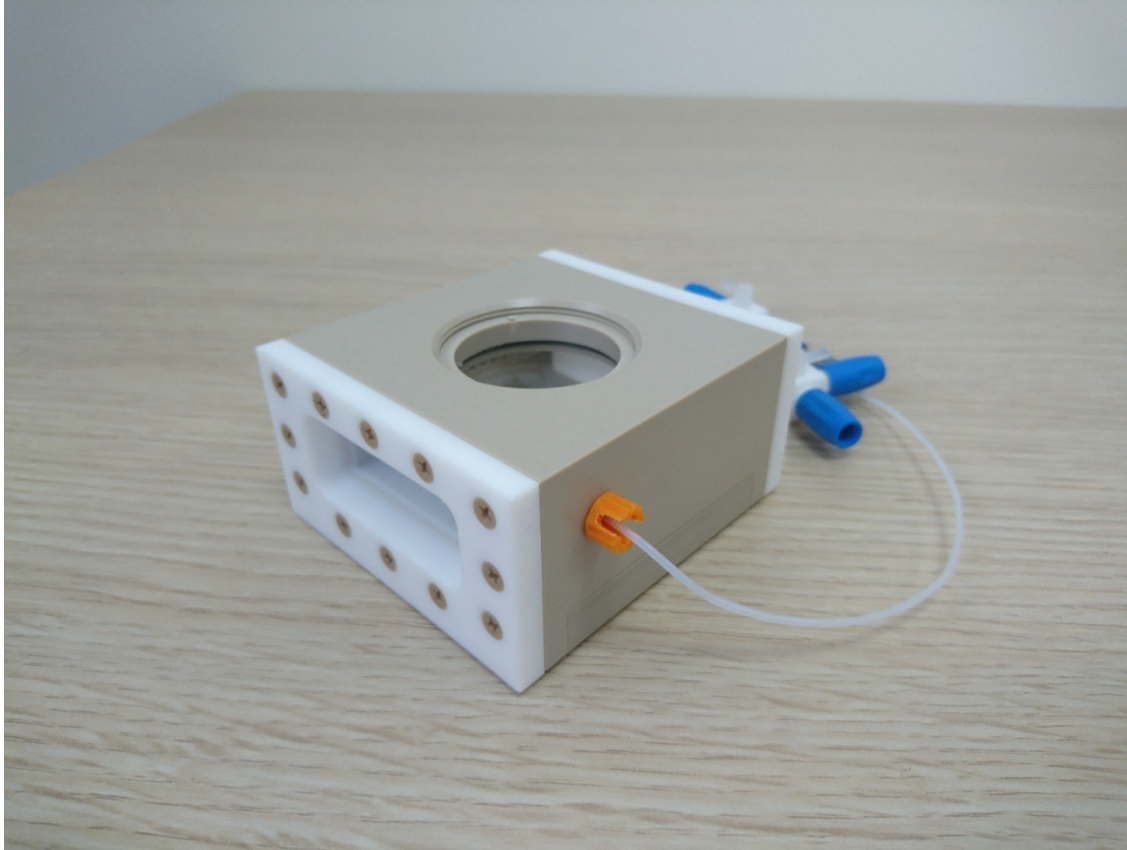
DP  
PC  
DS  
PC



**Fluid floating bilayer + BR**  
**D17 reflectometer**

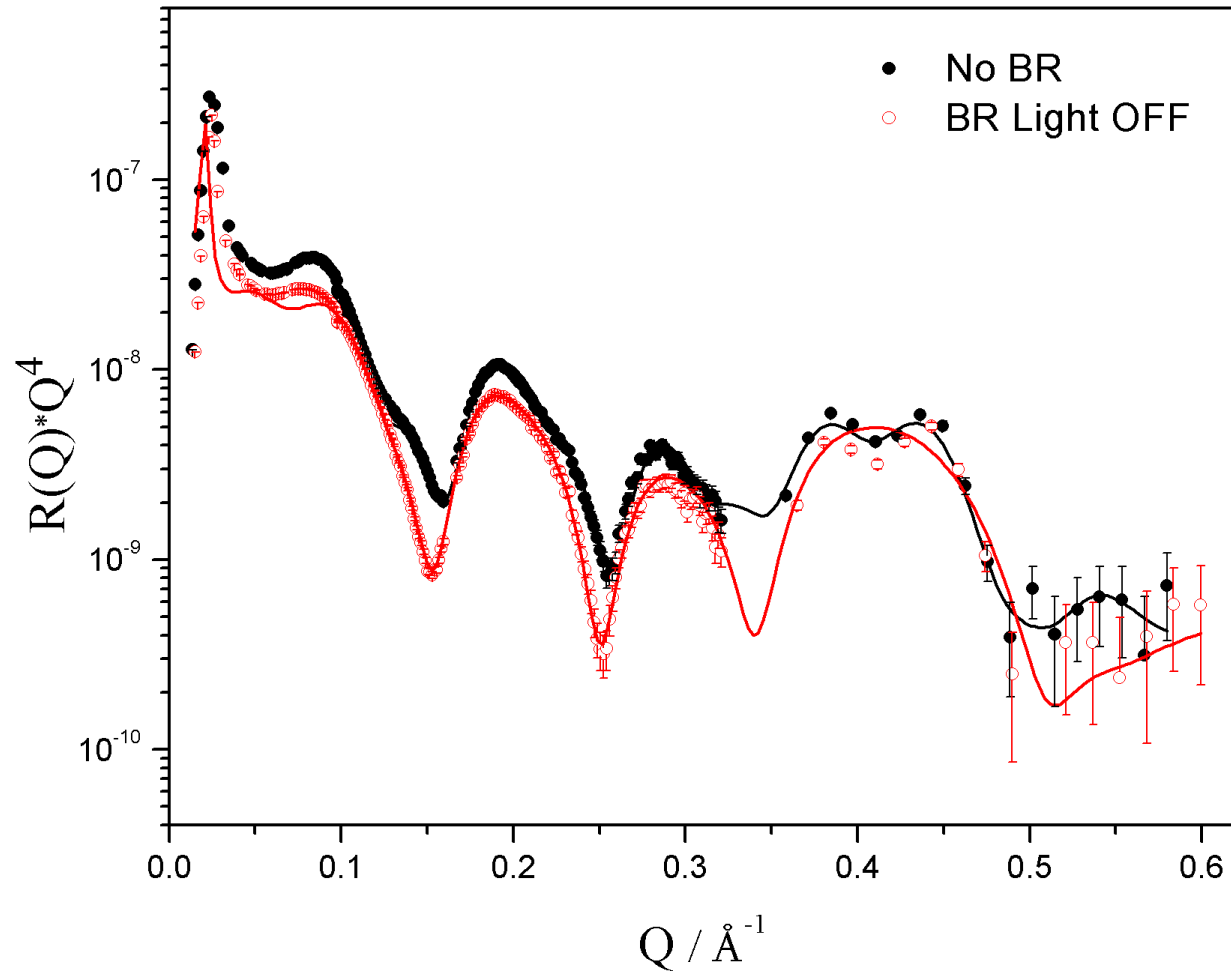
*BR insertion into the floating bilayer  
 + BR layer on top*

# Sample environment for X-ray reflectivity experiment



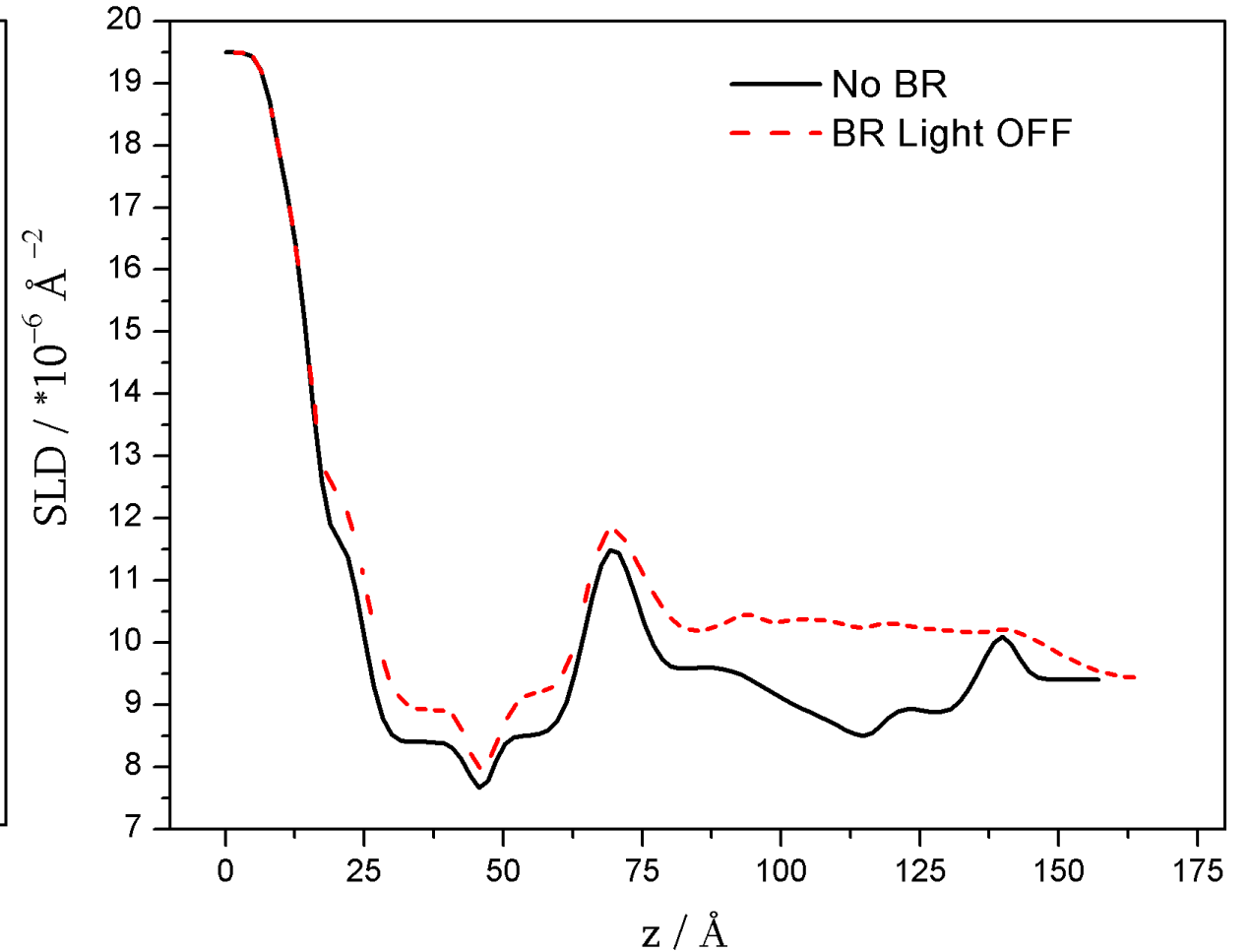
**SixS beamline**  
SOLEIL synchrotron, France

# Specular SR Reflectometry



*DSPC **double bilayer** at 25°C before and after protein BR incorporation.*

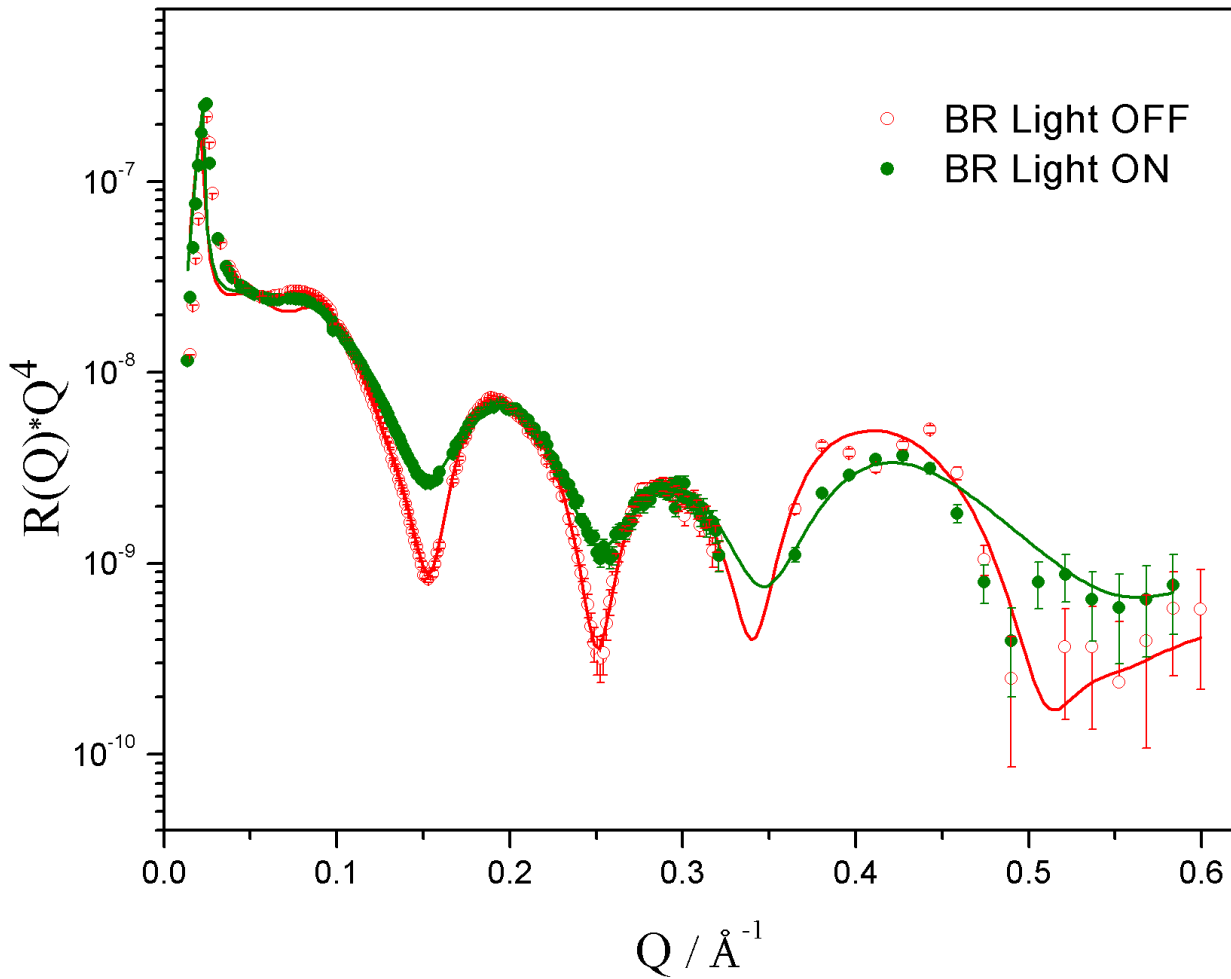
***SixS beamline (SOLEIL synchrotron, France).***



*SLD profiles corresponding to the fits.*

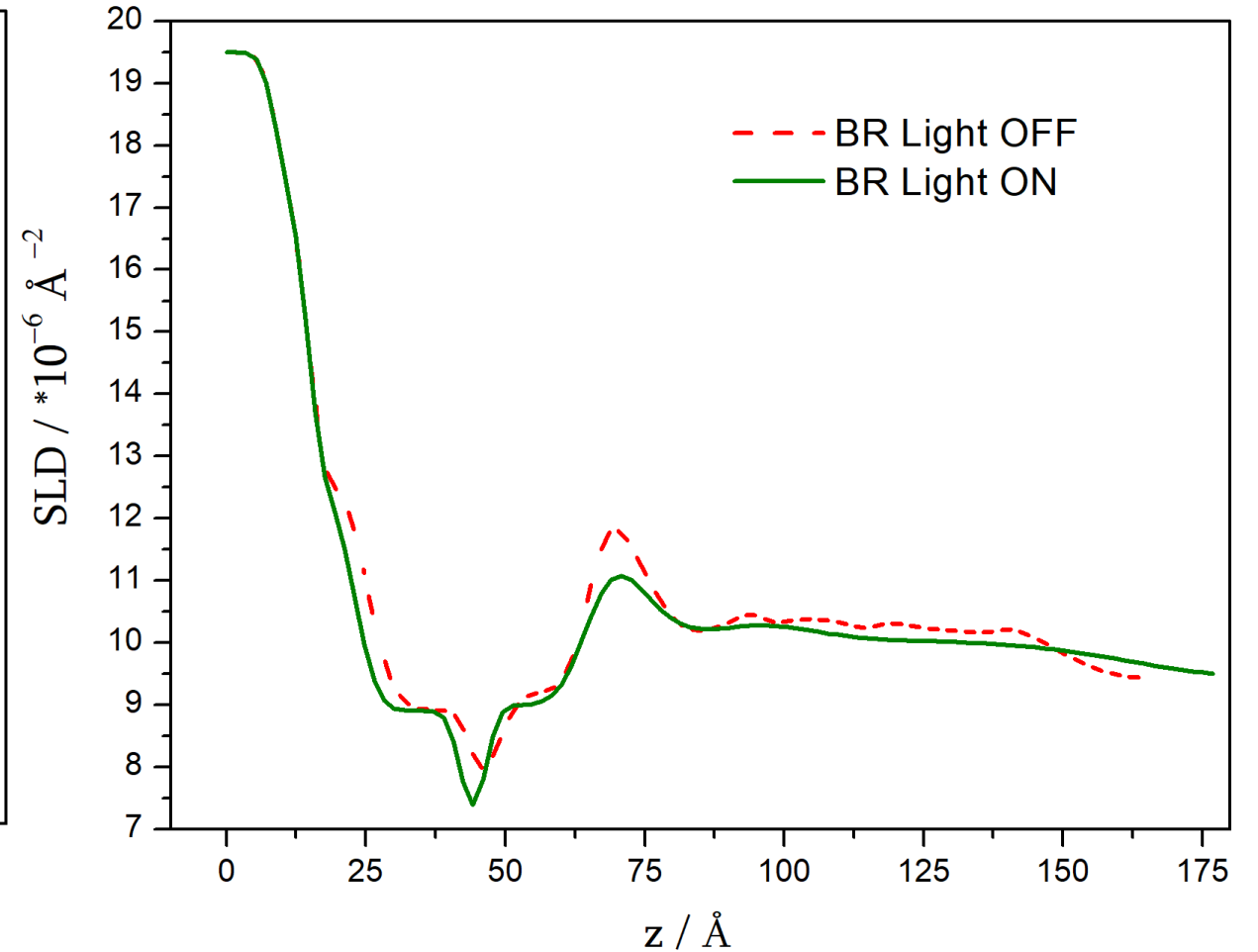


# Specular SR Reflectometry



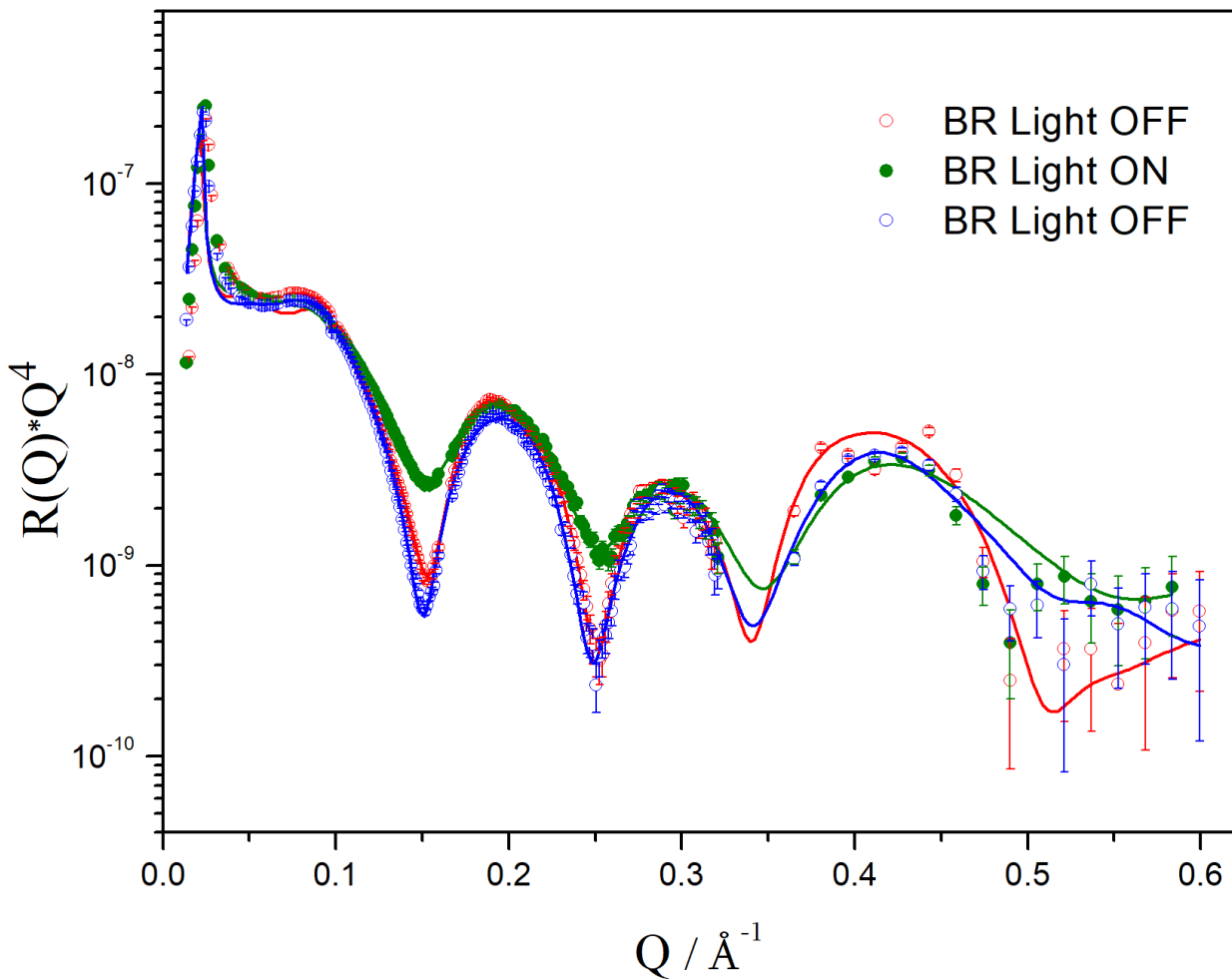
*DSPC **double bilayer** at 25°C after protein BR incorporation, with and without illumination.*

**SixS beamline (SOLEIL synchrotron, France).**



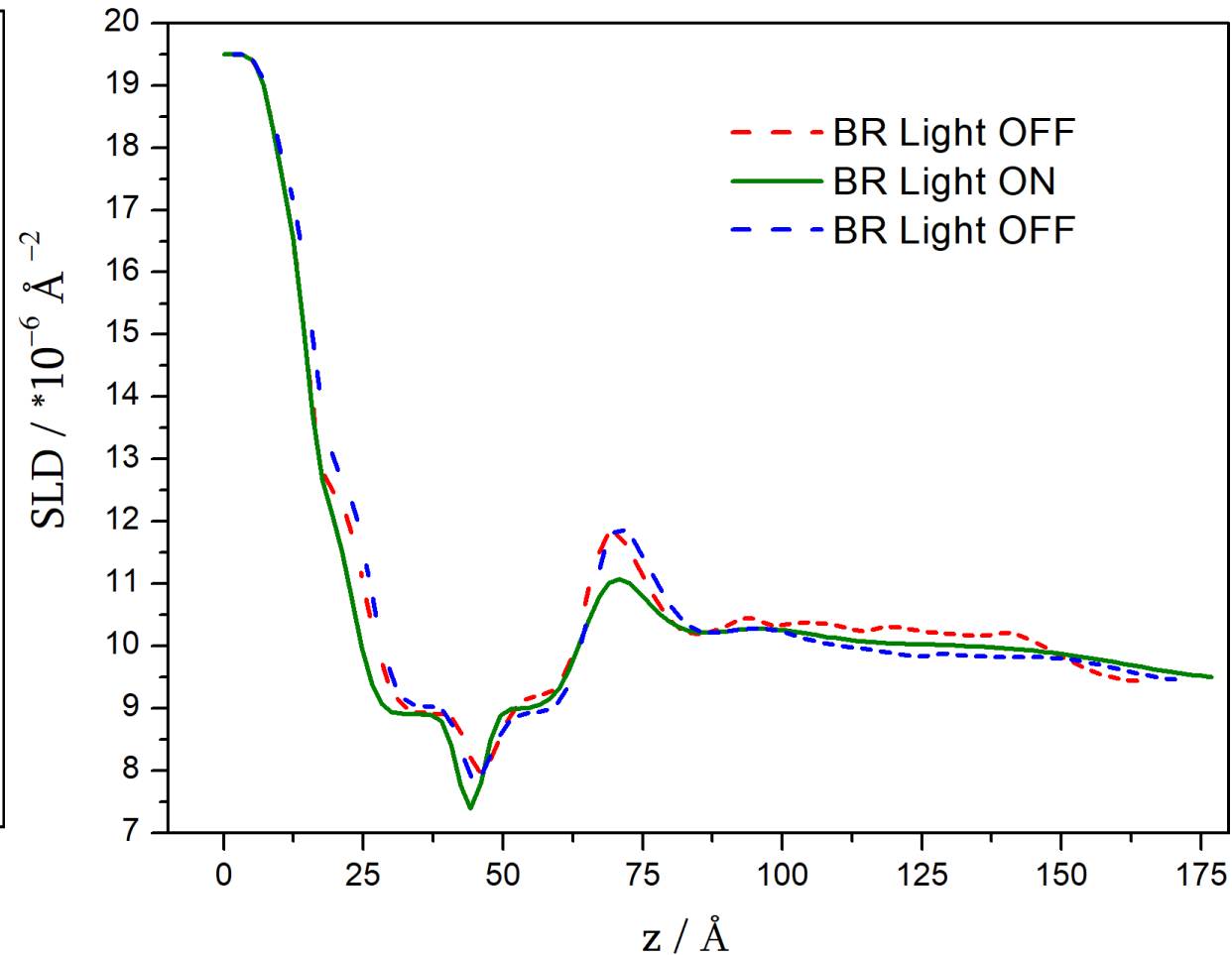
*SLD profiles corresponding to the fits.*

# Specular SR Reflectometry



*DSPC **double bilayer** at 25°C after protein BR incorporation, with and without illumination.*

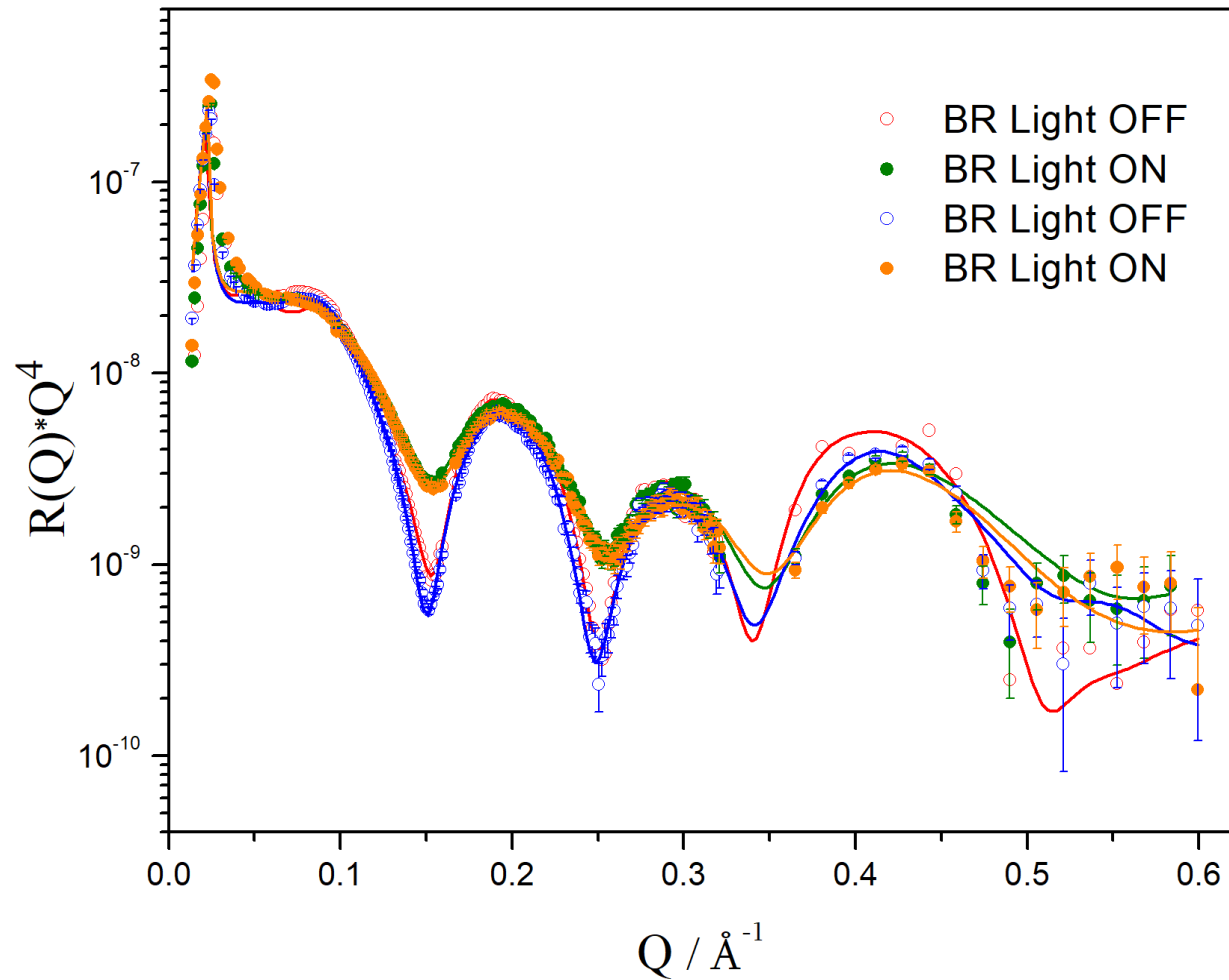
**SixS beamline (SOLEIL synchrotron, France).**



*SLD profiles corresponding to the fits.*

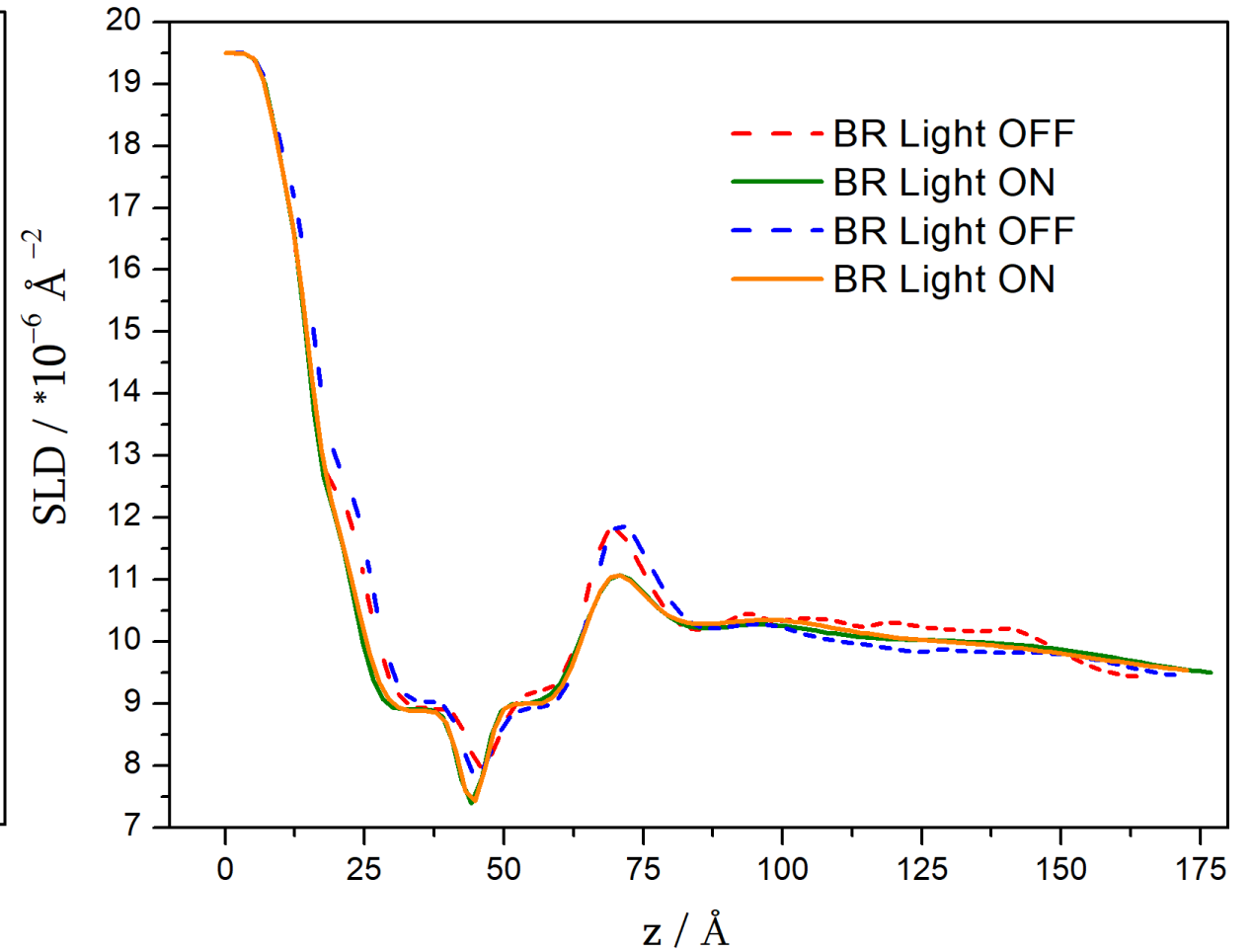
*The reversible effect of light illumination is shown.*

# Specular SR Reflectometry



*DSPC **double bilayer** at 25°C after protein BR incorporation, with and without illumination.*

**SixS beamline ( SOLEIL synchrotron, France).**

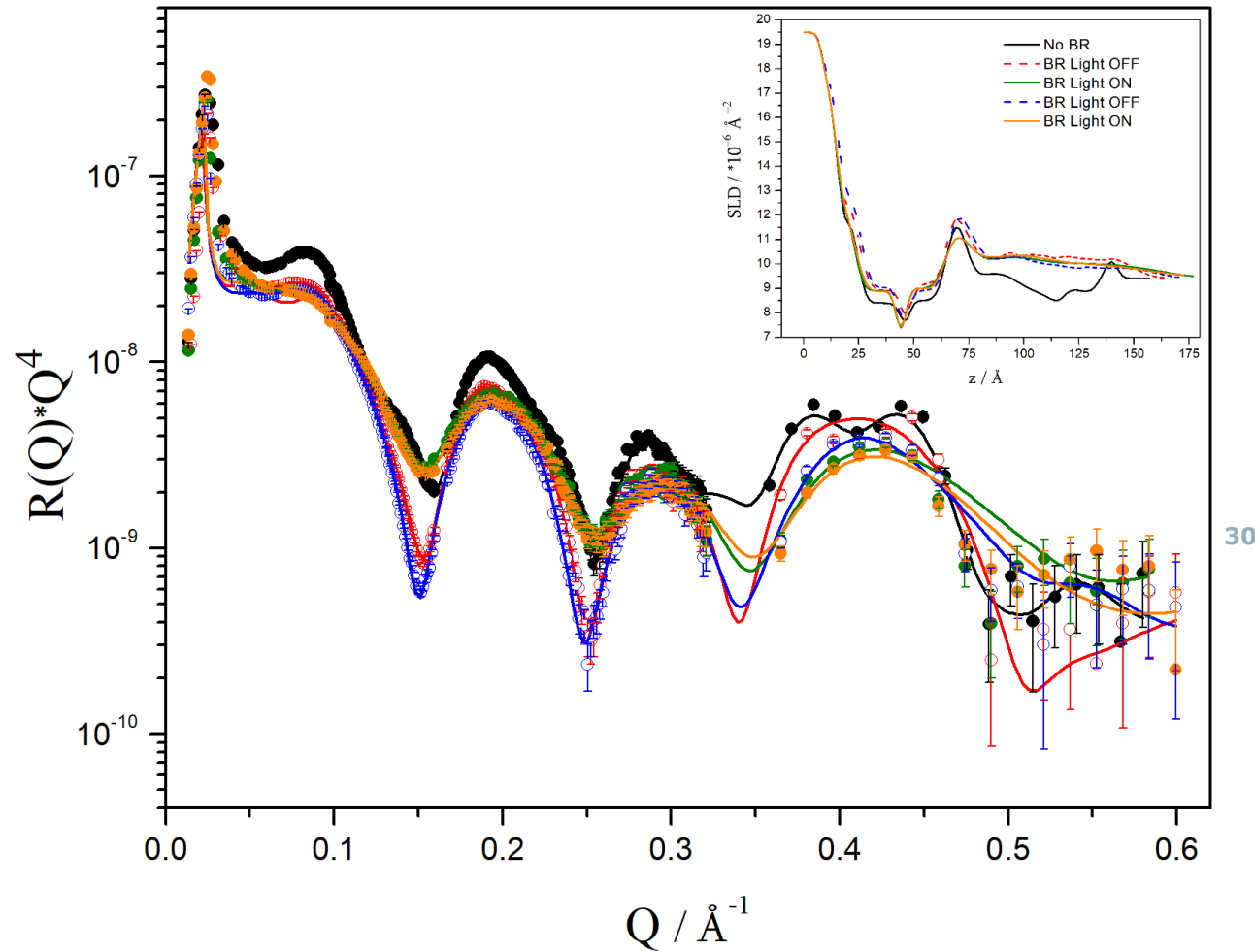
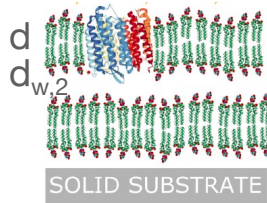


*SLD profiles corresponding to the fits.*

*The reversible effect of light illumination is shown.*



# Specular SR Reflectometry



DSPC **double bilayer** at 25°C with and without BR, with and without illumination. **Inset:** SLD profiles.

**SixS beamline (SOLEIL synchrotron, France).**

BR incorporation:

	<b>No BR</b>	<b>BR</b>
SLD <sub>ch21</sub> [ $10^{-4} \text{ nm}^2$ ]	$8.85 \pm 0.05$	$10.5 \pm 0.05$
d [nm]	$5.4 \pm 0.3$	$6.1 \pm 0.3$
d <sub>w,2</sub> [nm]	$1.5 \pm 0.2$	$1.3 \pm 0.2$
$\sigma_{\text{spec}}$ [nm]	$0.2 \pm 0.1$	$0.8 \pm 0.2$

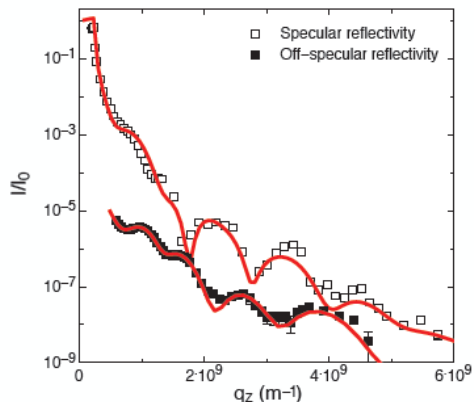
Light OFF ↔ Light ON:

	<b>Light OFF</b>	<b>Light ON</b>
d [nm]	$6.1 \pm 0.5$	$7.2 \pm 0.5$
d <sub>w,2</sub> [nm]	$1.3 \pm 0.2$	$1.8 \pm 0.2$
$\sigma_{\text{spec}}$ [nm]	$0.8 \pm 0.2$	$1.5 \pm 0.2$

30

# Off-specular reflection with and without illumination

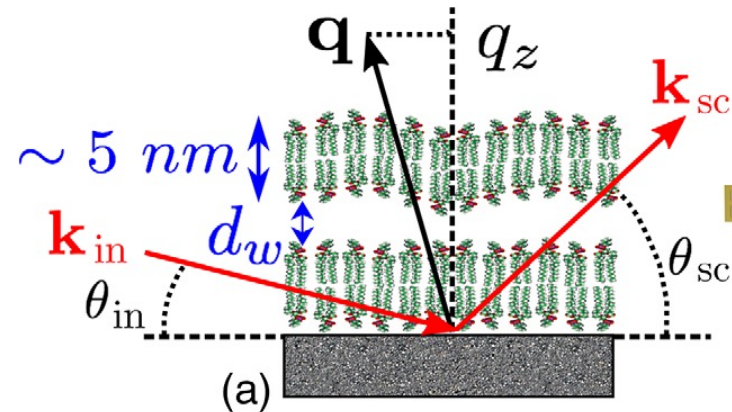
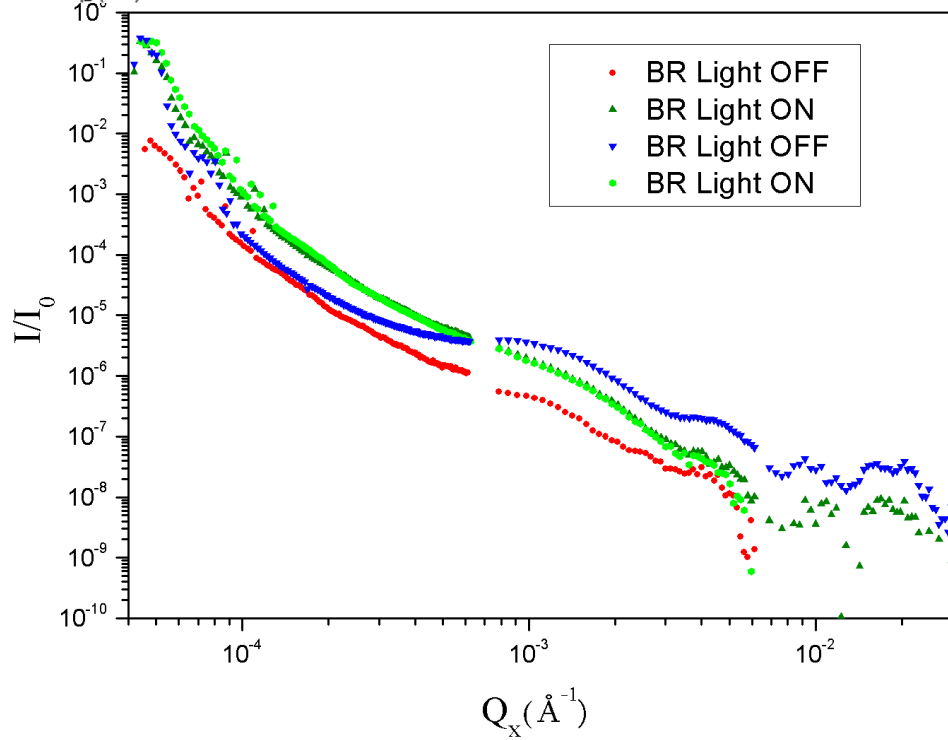
$\langle \delta\rho(r)\delta\rho(r') \rangle \sim$  *static* roughness, defects, fluctuations ...



Fitting with models of correlation functions:

- ➔ **structure**,
- ➔ **elastic parameters** (bending modulus, surface tension),
- ➔ **interaction potentials**.

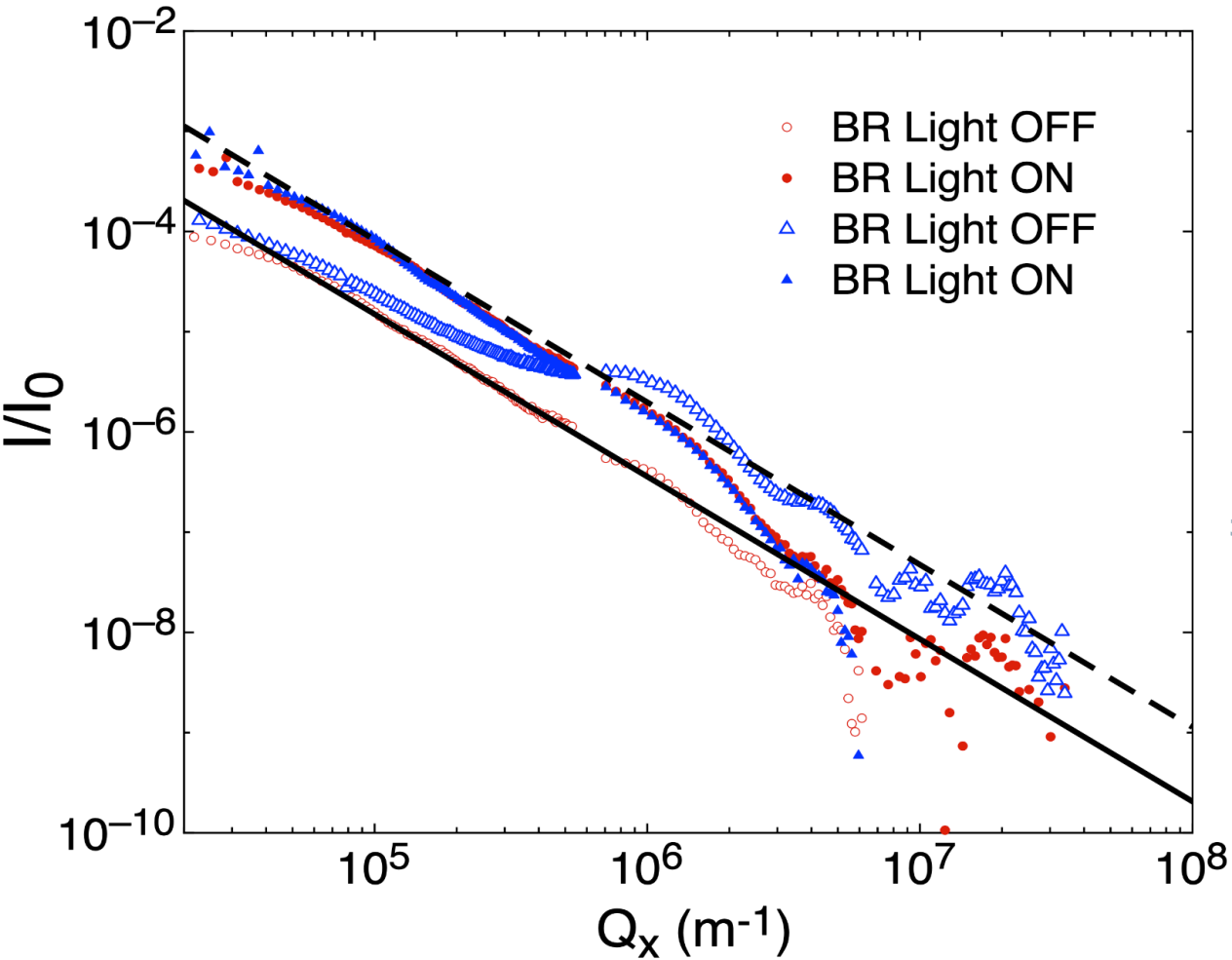
Malaquin et al., EPJE (2010), Hemmerle et al., PNAS (2012)



$$\langle z(q_{\parallel})z(q_{\parallel}) \rangle = \frac{k_B T}{A + \gamma q_{\parallel}^2 + \kappa q_{\parallel}^4}$$

... Analysis in progress

# Off-specular X-ray Reflectometry



*Off-specular XRR for the same DSPC double bilayer.*

*Reversible effect of light illumination.*

$$\frac{(I/I_0)_{\text{ON,off-spec}, Q_x \rightarrow 0}}{(I/I_0)_{\text{OFF,off-spec}, Q_x \rightarrow 0}} = \frac{\sigma_{\text{ON,off-spec}}^2}{\sigma_{\text{OFF,off-spec}}^2} = 4.1 \pm 0.8$$

**Effective temperature:**

$$\left(\frac{T_{\text{eff}}}{T}\right)_{\text{spec}} = \frac{\sigma_{\text{ON,spec}}^2}{\sigma_{\text{OFF,spec}}^2} = 3.5 \pm 2.5$$

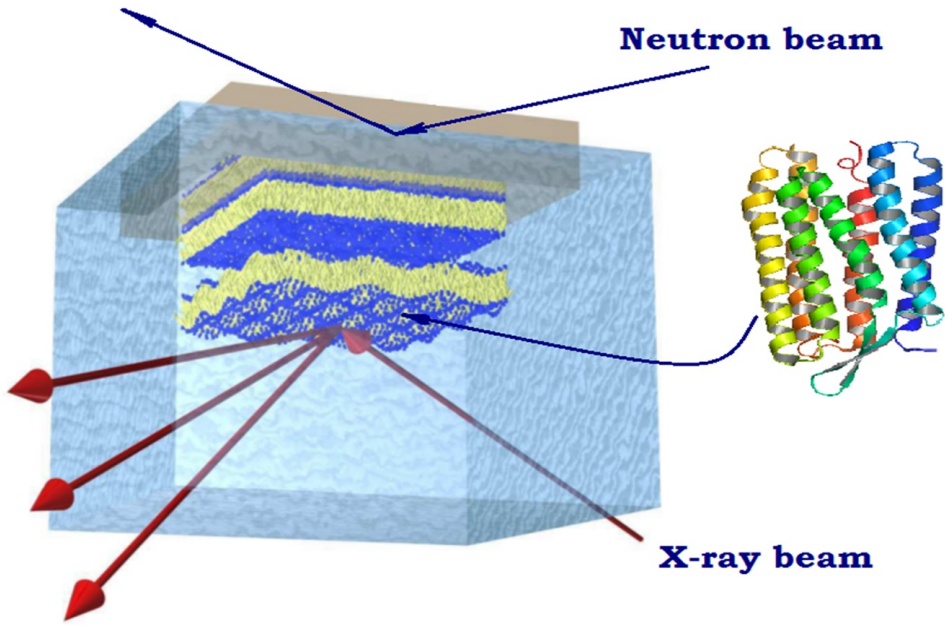
$$\left(\frac{T_{\text{eff}}}{T}\right)_{\text{off-spec}} = \frac{\sigma_{\text{ON,off-spec}}^2}{\sigma_{\text{OFF,off-spec}}^2} = 4.1 \pm 0.8$$

Micropipette aspiration experiments  
by J.-B. Manneville et al.:

$$1.7 \leq \frac{T_{\text{eff}}}{T} \leq 2.7$$



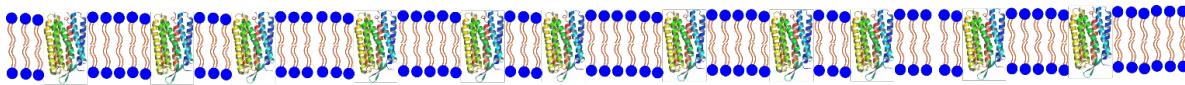
# SUMMARY



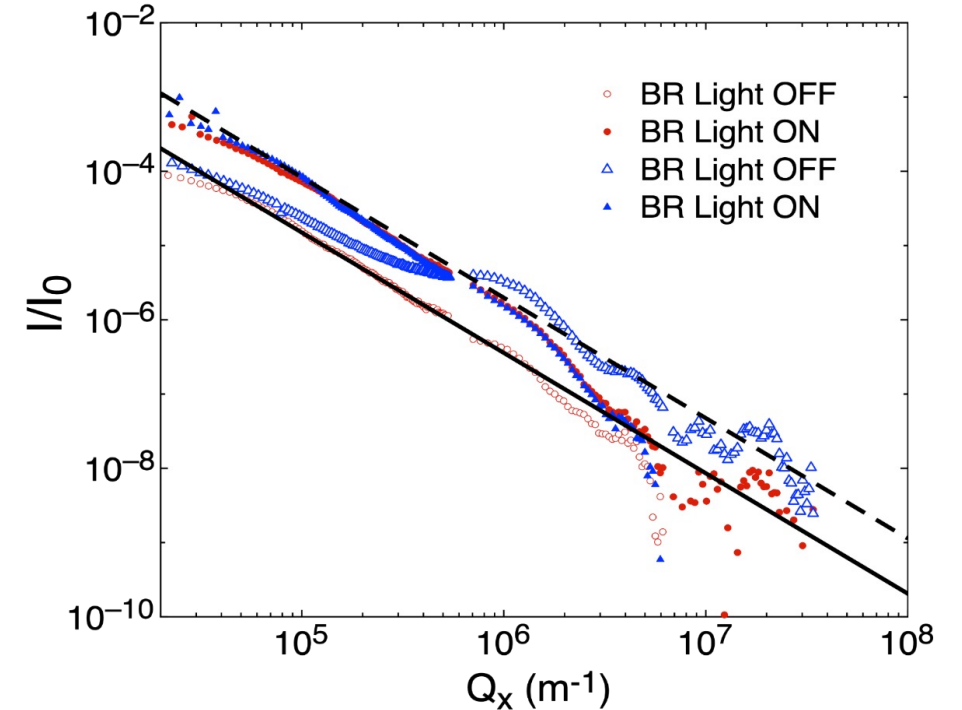
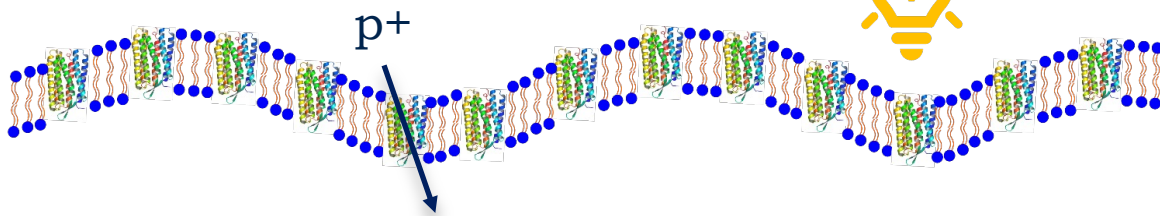
- Successful BR incorporation into membrane-mimic systems
- Reversible effect of light-induced protein activity on membrane structure and fluctuations
- Magnification of membrane fluctuations

33

No activity

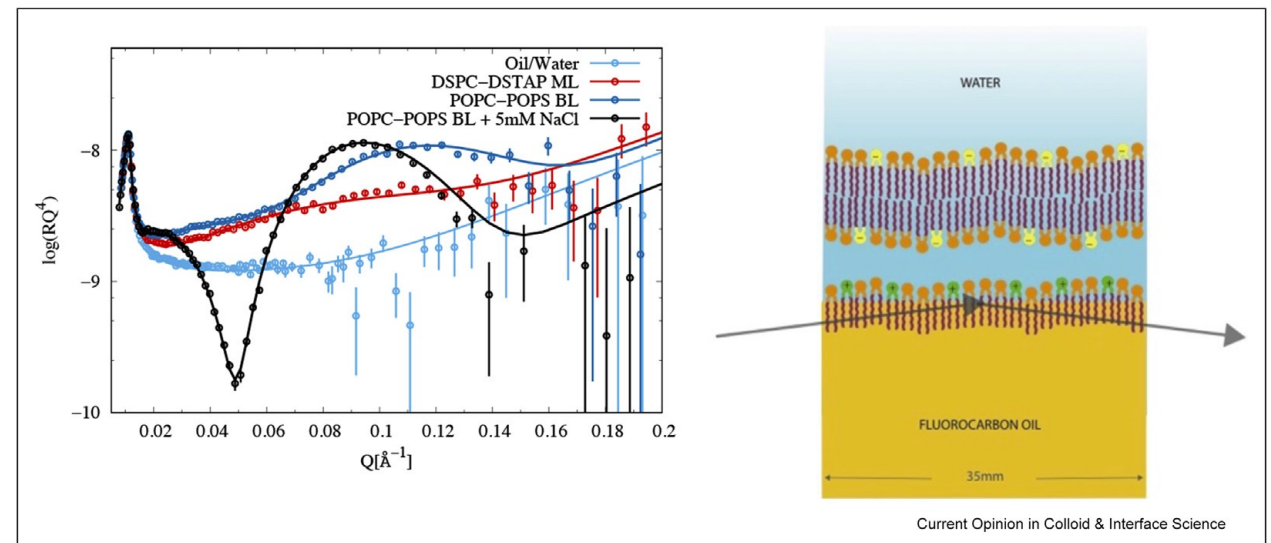
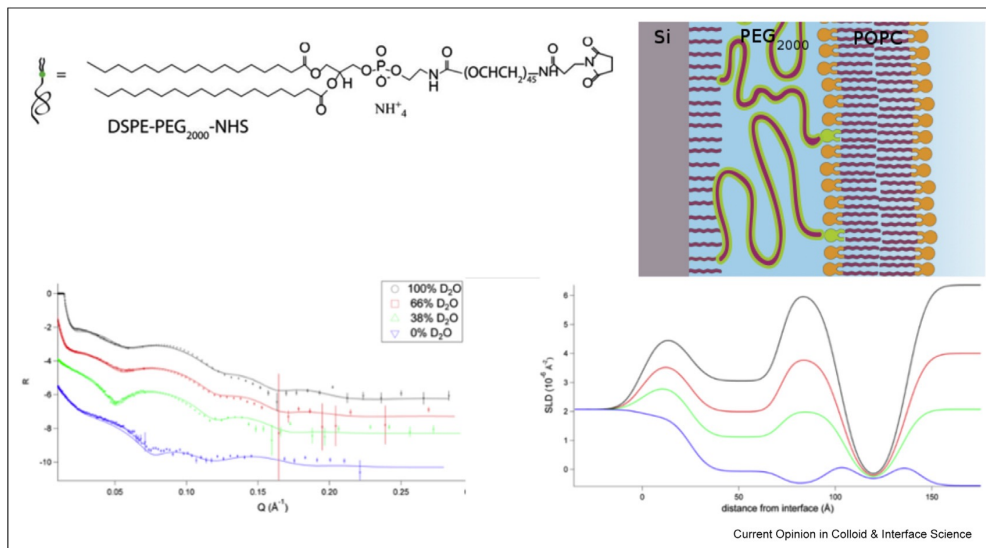
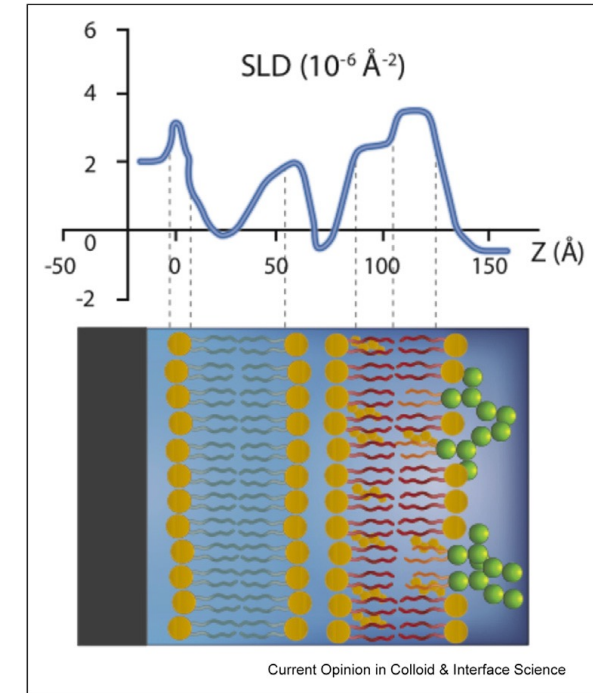


BR is active



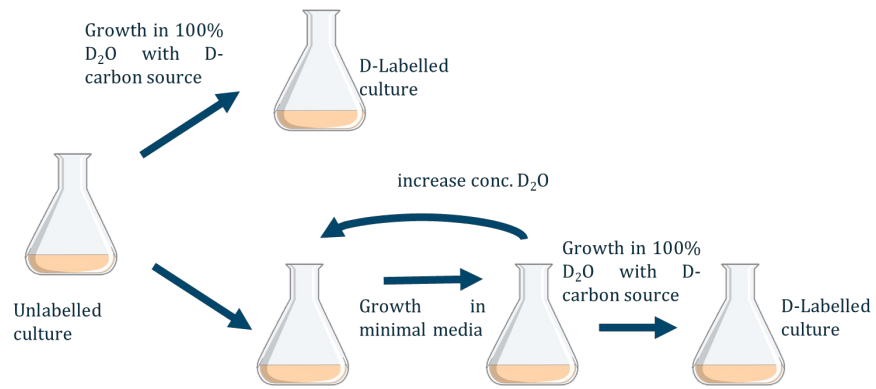
# Neutrons and model membranes: Moving towards complexity

Giovanna Fragneto<sup>1</sup>  , Robin Delhom<sup>2</sup>, Loïc Joly<sup>1,3</sup>, Ernesto Scoppola<sup>4</sup>





# Moving towards complexity: deuterated (and non) lipid extraction and purification



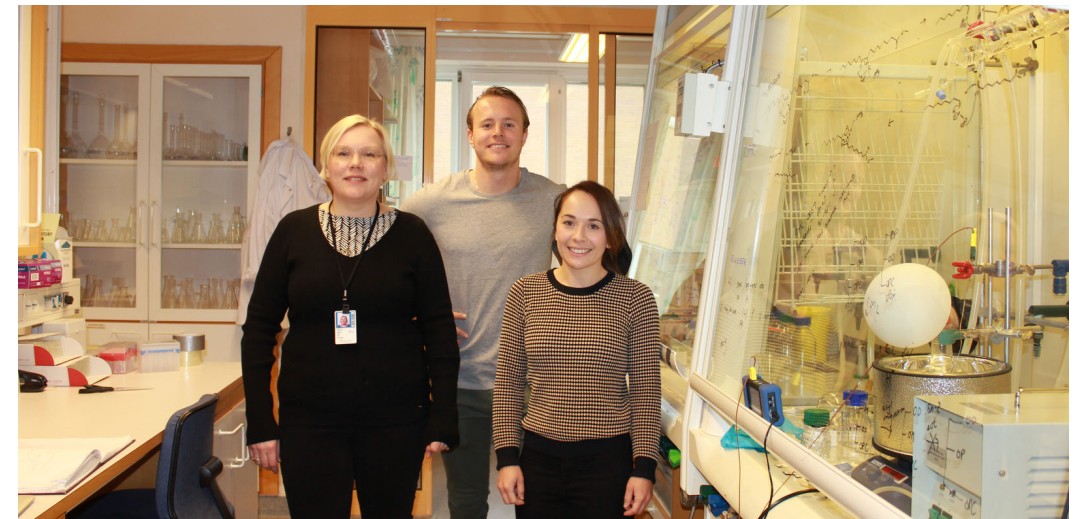
[www.ill.eu/L-Lab](http://www.ill.eu/L-Lab)





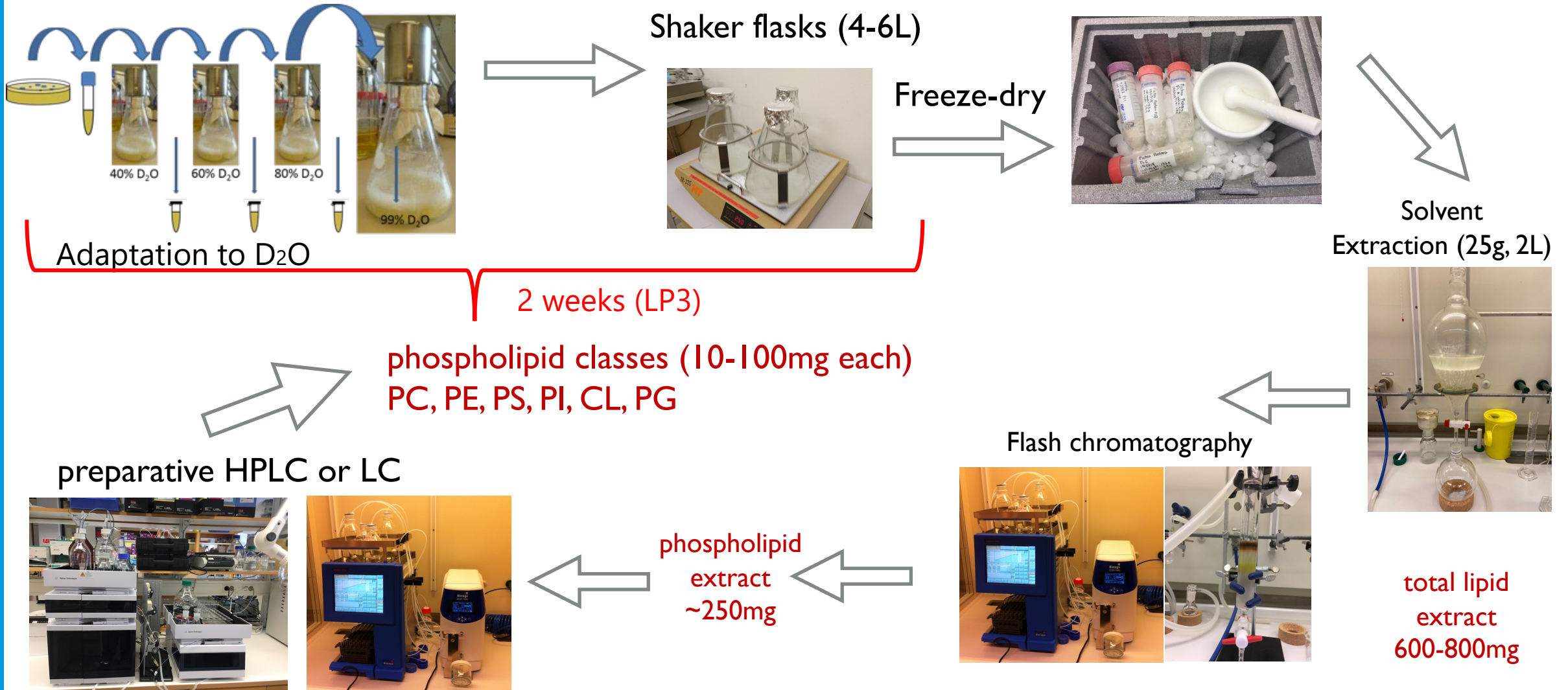
# Yeast lipid production at DEMAX

The Deuteration and  
Macromolecular Crystallisation  
(DEMAX) platform supports life  
science and soft matter research  
users of neutron instruments.



# Workflow for purification

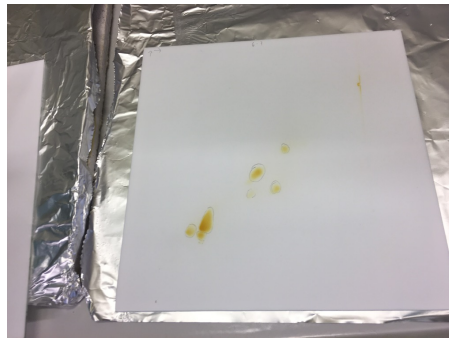
From cell culture to phospholipid extracts ~ 4 PW



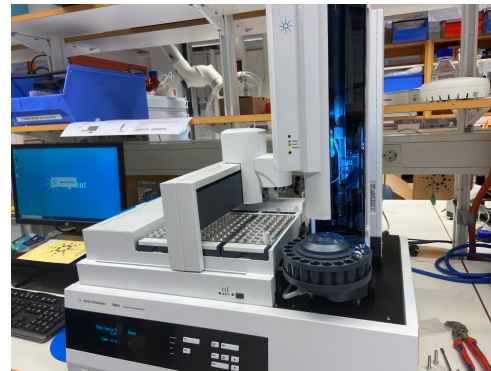
# Workflow for lipid analysis

Lipid composition ~1 PW per TLC plate/ batch

Preparative TLC



Automated GC sample preparation



% deuteration:  
FTIR check internal (1 day)  
GC/LC-MS external (availability)



GC-FID



- total PL extract
- each PL class

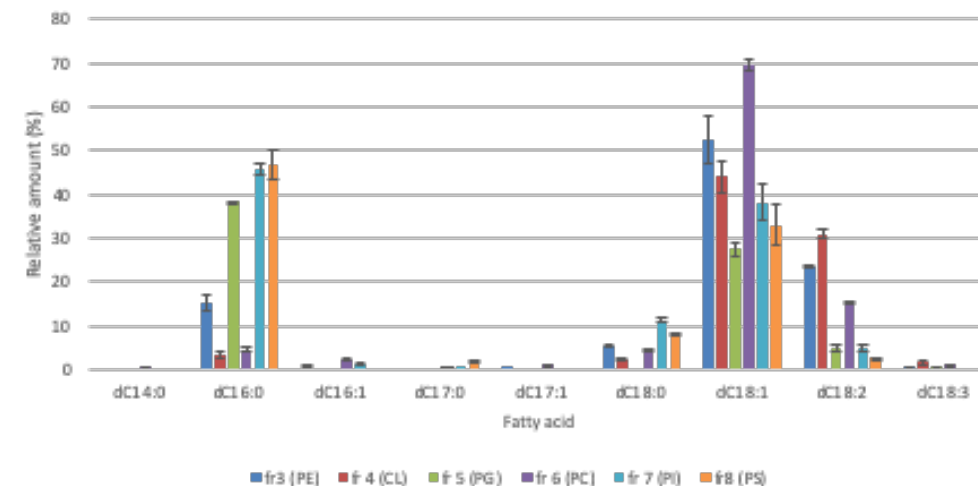
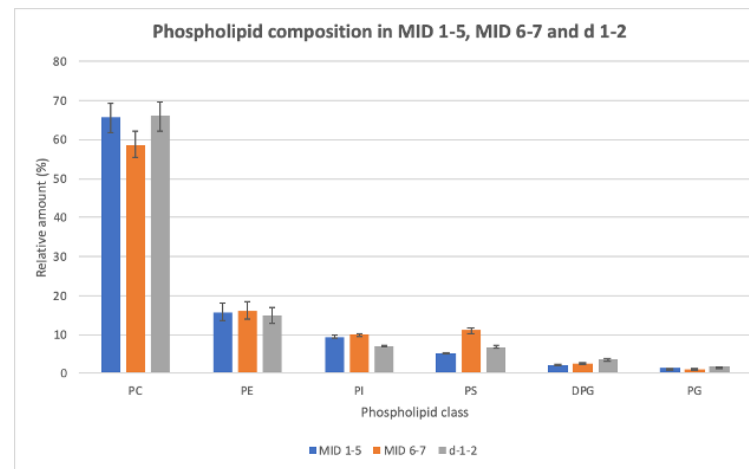
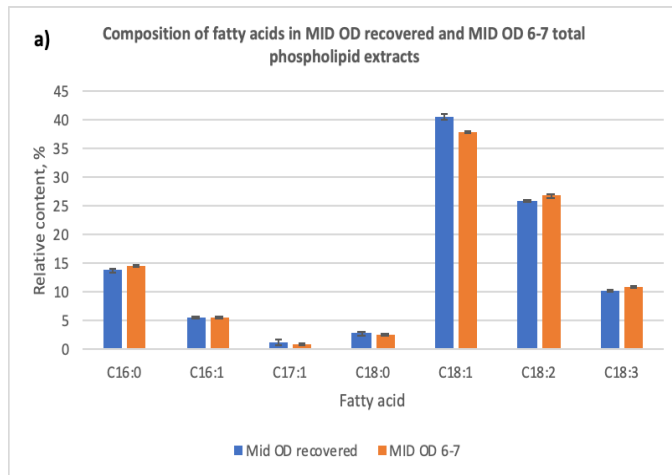
analysis in triplicate for errors in composition

reusability good after neutrons:

Phospholipid composition

Lipid chain composition in each PL class

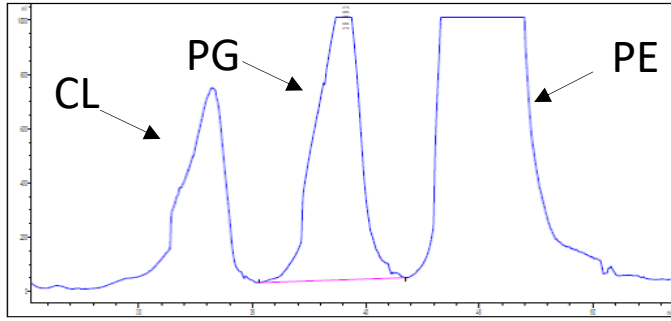
Composition of fatty acids deuterated *Pichia* Phospholipids





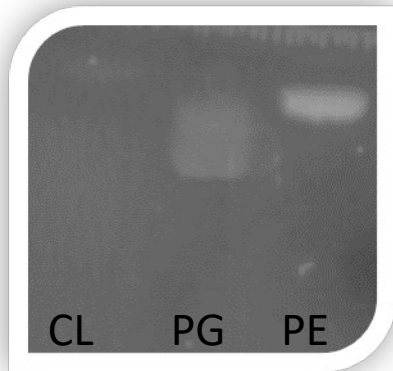
# Separation of the phospholipid classes by Normal phase-HPLC

Successful separation of phospholipid classes from *P.pastoris* and *E.coli* total lipid extracts.



*E.Coli* lipids chromatogram

PL	RT
CL	30 m
PG	35 m
PE	40 m



PC = ~51 mg  
 PE = ~24 mg  
 PI = ~13 mg  
 PS = ~18 mg

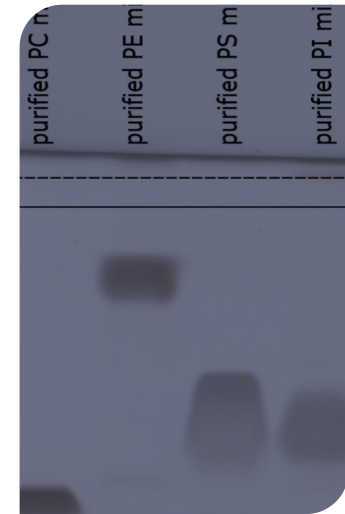
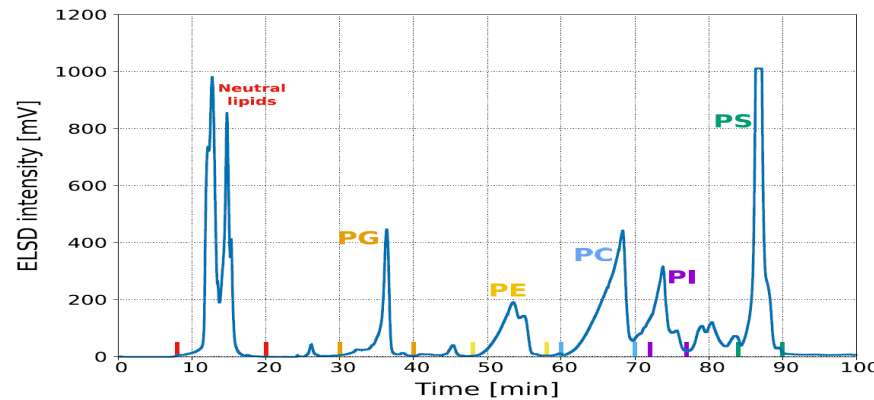
A total of ~106 mg  
 GPL mixture from 5  
 g/yeast biomass

TOTAL YIELD

A total of ~89 mg  
 GPL mixture from 5  
 g/*E.coli* biomass

PE = ~54 mg  
 PG = ~23 mg  
 CL = ~13 mg

*Pichia pastoris* lipids chromatogram



PL	RT
CL	30 m
PG	35 m
PE	43 m
PC	59 m
PI	72 m
PS	83 m

The implementation of an additional purification step by High Performance Liquid Chromatography-Evaporative Light Scattering Detector (HPLC-ELSD) enabled a better separation of the GPL mixtures from the neutral lipid fraction that includes sterols, and also allowed for the GPLs to be purified according to their different polar headgroups.

# Fatty acyl chain composition – *Pichia pastoris*

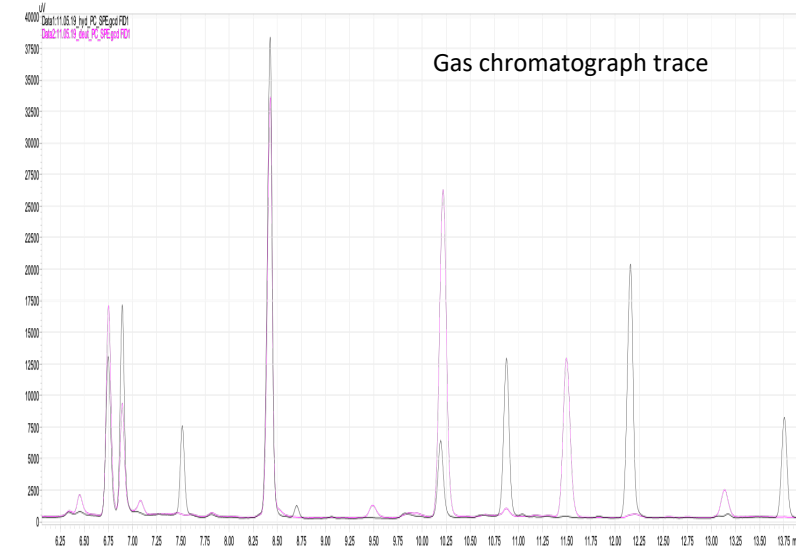
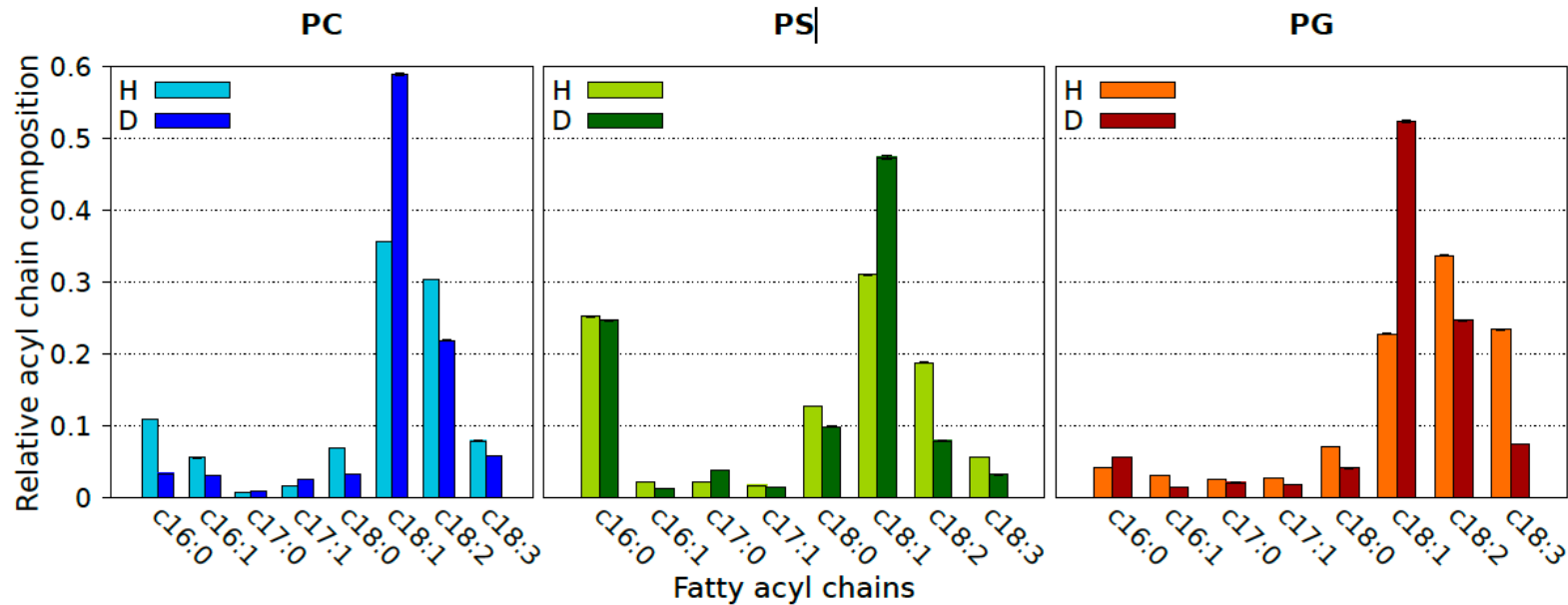


Figure 2: GC-FID analysis of the fatty acid distribution for the investigated GPL classes. Data are mean  $\pm$  S.D. of three technical repeats. A complete table with all the values can be found in *Supplementary Material Table S1*.

FAMES analyses revealed that deuteration triggered a significant increase in the oleic acid content = reflected across classes

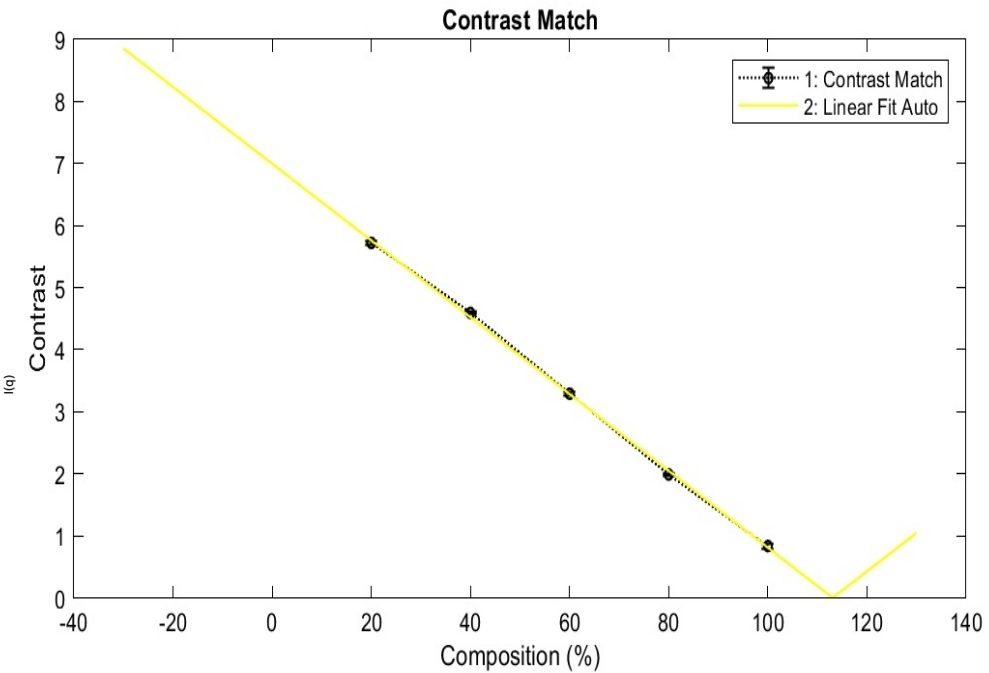
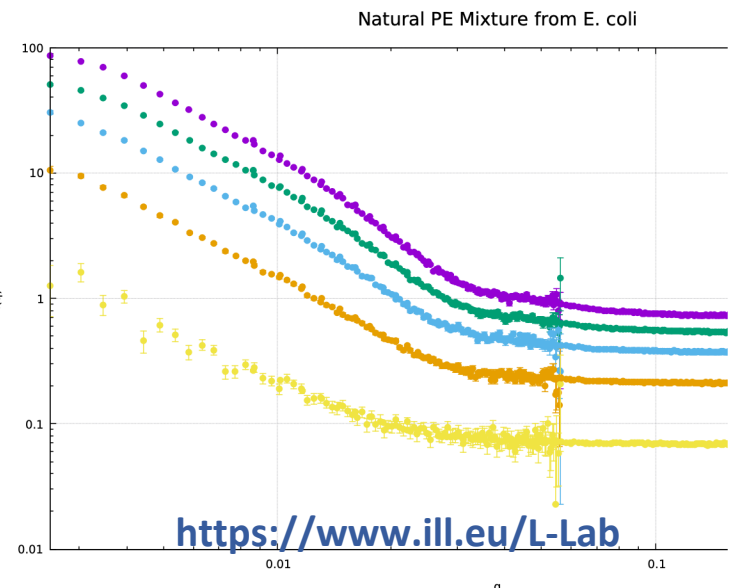
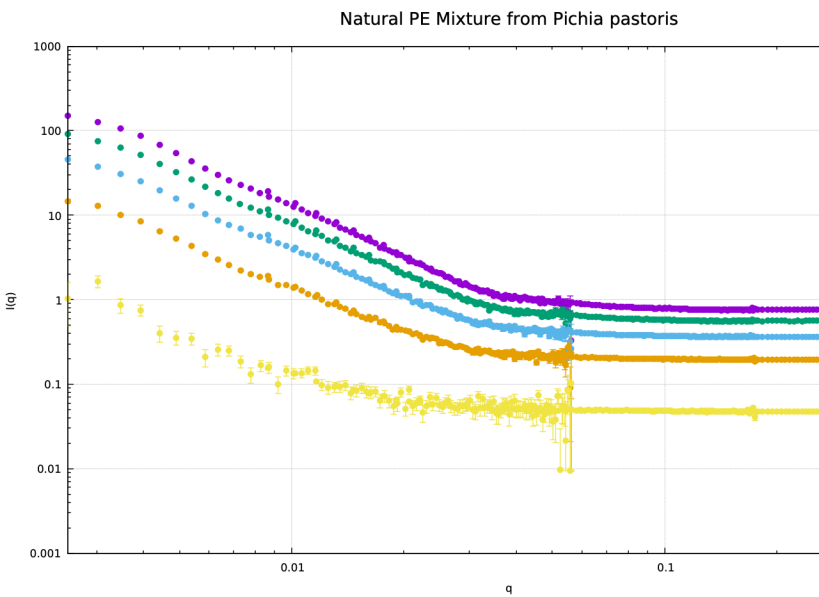
Decrease in the polyunsaturated FA content.

Decreased palmitic acid levels across the classes while an increase in the stearic acid is noticed in the acidic phospholipids

# Structural Characterization of the Purified Lipid Mixtures

- The structure of membrane mono/bilayers has been characterized by techniques (DSC, QCM) that are available within the PSCM.
- Neutron characterization of membrane mimics from the purified lipid mixtures has been carried out within the LSS and SMSS groups including:

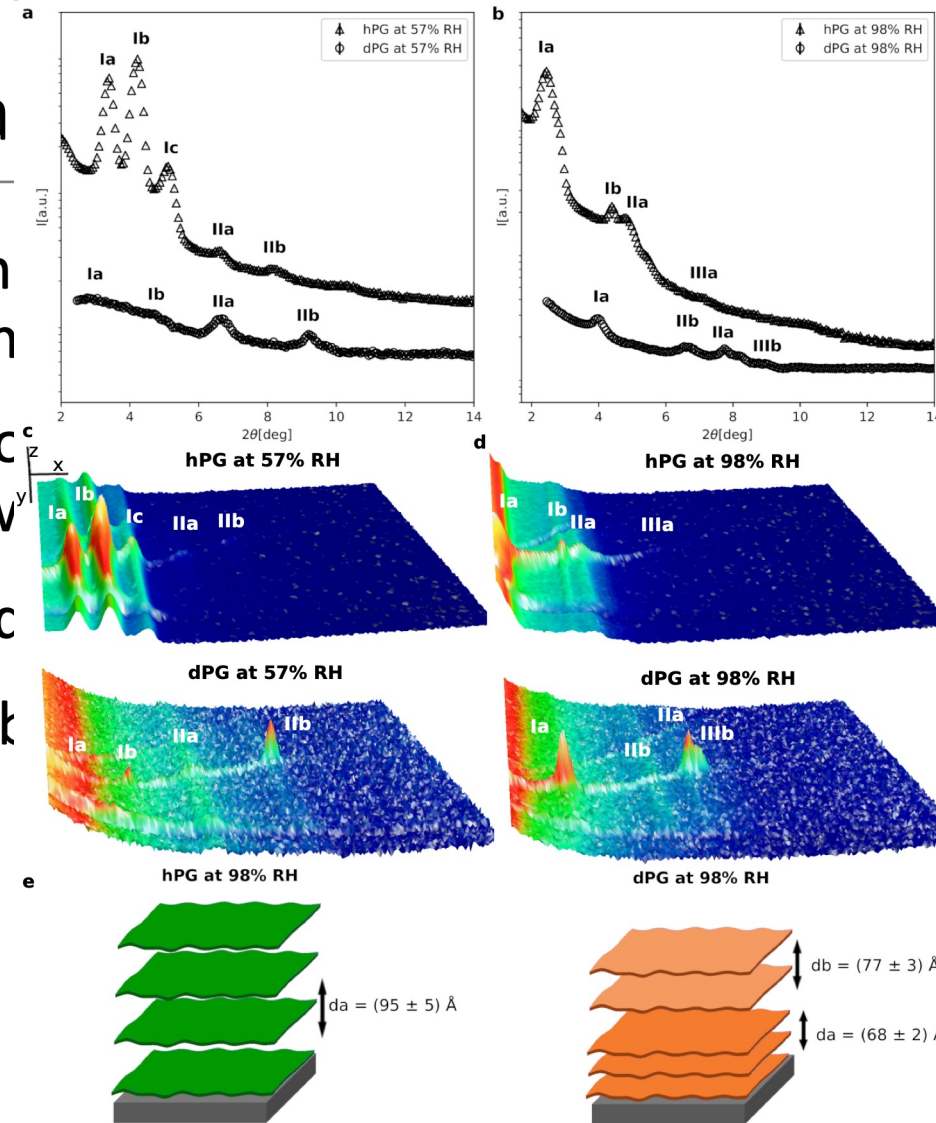
## 1.) SANS from liposomes, nanodiscs, vesicles.





# Structural Cha

- The structure of membran (QCM) that are available with
- Neutron characterization c been carried out at the ILL w
  - 1.) SANS from liposomes and
  - 2.) Diffraction from stacked k



# Lipid Mixtures

ized by techniques (DSC, ed lipid mixtures has ding: umidity),



## Structural Characterization of Natural Yeast Phosphatidylcholine and Bacterial Phosphatidylglycerol Lipid Multilayers by Neutron Diffraction

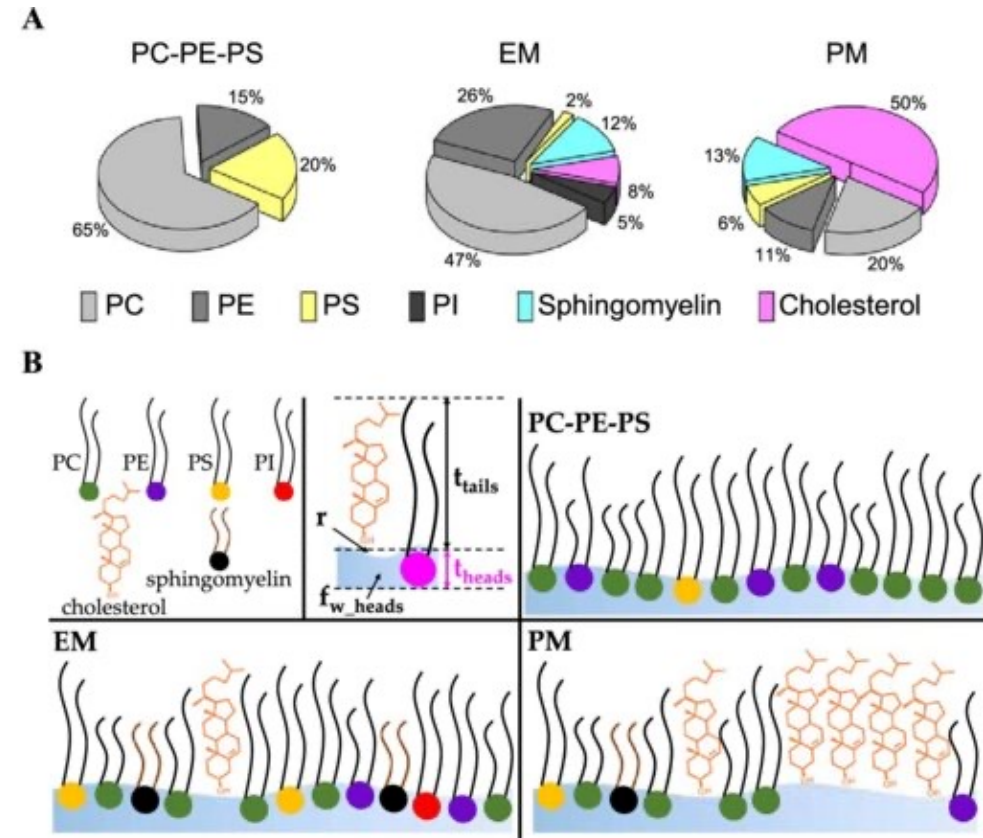
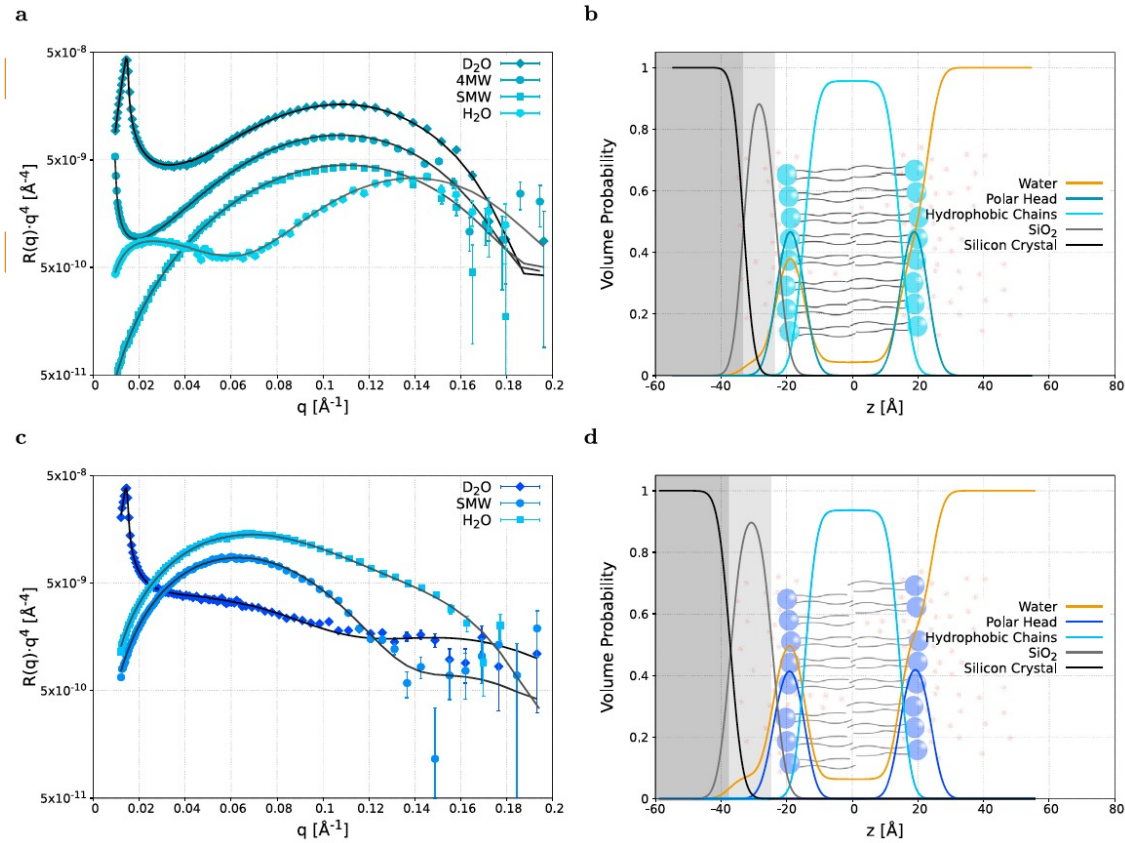
Alessandra Luchini<sup>1\*</sup>, Giacomo Corucci<sup>2,3</sup>, Krishna Chaitanya Batchu<sup>2</sup>, Valerie Laux<sup>2</sup>, Michael Haertlein<sup>2</sup>, Viviana Cristiglio<sup>2</sup> and Giovanna Fragneto<sup>2,3</sup>



The impact of deuteration on natural and synthetic lipids: A neutron diffraction study

Alessandra Luchini<sup>a,b</sup>, Robin Delhom<sup>a</sup>, Bruno Demé<sup>a</sup>, Valérie Laux<sup>a</sup>, Martine Moulin<sup>a</sup>, Michael Haertlein<sup>a</sup>, Harald Pichler<sup>c,d</sup>, Gernot A. Strohmeier<sup>c,e</sup>, Hanna Wacklin<sup>f,g</sup>, Giovanna Fragneto<sup>a,\*</sup>

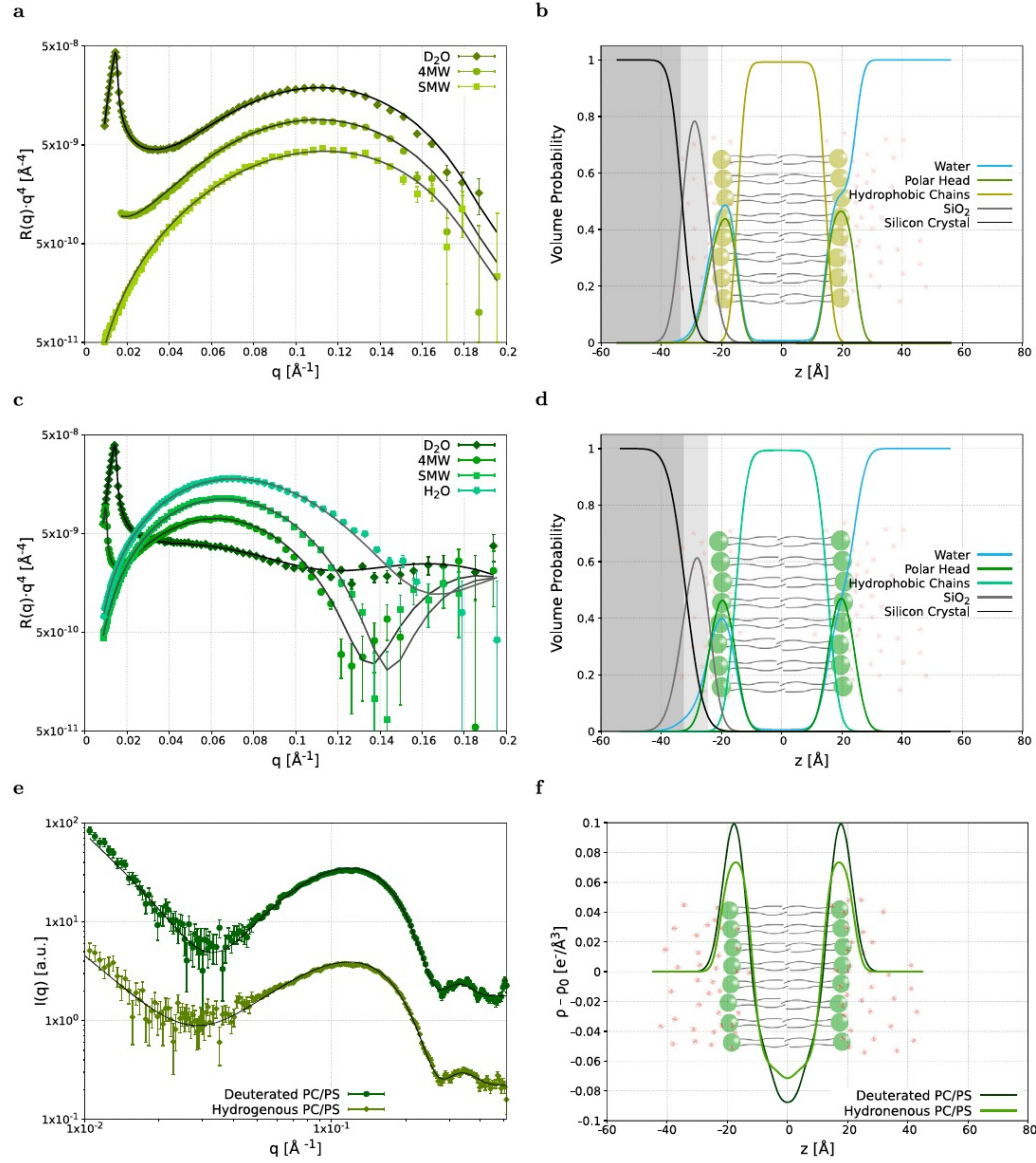
# Structural Characterization of the Purified Lipid Mixtures



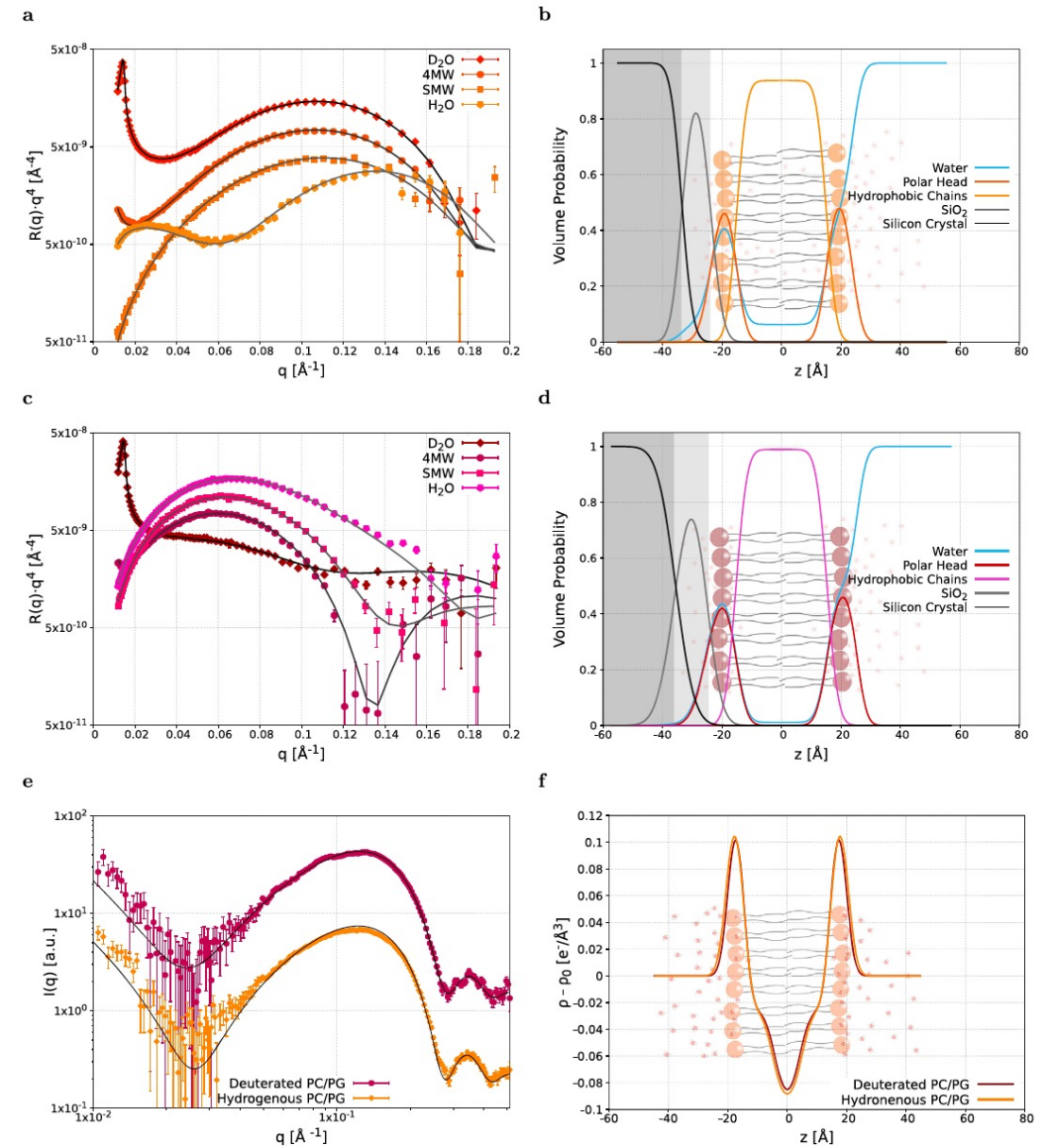
Santamaria, JCIS 2023



# PC/PS



# PC/PG



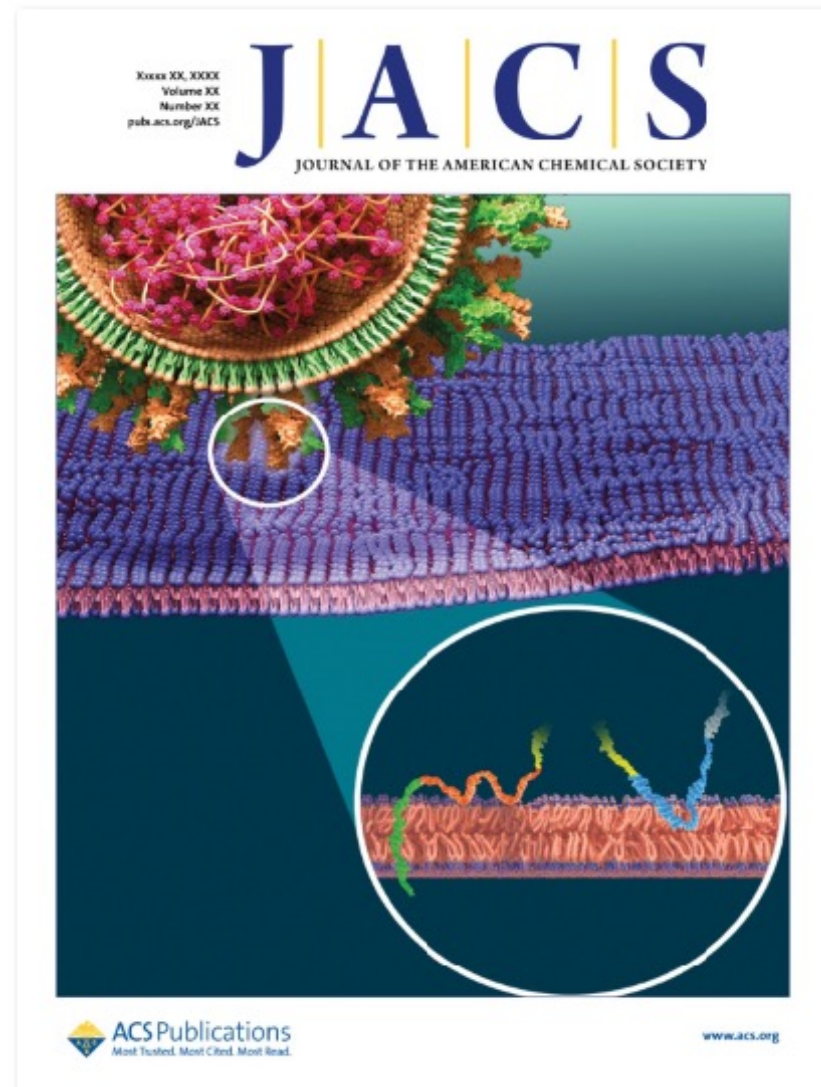
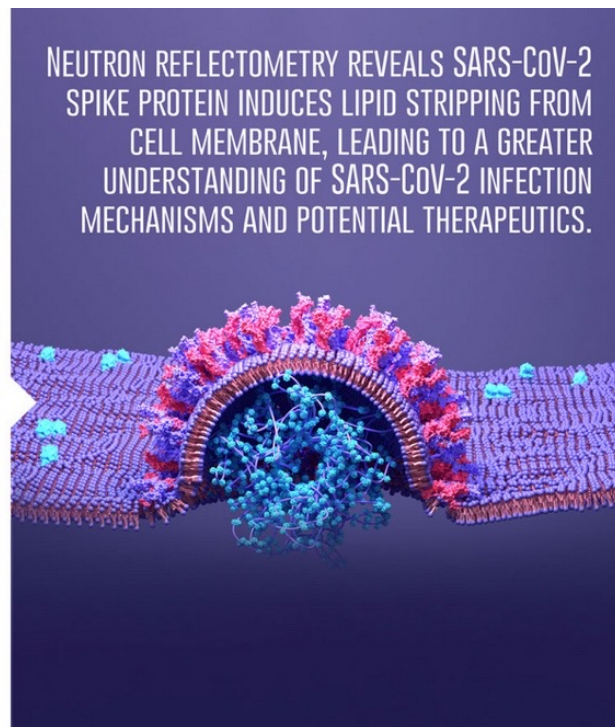
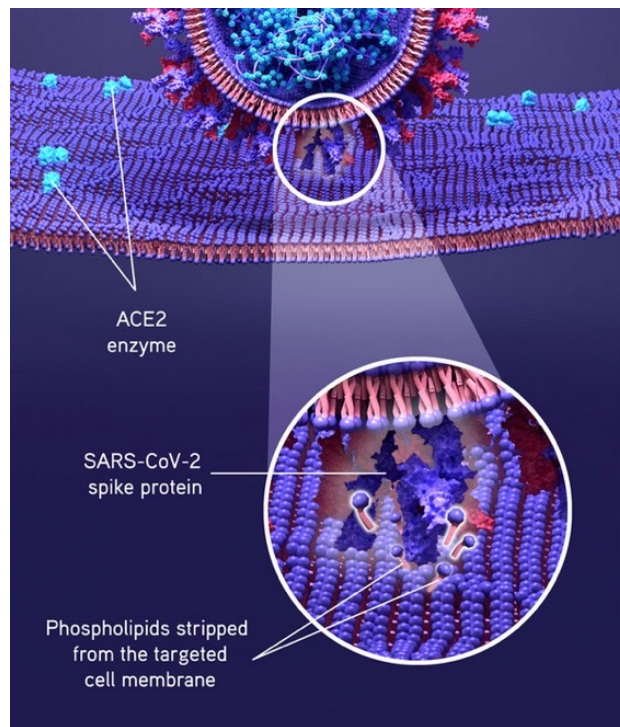
**All bilayers from natural extracts show the same structure at the interface (within errors)**





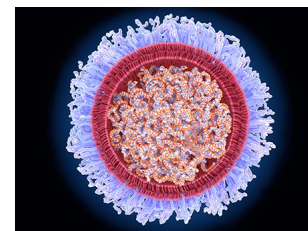
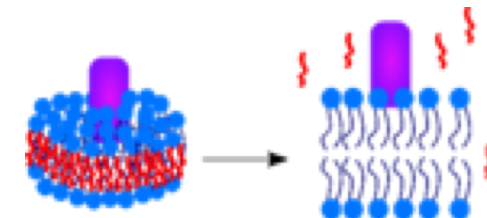
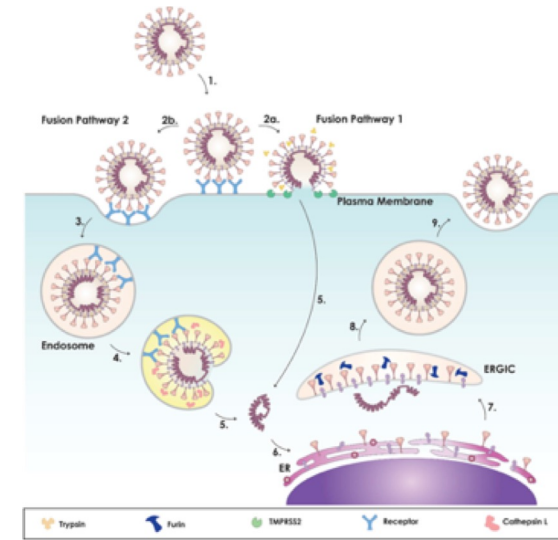
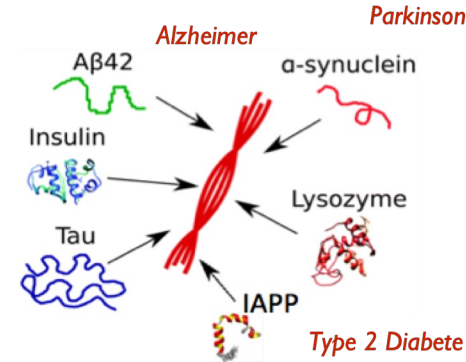
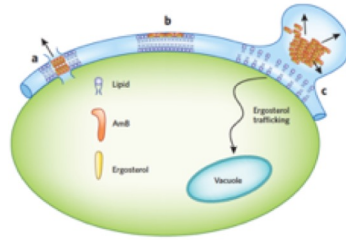
**OPEN** Lipid bilayer degradation induced by SARS-CoV-2 spike protein as revealed by neutron reflectometry

Alessandra Luchini<sup>1,5</sup>, Samantha Micciulla<sup>2,5</sup>, Giacomo Corucci<sup>2</sup>, Krishna Chaithanya Batchu<sup>2</sup>, Andreas Santamaria<sup>2</sup>, Valerie Laux<sup>2</sup>, Tamim Darwish<sup>3</sup>, Robert A. Russell<sup>3</sup>, Michel Thepaut<sup>4</sup>, Isabelle Bally<sup>4</sup>, Franck Fieschi<sup>4</sup> & Giovanna Fraeneto<sup>2,5</sup>

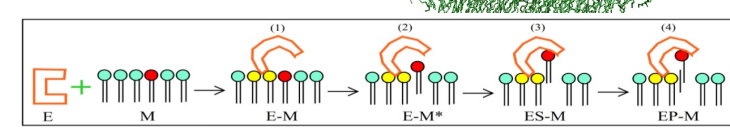
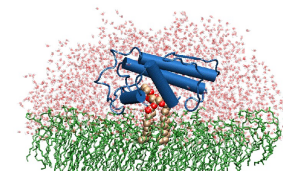
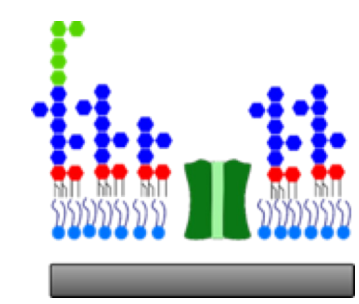


# Perspectives:

- ❖ Antibiotic resistance
- ❖ Amyloid interactions with membranes
- ❖ Viral entry
- ❖ Protein/peptide/drug/... membrane interactions
- ❖ Matched nano discs: low resolution protein structures
- ❖ Transmembrane protein reconstitution
- ❖ Effects of deuteration on membrane composition
- ❖ Enzyme mechanisms at membrane surface

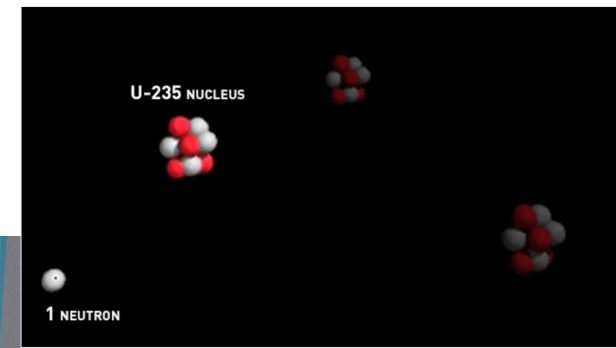
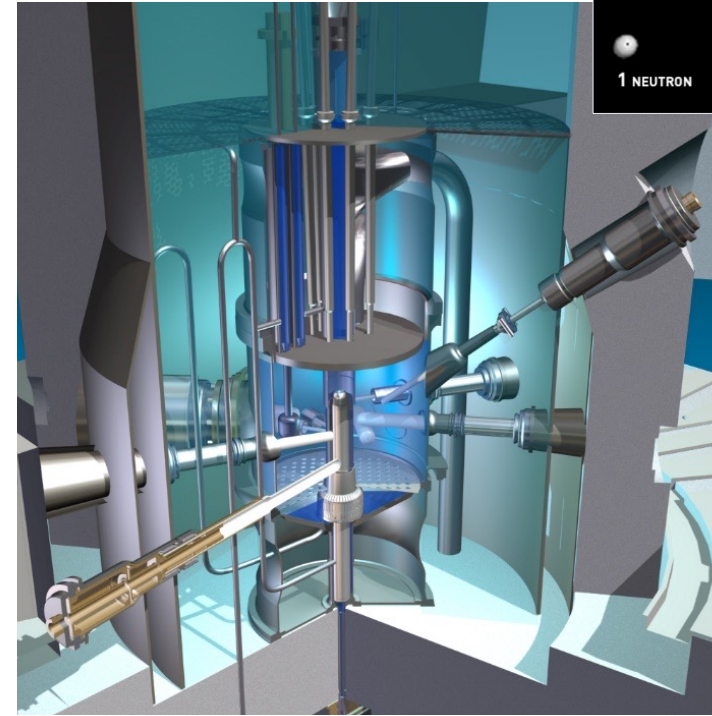
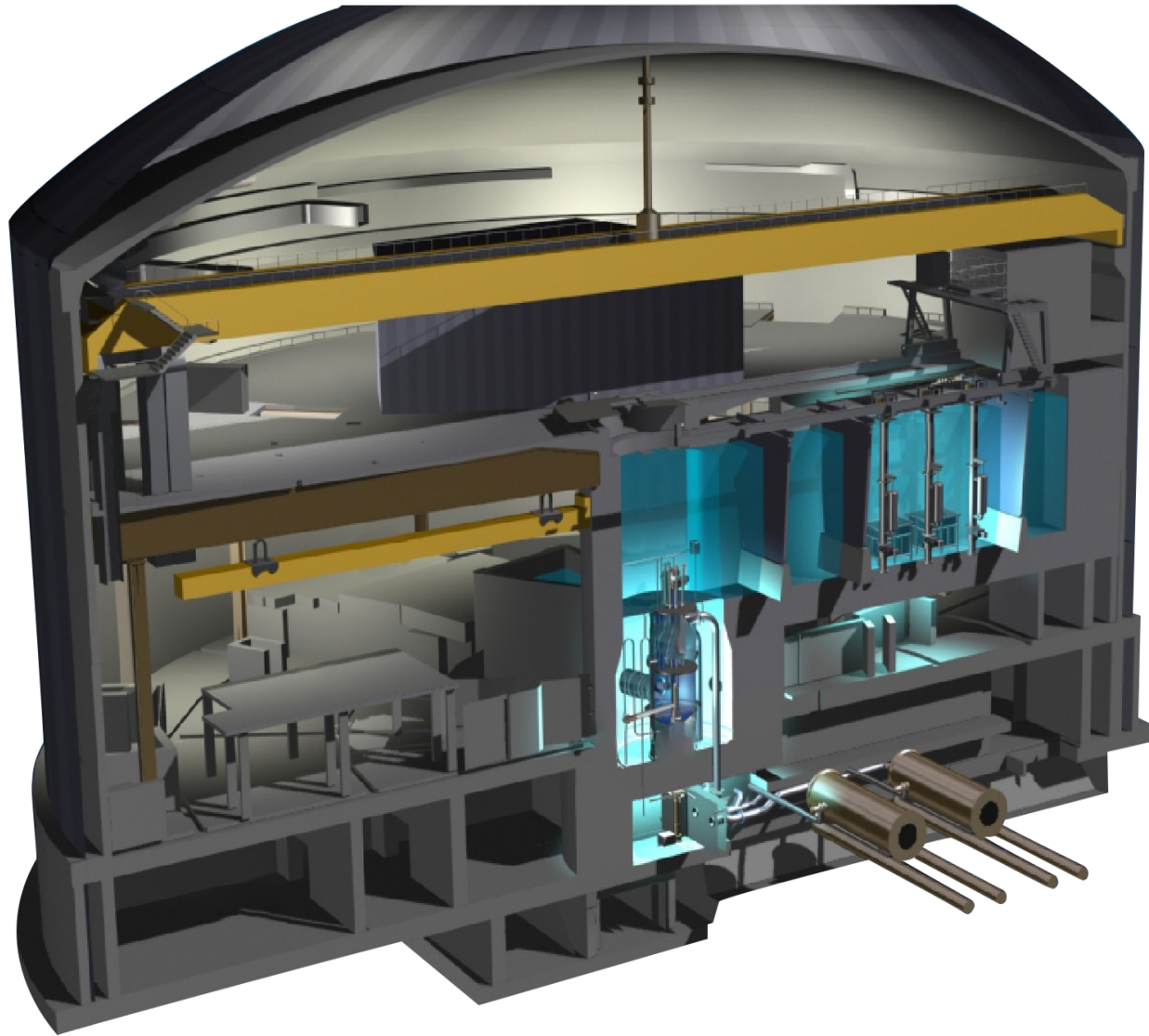


ACS Nano 2021





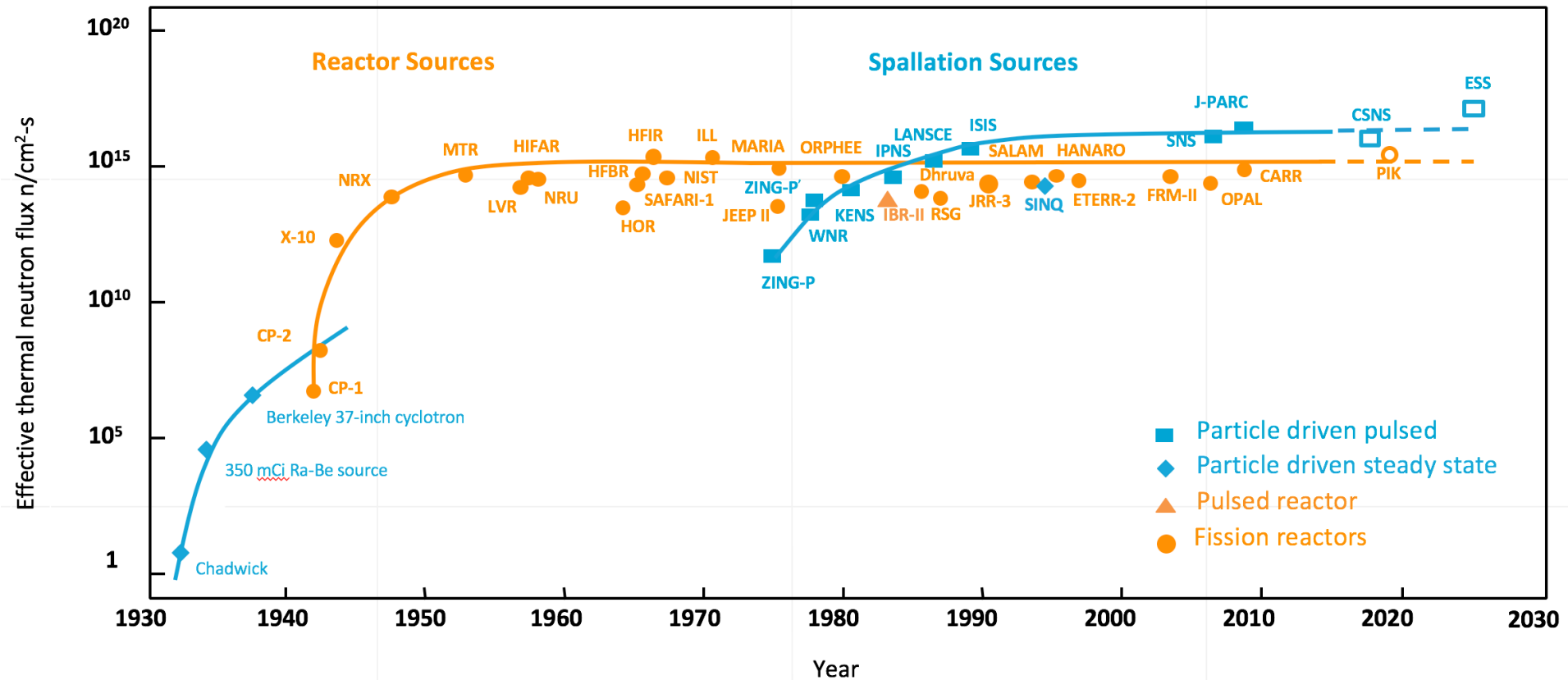
# The ILL Reactor



**A neutron source generating  
 $5 \times 10^{18}$  fast neutrons/sec  
at a max power of 58 MW**



# Neutron Source Brightness

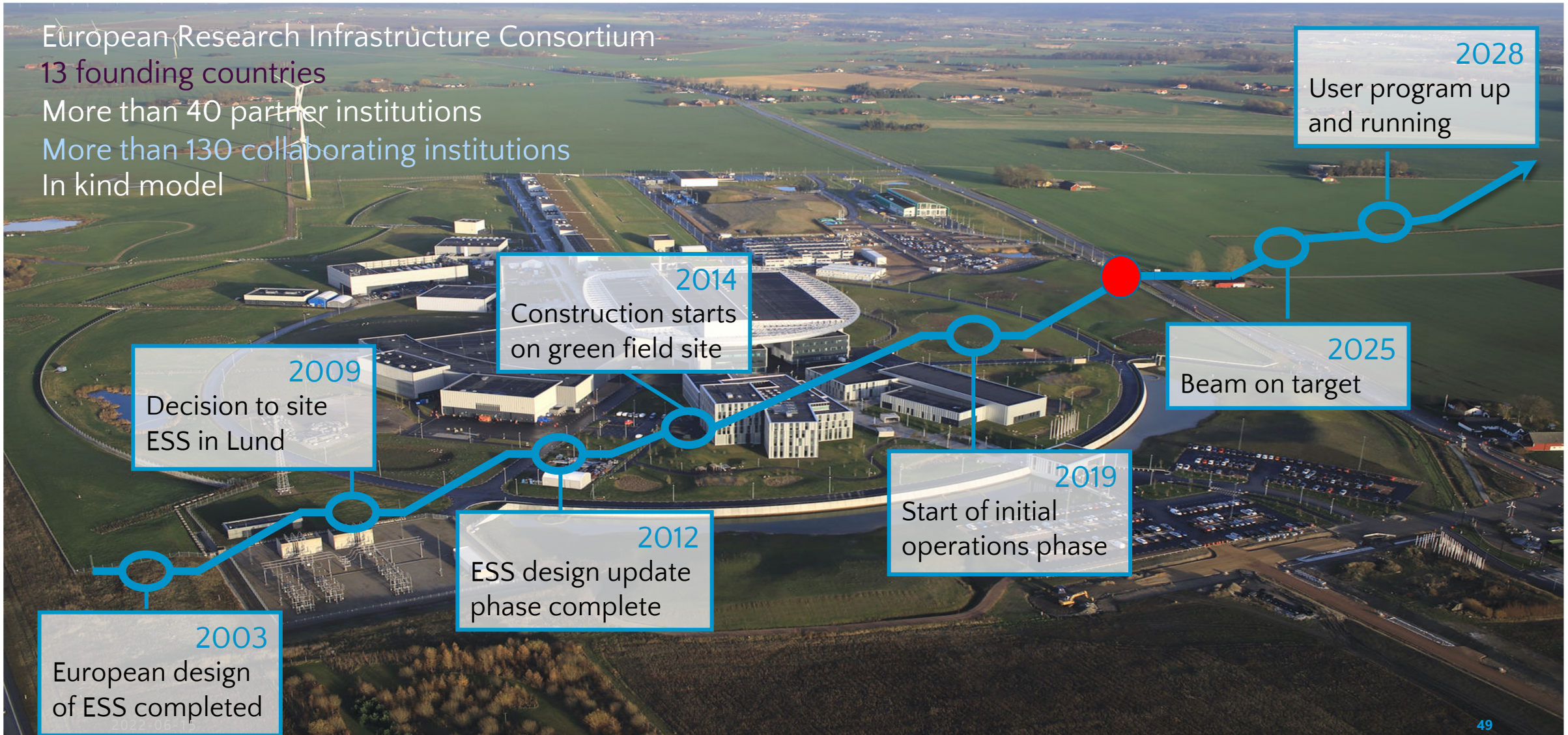


(Updated from *Neutron Scattering*, K. Skold and D. L. Price, eds., Academic Press, 1986)

# European Spallation Source



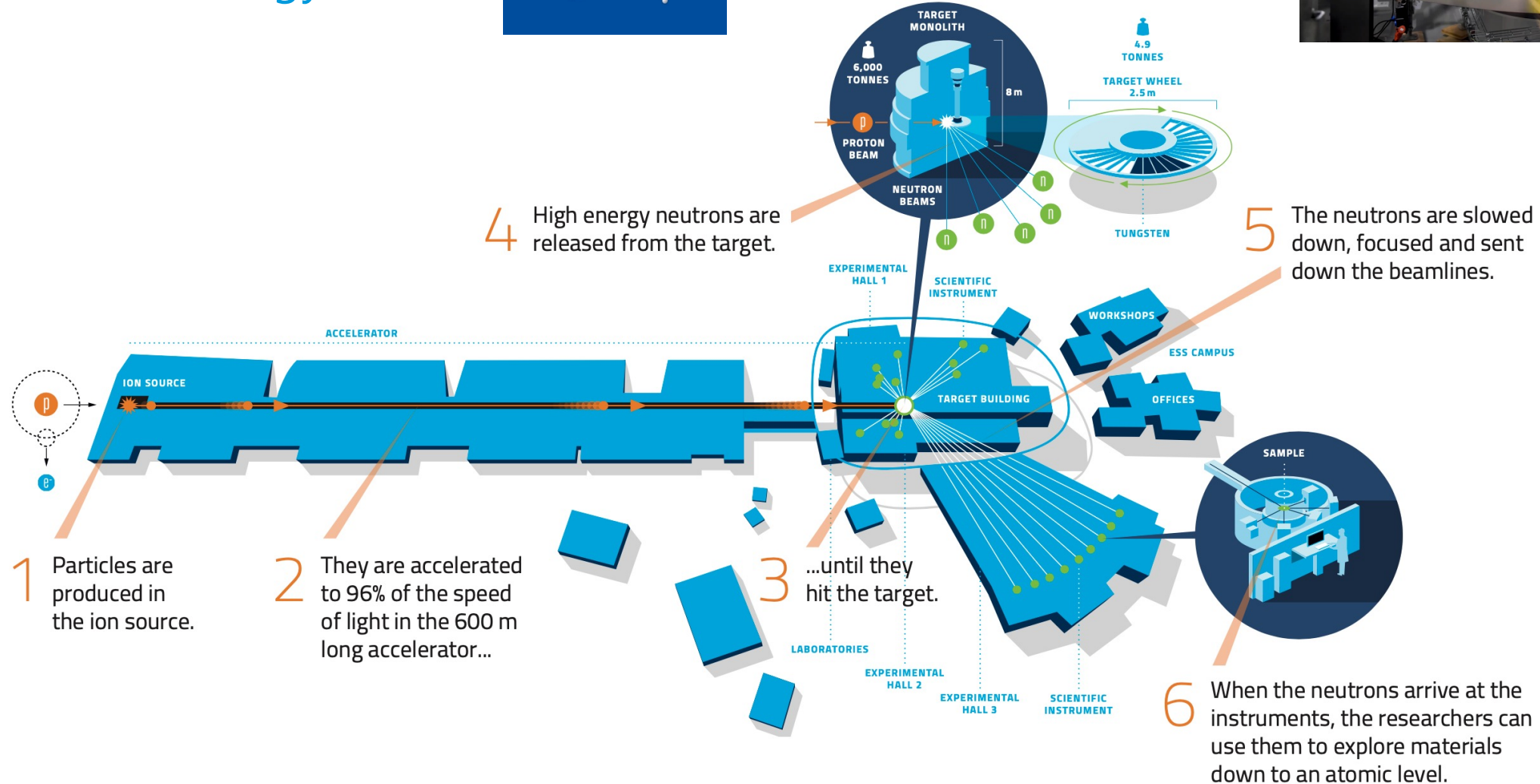
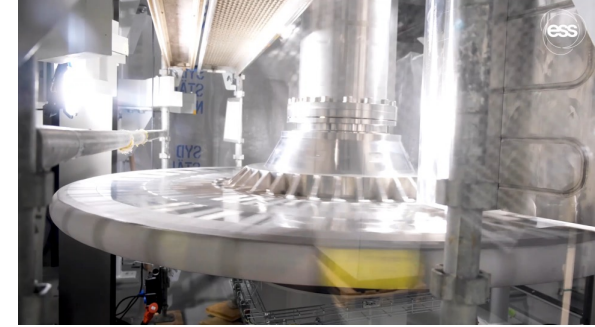
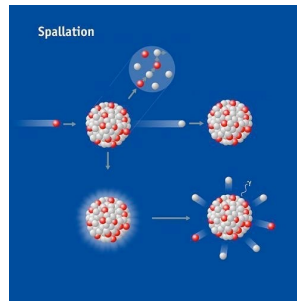
European Research Infrastructure Consortium  
13 founding countries  
More than 40 partner institutions  
More than 130 collaborating institutions  
In kind model





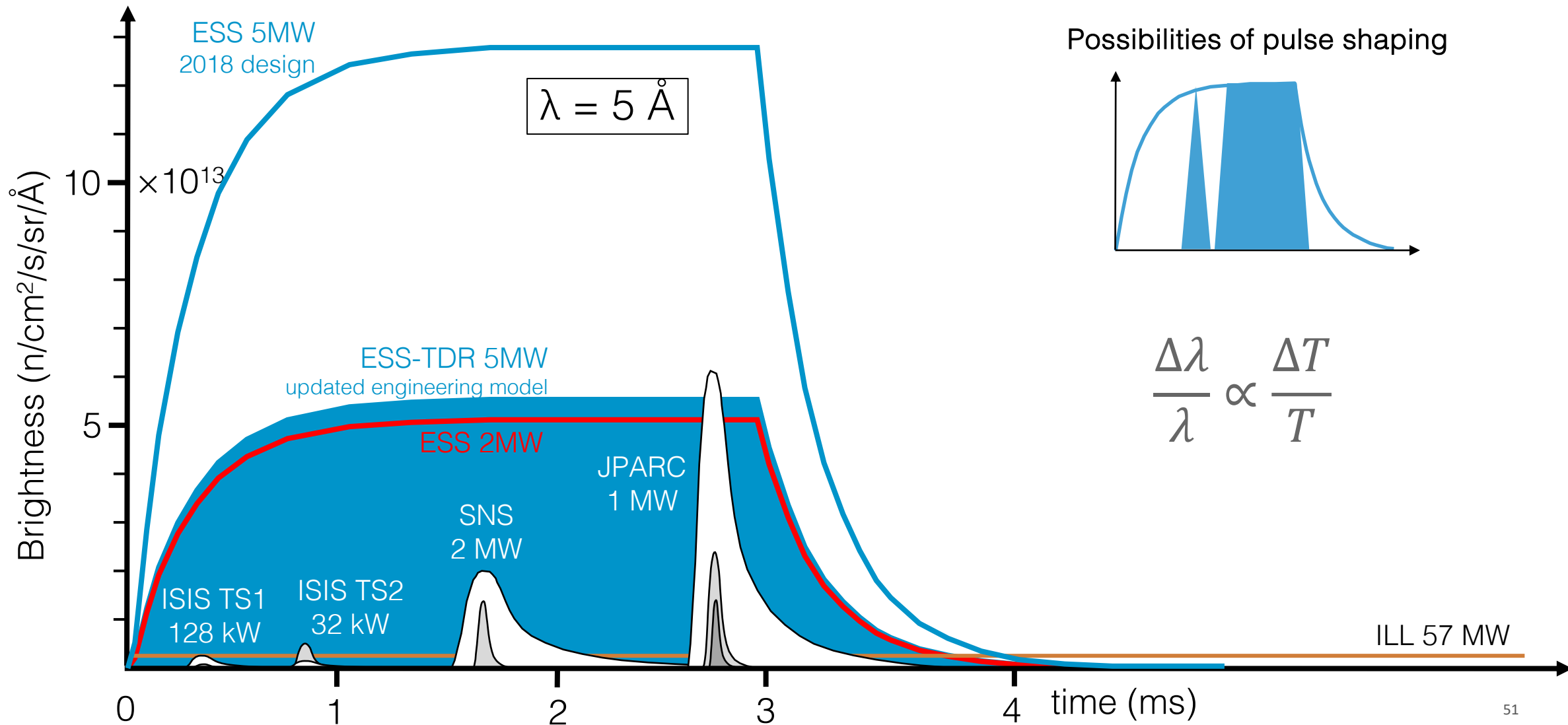
# How it works

## The technology

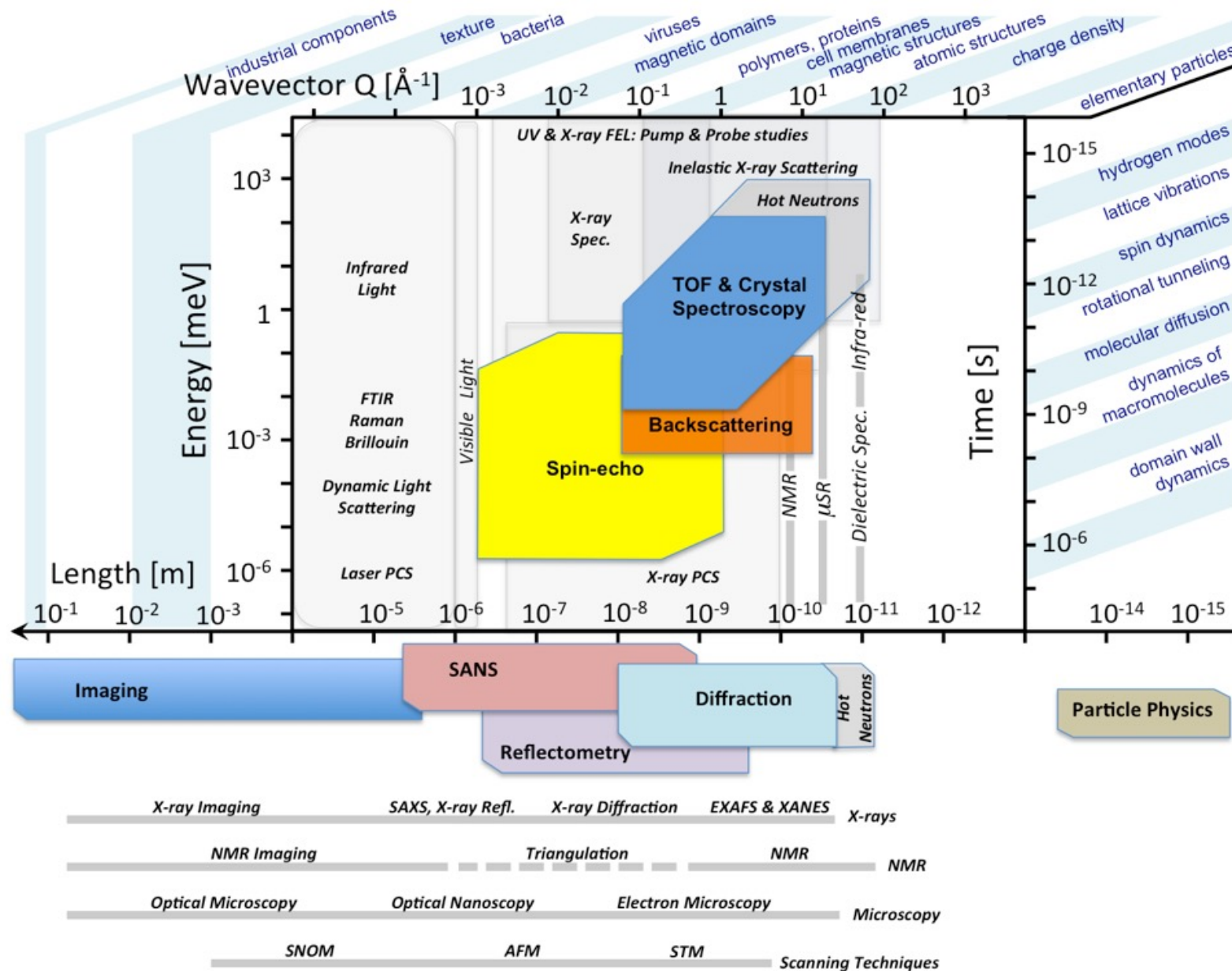




# Long-pulse Performance and Flexibility



# Neutron Scattering Techniques



# Neutron Instruments

Andersen, K. H.; Argyriou, D. N.; Jackson, A. J. et al. The Instrument Suite of the European Spallation Source. *Nuclear Instruments and Methods in Physics Research Section A*: **2020**, 957, 163402. <https://doi.org/10.1016/j.nima.2020.163402>.

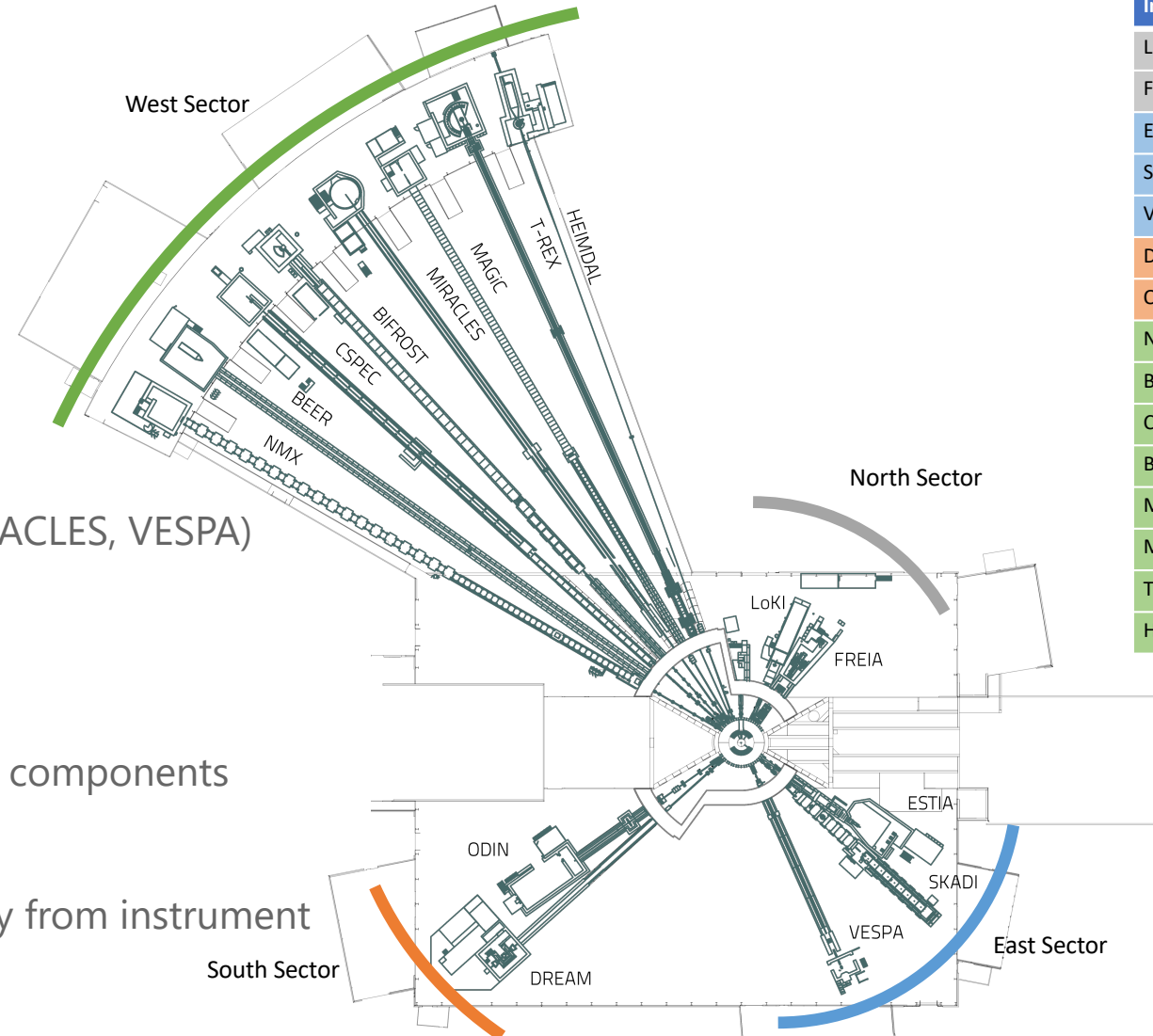


## 15 instruments + Test Beamline

- Diffractometers (DREAM, MAGiC, HEIMDAL)
- SANS (LoKI, SKADI)
- Reflectometers (Estia, FREIA)
- Imaging (ODIN)
- Engineering Diffraction (BEER)
- Macromolecular Crystallography (NMX)
- Spectrometers (CSPEC, T-REX, BIFROST, MIRACLES, VESPA)

*Novel detector technologies and geometries*  
*Complex pulse-shaping*

- Shared neutron bunker – common space for components
- Common timing system for facility
- Single controls infrastructure (EPICS)
- Control and data recording running remotely from instrument

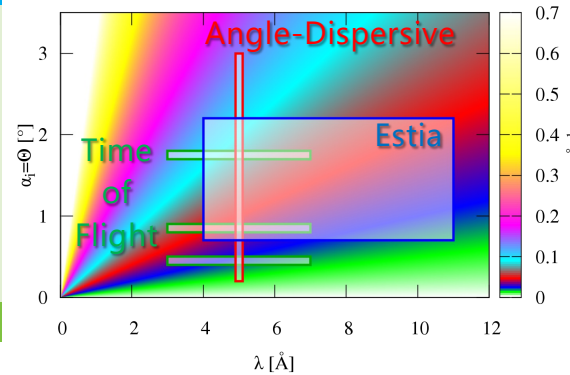
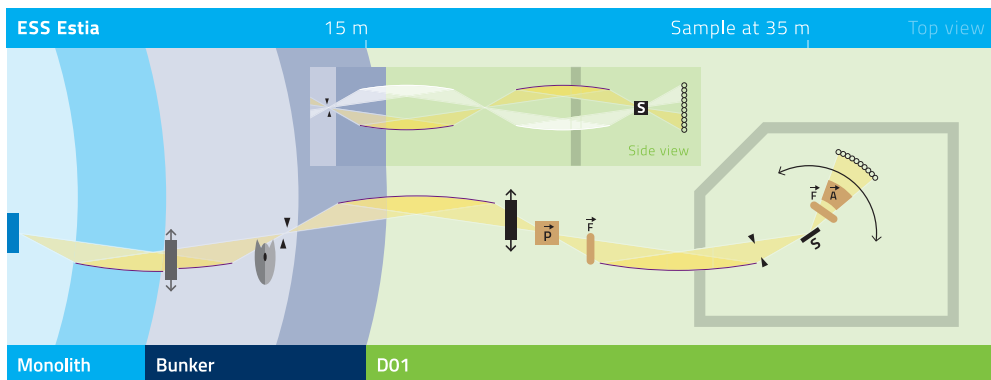


Instrument	Beamport
LoKI	N7
FREIA	N5
Estia	E2
SKADI	E3
VESPA	E7
DREAM	S3
ODIN	S2
NMX	W1
BEER	W2
CSPEC	W3
BIFROST	W4
MIRACLES	W5
MAGIC	W6
T-REX	W7
HEIMDAL	W8



# Estia

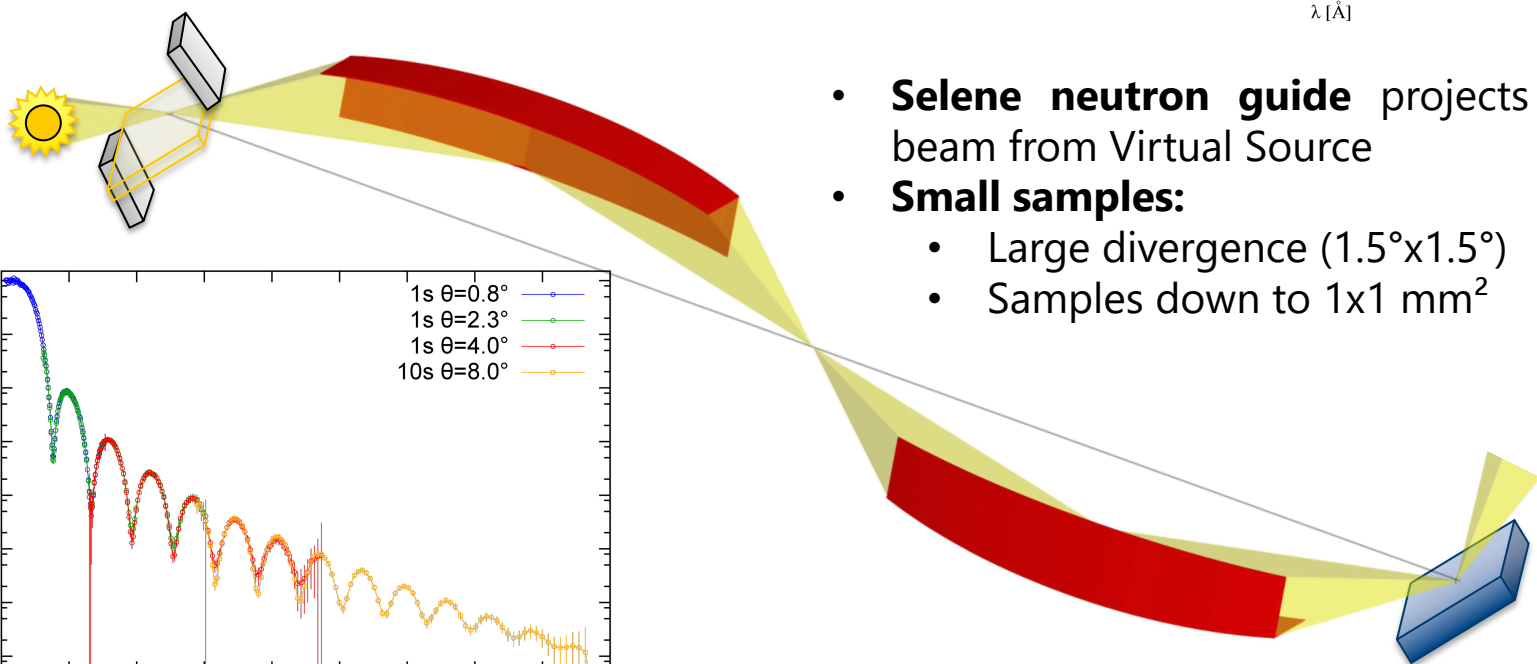
## Focussing Polarised Reflectometer for Tiny Samples



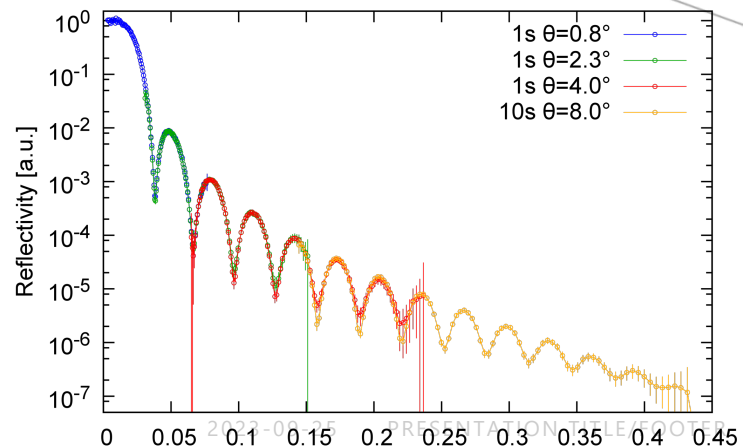
### Estia Quick Facts.

Estia Quick Facts	
Instrument Class	Reflectometry
Moderator	Cold
Primary Flightpath	35 m
Secondary Flightpath	4 m
Wavelength Range	3.75–28 Å
Polarised Incident Beam	Optional
Polarisation Analysis	Optional
Sample Orientation	Vertical
Total Q-Range	0.001 to 3.15 Å <sup>-1</sup> /–0.001 to –0.3 Å <sup>-1</sup>
Standard Mode (14 Hz)	
Bandwidth	7 Å
Flux at Sample at 2 MW <sup>a</sup>	6 × 10 <sup>8</sup> n s <sup>-1</sup> cm <sup>-2</sup>
Relative Q-Range	Q <sub>max</sub> = 2.85 × Q <sub>min</sub>
Q-Resolution ΔQ/Q	7.8%–3.0% over Q-range
2-Pulse Skipping Mode (4.7 Hz)	
Bandwidth	21 Å
Flux at Sample at 2 MW <sup>a</sup>	2 × 10 <sup>8</sup> n s <sup>-1</sup> cm <sup>-2</sup>
Relative Q-Range	Q <sub>max</sub> = 6.6 × Q <sub>min</sub>
Q-Resolution ΔQ/Q	7.8%–1.3% over Q-range

<sup>a</sup>Full-divergence beam averaged over 5(H) × 10(V) mm<sup>2</sup>.



- **Selene neutron guide** projects tiny beam from Virtual Source
- **Small samples:**
  - Large divergence (1.5°x1.5°)
  - Samples down to 1x1 mm<sup>2</sup>

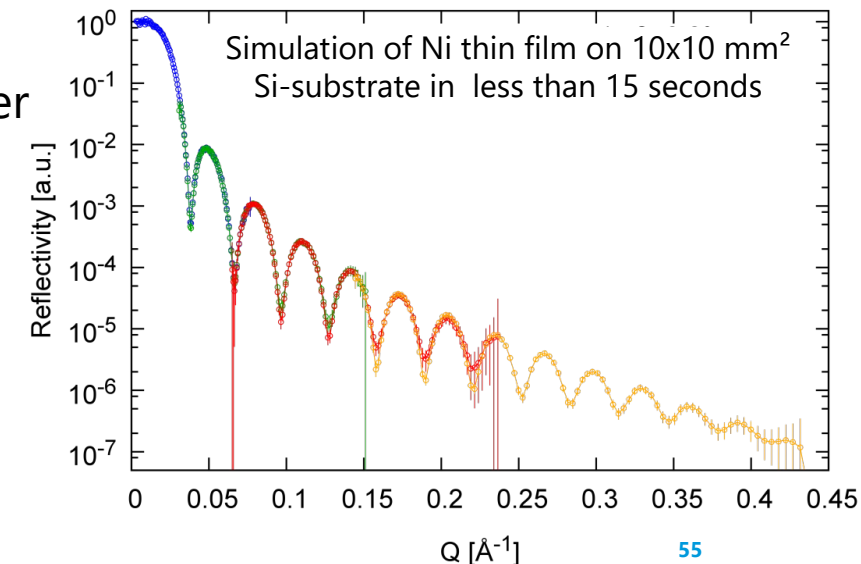


For the study of surfaces and interfaces including magnetic layers

ESTIA is optimised for small samples and polarisation analysis:

The investigation of the **chemical and magnetic** depth-profile near surfaces and of **lateral correlations and structures**

- functional devices: spin-valves, spintronics
- diffusion processes: Li batteries, corrosion protection
- multifunctional materials: interface-coupled electric and magnetic properties
- towards real materials: raster-scanning of bent, faceted or multi-domain surfaces
- **Small samples:**
  - Large divergence ( $1.5^\circ \times 1.5^\circ$ )
  - Samples down to  $1 \times 1 \text{ mm}^2$
- **Polarization >99%** for curved transmission polarizer and analyser
- Simultaneous measurement of two polarization states

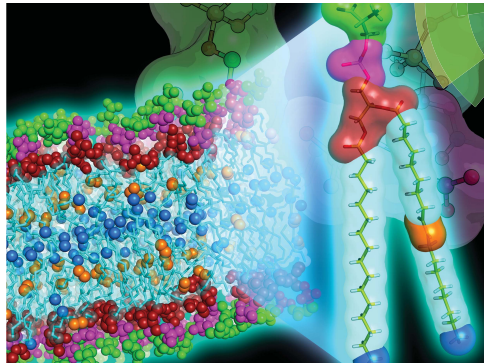


# FREIA

## Science Case



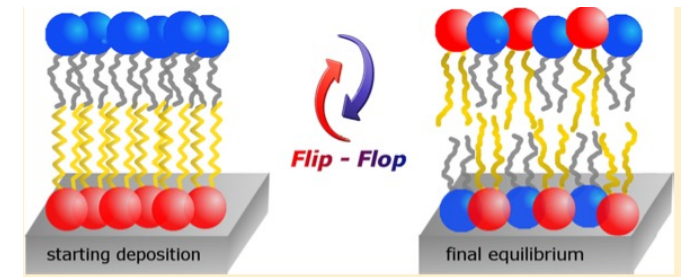
In-situ time-resolved reflectometry for soft condensed matter, life science and functional materials



Instrument characteristics to allow very fast measurements:

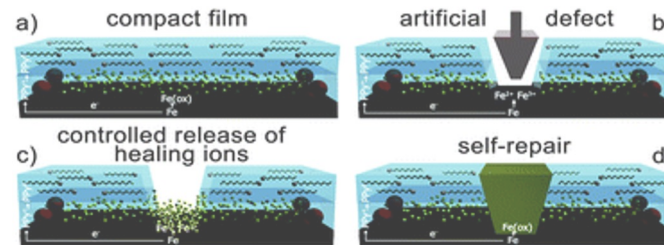
- Very high flux
- Horizontal sample geometry
- Flexible collimation
- Variable resolution
- Broad simultaneous Q
- No sample movement

## Dynamics



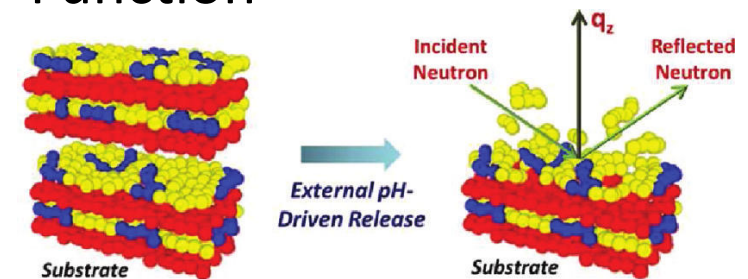
- deposition, structure and phase behavior
- adsorption, self-assembly and reactions
- gas/liquid/solid interfaces

## Applications



- response to external stimuli
- in situ and in operando
- complex sample environments

## Function



## Design





# Expected performances of ESS reflectometers compared to ILL ones

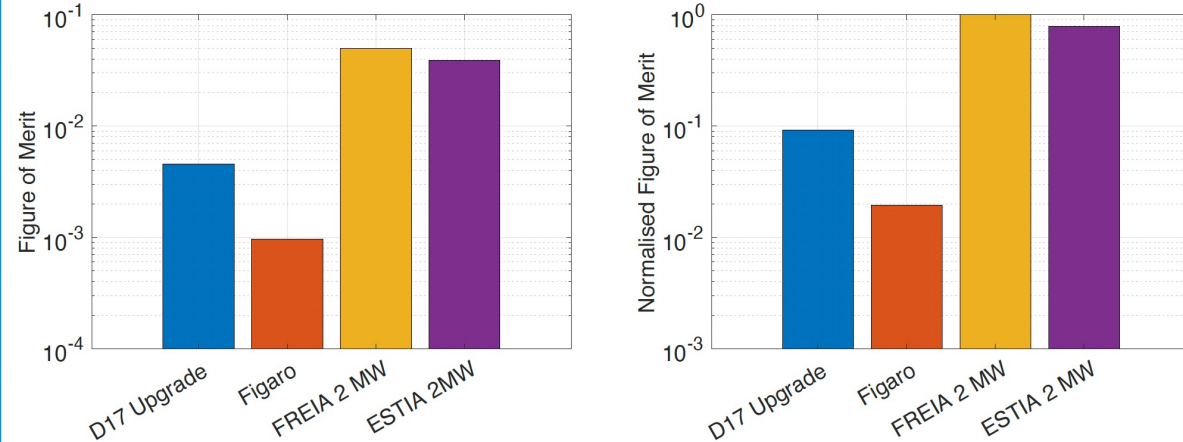


$$\text{FOM} = \text{Peak brightness} * \text{divergence (vertical \& horizontal)} * \lambda_{\text{max}}/\lambda_{\text{min}}$$

FREIA and ESTIA: wide divergence mode: large q-range kinetic measurements

FOM = Peak brightness\* divergence (vertical & horizontal) \*  $\lambda_{\text{max}}/\lambda_{\text{min}}$

ESS-2MW: D17 new guide: usable flux on the sample by a factor of 2.5

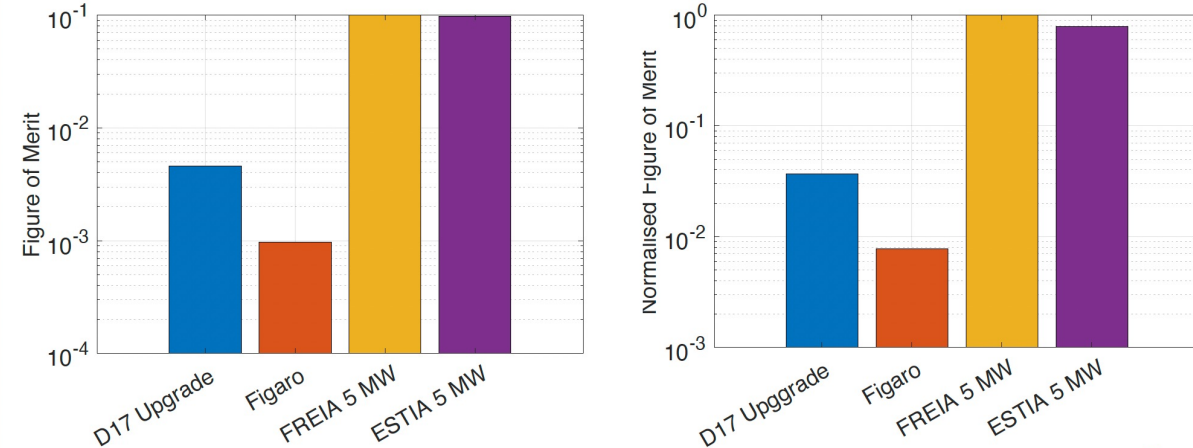


2MW

FREIA and ESTIA: wide divergence mode: large q-range kinetic measurements

FOM = Peak brightness\* divergence (vertical & horizontal) \*  $\lambda_{\text{max}}/\lambda_{\text{min}}$

ESS-5MW: D17 new guide: usable flux on the sample by a factor of 2.5



5MW



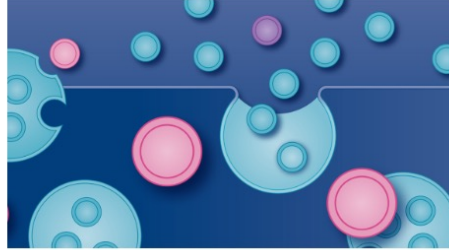
# Challenges for neutron science

[nature](#) > collection

Collection | 17 February 2023

## Extracellular vesicles

Extracellular vesicles (EVs) have emerged as important means of cell–cell communication, having the potential to transfer various cargoes – encompassing proteins, nucleic acids, metabolites or even entire organelles – between cells. By now, the importance of EV-mediated cell–cell communication has been documented in a plethora of physiological and pathological situations, across the different kingdoms. In addition, their secretion and cargo composition can change depending on the biological context, making EVs suitable biomarkers for several diseases. EVs have also been harnessed as drug delivery agents and standalone therapeutics.

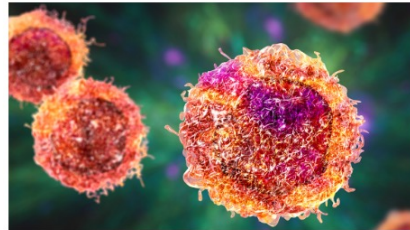


[nature](#) > collection

Collection | 14 April 2023

## Cancer research

Cancer is a leading cause of death, accounting for nearly one in six deaths worldwide. Many cancers can be cured, especially if detected early and treated effectively. Nevertheless, an unmet need for the development of treatments for aggressive and often metastatic tumors remains. Preclinical and clinical research in the areas of cancer screening and detection, as well as development of new therapies are at the core of this challenge. This development is cemented by an understanding of basic cancer biology and tumor immunology and tumor profiling studies that link bench and bedside to allow for an improved understanding of therapy response and resistance. In this collection, we highlight the breadth of cancer research in these areas at the Nature Portfolio.



Article | [Open Access](#) | [Published: 14 April 2023](#)

## Temporal nanofluid environments induce prebiotic condensation in water

[Andrea Greiner de Herrera](#), [Thomas Markert](#) & [Frank Trixler](#) ✉

[Communications Chemistry](#) 6, Article number: 69 (2023) | [Cite this article](#)

468 Accesses | 2 Altmetric | [Metrics](#)

2023-09-25

COMMENT | 02 May 2023

## Address the growing urgency of fungal disease in crops

More political and public awareness of the plight of the world’s crops when it comes to fungal disease is crucial to stave off a major threat to global food security.

## Bacterial cellulose comes out of the woodwork

Polymer scientists in Japan are harnessing the power of botany and bacteria to produce bioplastics that don’t harm the environment.

NATURE INDEX | 14 December 2022

## Three ways to combat antimicrobial resistance

With a dearth of new antibiotics coming to market, researchers are finding creative ways to keep bacteria at bay.

NEWS FEATURE | 04 April 2023

## Conquering Alzheimer’s: a look at the therapies of the future

Researchers are looking to drug combinations, vaccines and gene therapy as they forge the next generation of treatments for the condition.

E.g. what ESS could advance (with smaller samples/higher throughput/deuteriation):

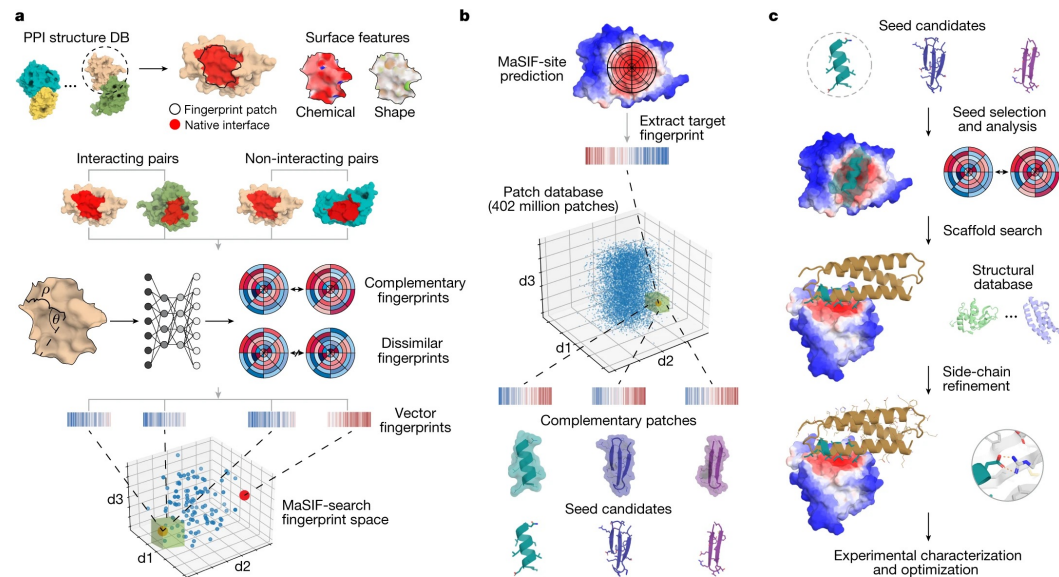


# Challenges for neutron science

RESEARCH BRIEFINGS | 26 April 2023

## New protein–protein interactions designed by a computer

Creating protein interactions through computational design is a key challenge in the fields of both basic and translational biology. An approach that uses the machine-learned fingerprints of protein-surface features was used to produce synthetic proteins that engage immunotherapeutic or viral targets with binding affinities comparable to those of naturally occurring proteins.



This is a summary of: [Gainza, P. et al. De novo design of protein interactions with learned surface fingerprints. Nature https://doi.org/10.1038/s41586-023-05993-x \(2023\).](https://doi.org/10.1038/s41586-023-05993-x)

nature > articles > article

Article | Published: 05 April 2023

## mRNA recognition and packaging by the human transcription–export complex

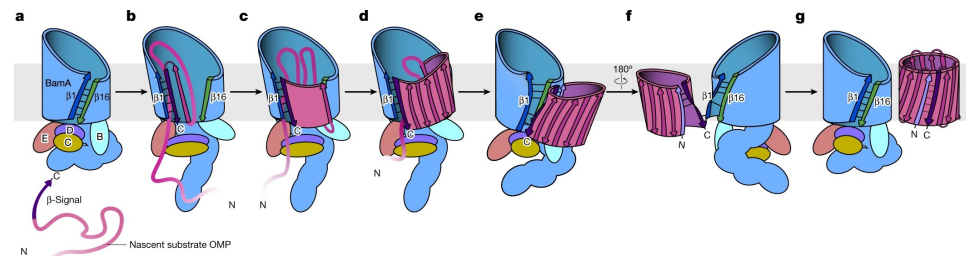
[Belén Pacheco-Fiallos](#), [Matthias K. Vorländer](#), [Daria Riabov-Bassat](#), [Laura Fin](#), [Francis J. O'Reilly](#), [Farja I. Ayala](#), [Ulla Schellhaas](#), [Juri Rappsilber](#) & [Clemens Plaschka](#) ✉

*Nature* **616**, 828–835 (2023) | [Cite this article](#)

RESEARCH BRIEFINGS | 26 April 2023

## Step-by-step assembly of a $\beta$ -barrel protein in a bacterial membrane

Gram-negative bacteria that are resistant to multiple drugs cannot survive without the cell-surface machinery that builds a  $\beta$ -barrel pore structure from outer membrane proteins. Snapshots of different stages in the assembly process provide insights into this crucial mechanism, and could lead to the development of new antibiotics.



This is a summary of: [Shen, C. et al. Structural basis of BAM-mediated outer membrane  \$\beta\$ -barrel protein assembly. Nature https://doi.org/10.1038/s41586-023-05988-8 \(2023\).](https://doi.org/10.1038/s41586-023-05988-8)



# Acknowledgements

## Colleagues at ILL and ESS



LSS group at ILL outing 2022

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Sebastain Kölher  
Giacomo Corucci  
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Rachel Morrison  
Ida Berts  
Jonathan Talbot  
Emanuel Schneck

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Marco Maccarini  
Francesca Baldelli Bombelli  
Debora Berti  
Giuseppe Vitiello  
Luigi Paduano  
Valerie Laux  
Michael Haertlein  
Jayne Lawrence  
Tommy Nylander  
Jian Ren Lu  
Michele Sferrazza  
Beate Kloesgen

...



NEUTRONS  
FOR SCIENCE



PARTNERSHIP FOR SOFT CONDENSED MATTER



EUROPEAN  
SPALLATION  
SOURCE

