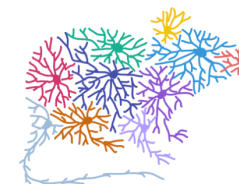
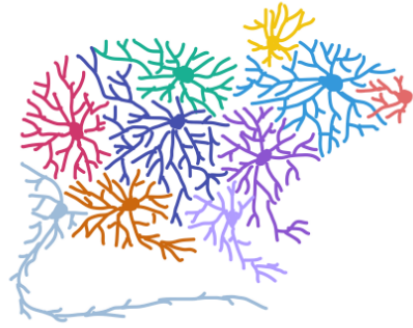


# 2023 A3D3 Workshop Hackathon DadarlatLab Dataset

Seungbin Park	<a href="mailto:park1377@purdue.edu">park1377@purdue.edu</a>
Megan Lipton	<a href="mailto:liptonm@purdue.edu">liptonm@purdue.edu</a>
Maria Dadarlat	<a href="mailto:mdadarla@purdue.edu">mdadarla@purdue.edu</a>



**DADARLAT  
LAB**



# DADARLAT LAB

<https://engineering.purdue.edu/DadarlatLab>

The Dadarlat Lab has two main areas of study:

(1) to understand how sensory information is processed in the visual and somatosensory systems

(2) to apply these insights towards the development of artificial sensation.

To address these questions, we use a combination of animal behavior, in-vivo electrophysiology, 2-photon-imaging, and electrical stimulation.



Dr. Maria  
Dadarlat-Makin



Megan Lipton



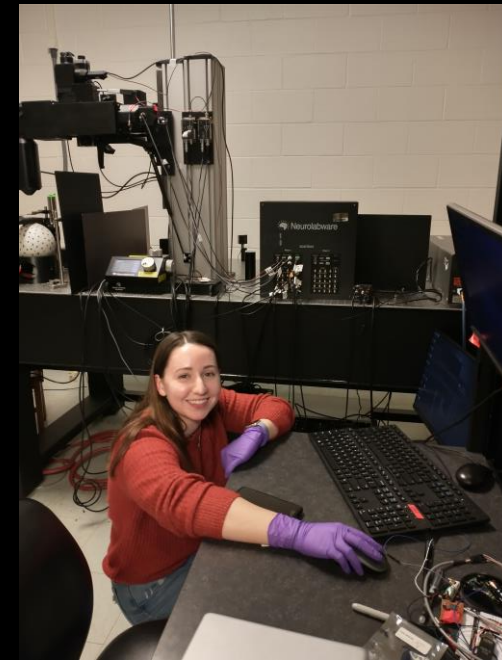
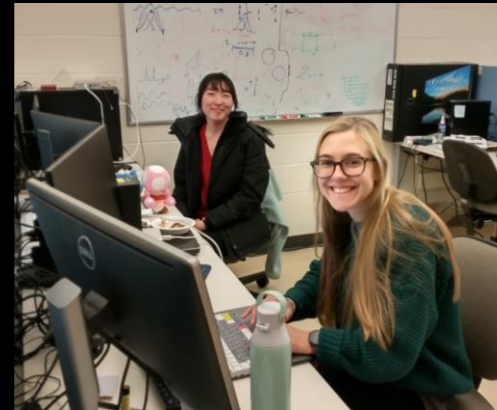
Seungbin Park



Elizabeth Frazier

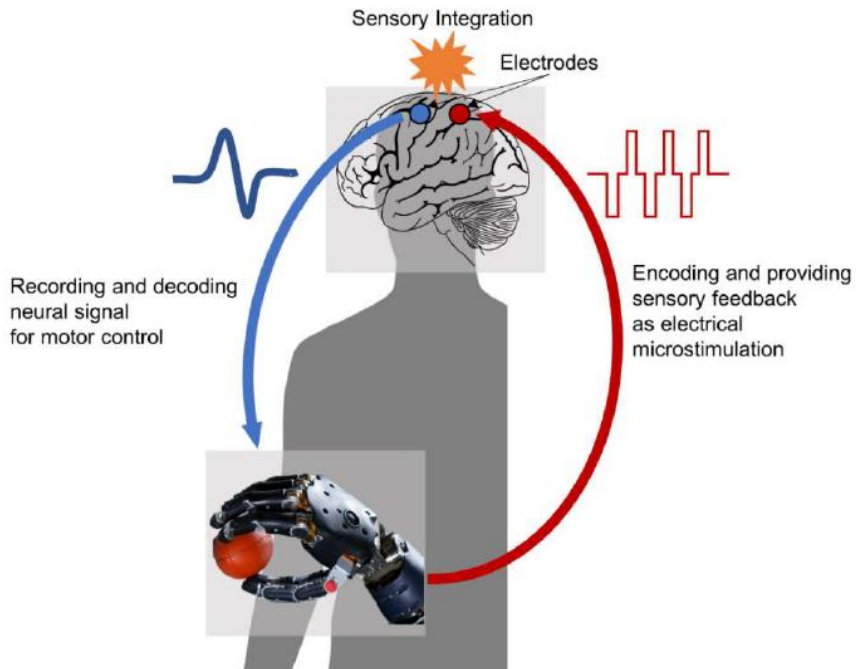
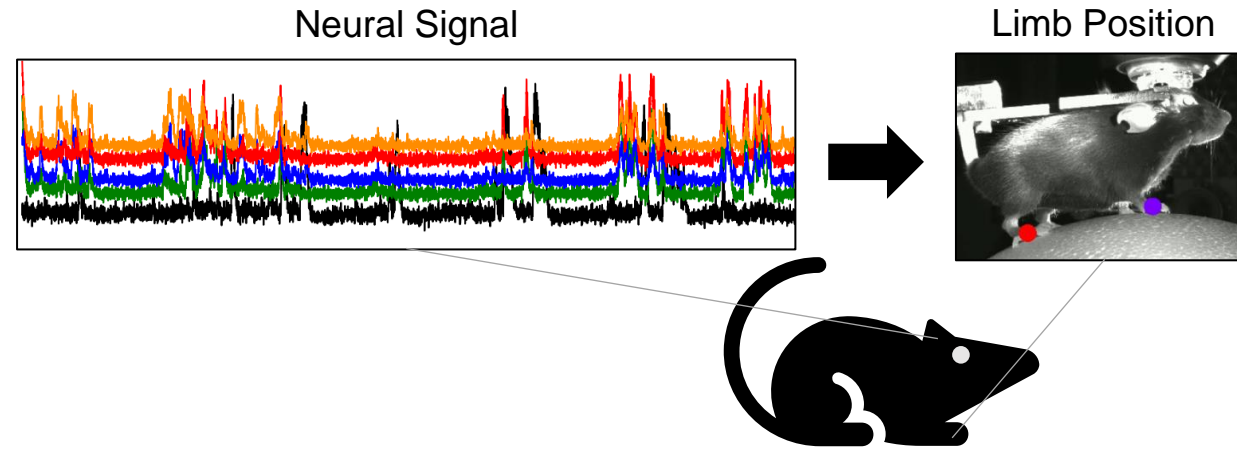


Samuel Senneka

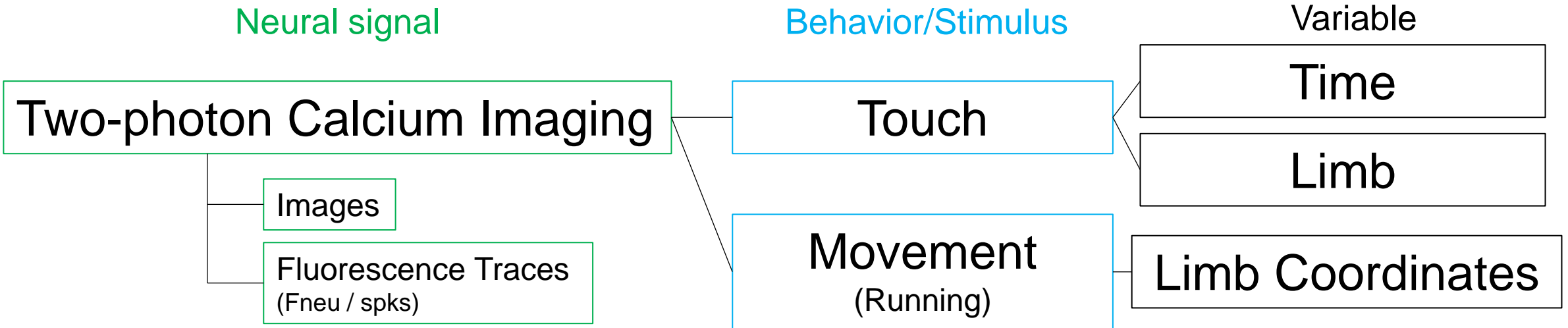
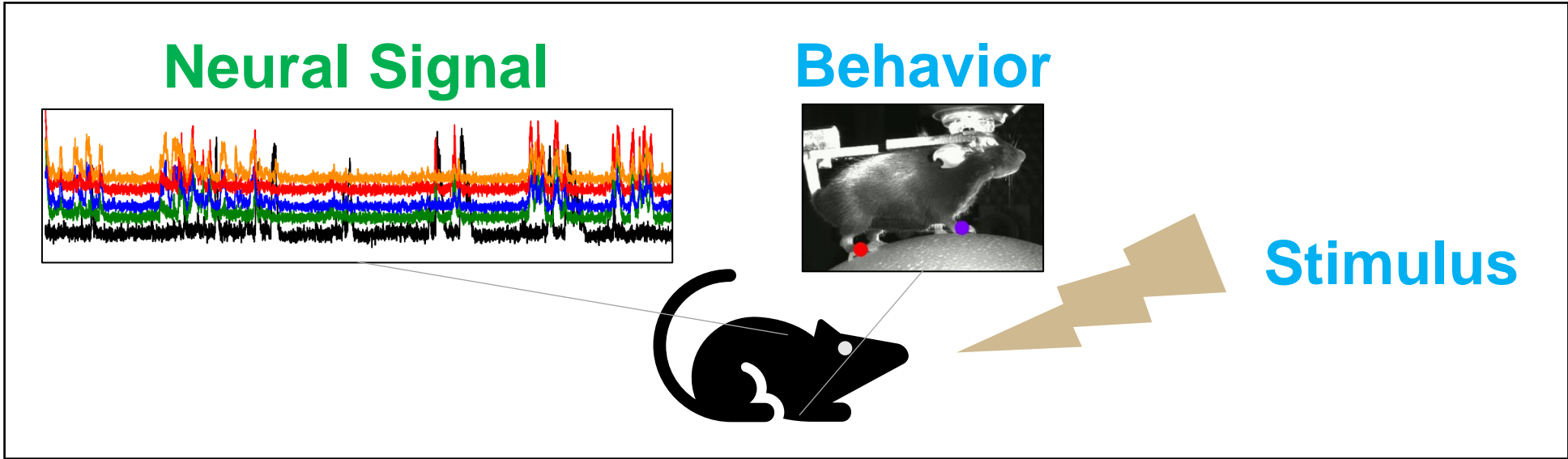


# Neural Decoding

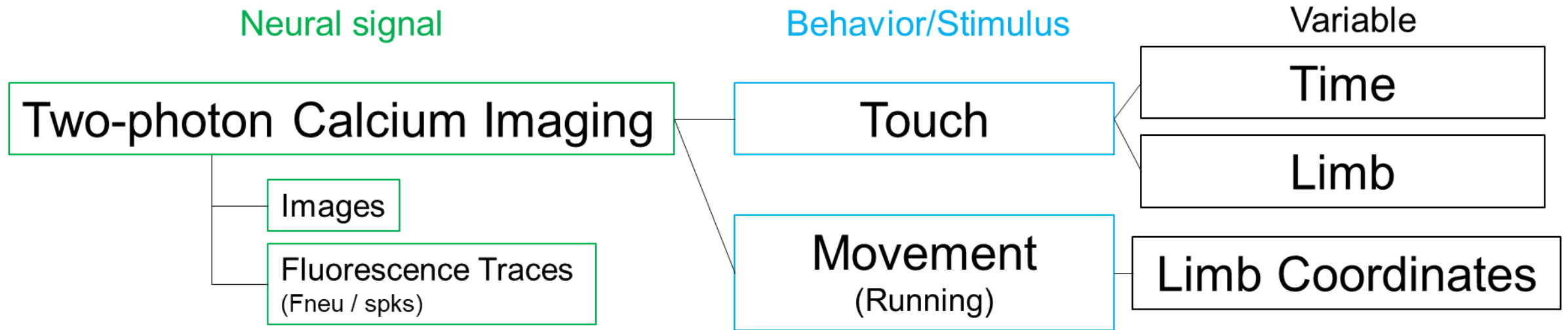
: Neural Signal → Behavior



- (1) To develop technology to aid patients suffering from neurological injury and disease  
ex) Brain-machine interface, Artificial sensation by electrical stimulation
- (2) To understand the function of the brain



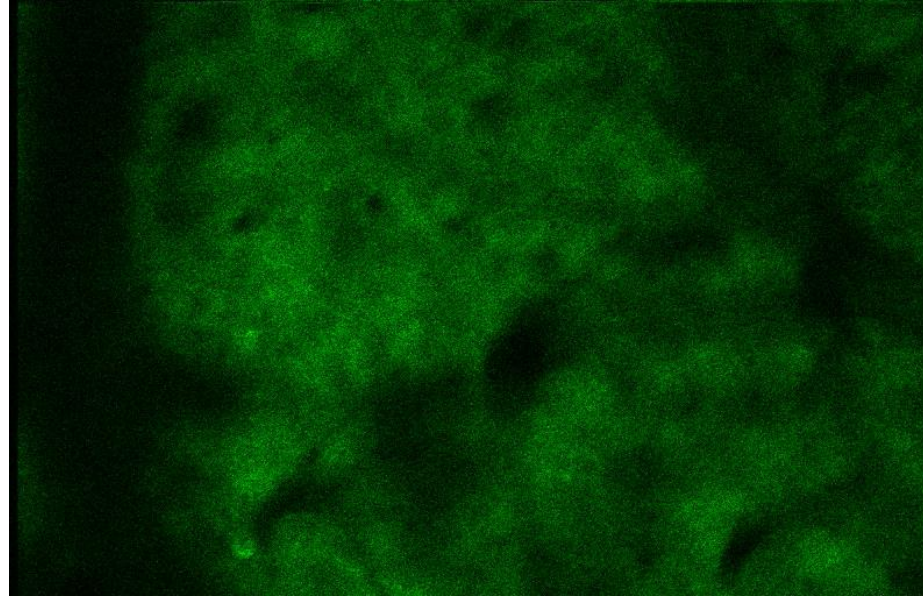
# Possible Hackathon Projects



- **Main goal: To explore the datasets**
- 2P+Touch
  - (Classification) Decoding the time when the touch stimulus was given from 2P calcium images
  - (Classification) Decoding the limb in which the touch stimulus was given from 2P calcium images
- 2P+Running
  - (Regression) Decoding continuous limb trajectories of a running mouse from 2P calcium images
- Reconstruction
- Anything you want

# Two-Photon Calcium Imaging

: a promising technique for recording large populations of neurons



When neurons fire action potentials, intracellular calcium levels rise. Two-Photon calcium imaging measures neural activity by detecting fluorescence from fluorescent molecules that bind to calcium.

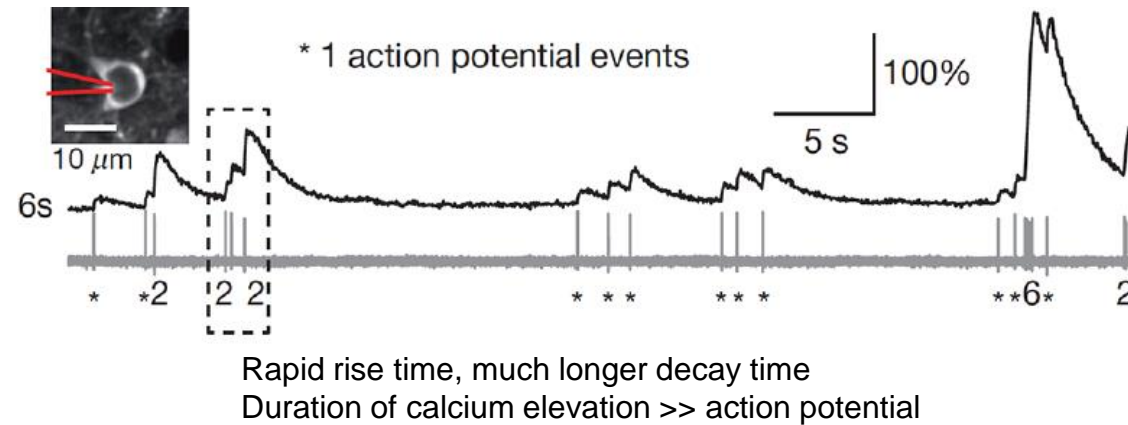
# Challenges in Decoding Two-Photon Calcium Imaging

- (1) Indirect and non-linear relation to action potentials
- (2) Slow kinetics with a long decay time
- (3) Time variance between neural activities in a frame
- (4) Low sampling rate



# Challenges in Decoding Two-Photon Calcium Imaging

- (1) Indirect and non-linear relation to action potentials
- (2) Slow kinetics with a long decay time
- (3) Time variance between neural activities in a frame
- (4) Low sampling rate

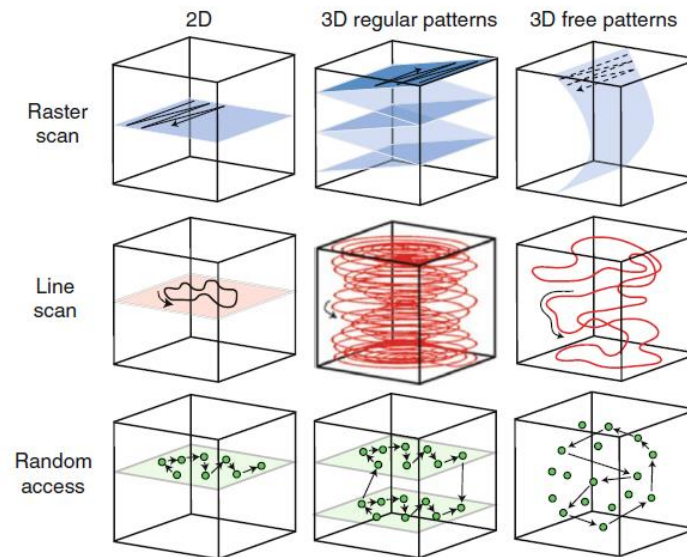


Ali et al., 2020



# Challenges in Decoding Two-Photon Calcium Imaging

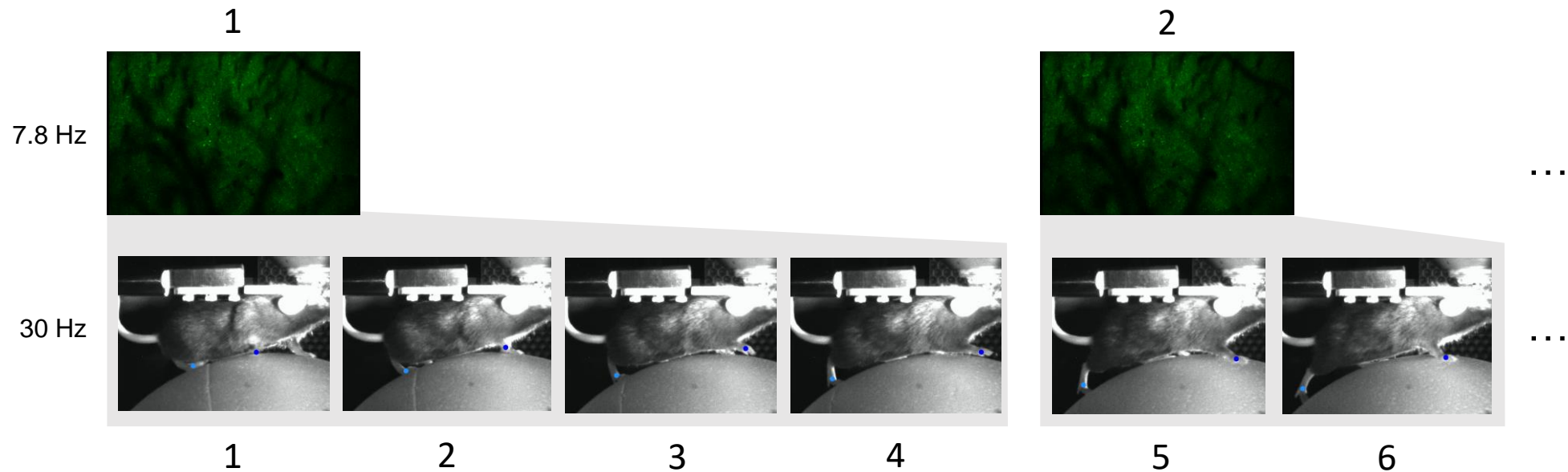
- (1) Indirect and non-linear relation to action potentials
- (2) Slow kinetics with a long decay time
- (3) Time variance between neural activities in a frame
- (4) Low sampling rate



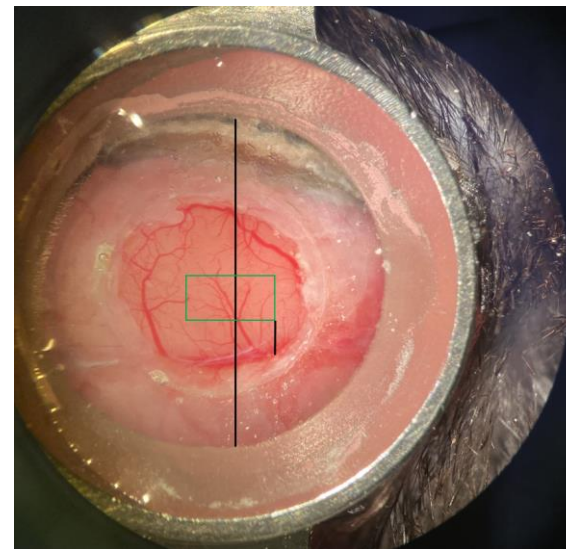
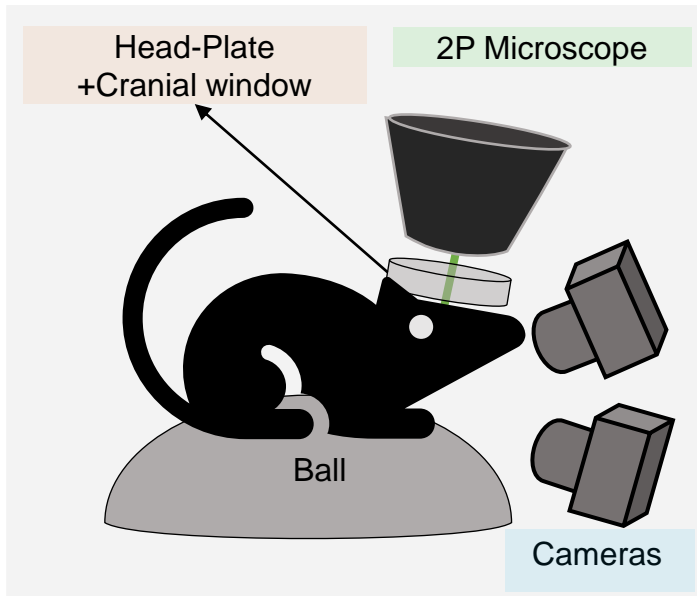
Weber et al., 2014

# Challenges in Decoding Two-Photon Calcium Imaging

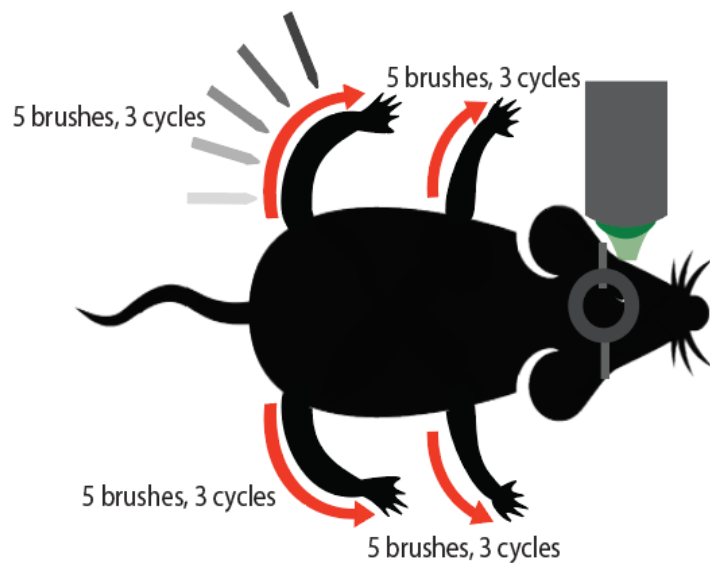
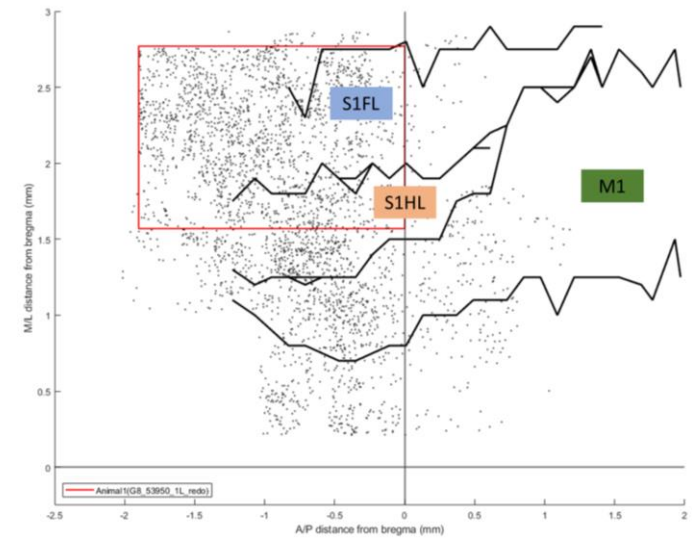
- (1) Indirect and non-linear relation to action potentials
- (2) Slow kinetics with a long decay time
- (3) Time variance between neural activities in a frame
- (4) Low sampling rate



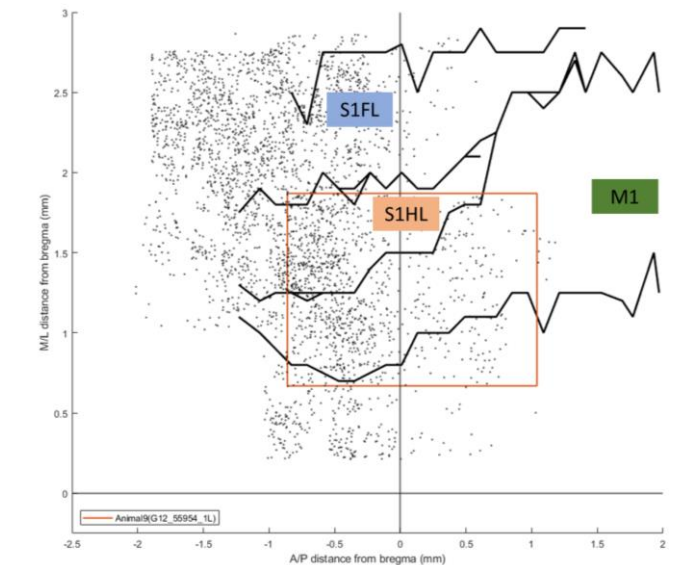
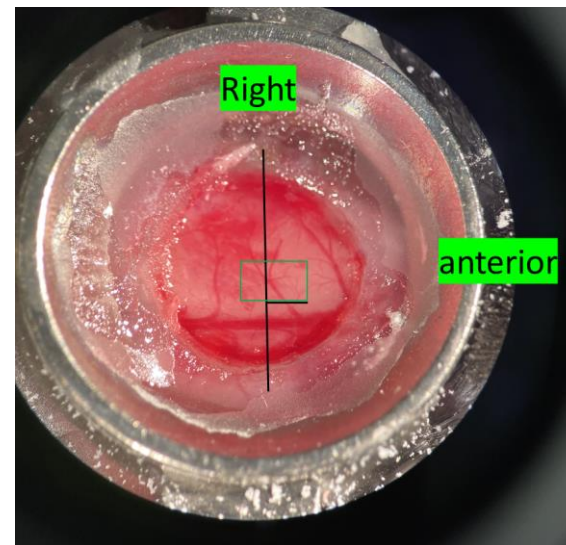
# Experiments



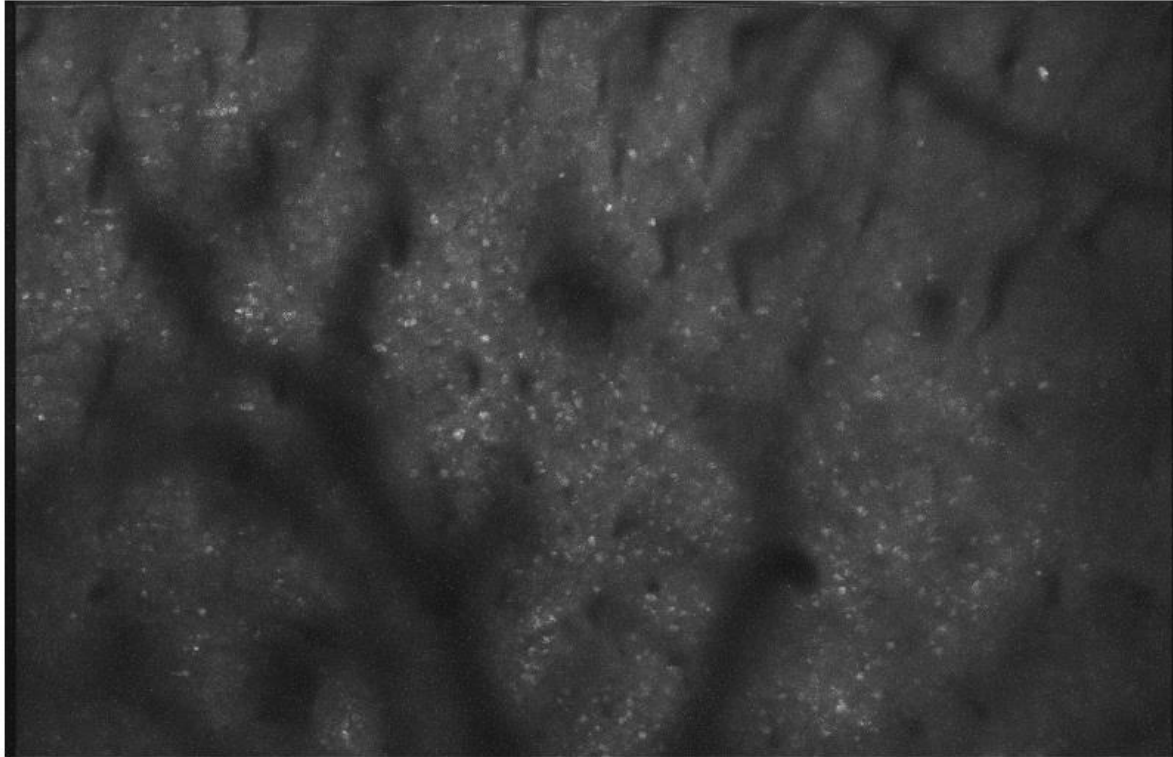
Animal 1



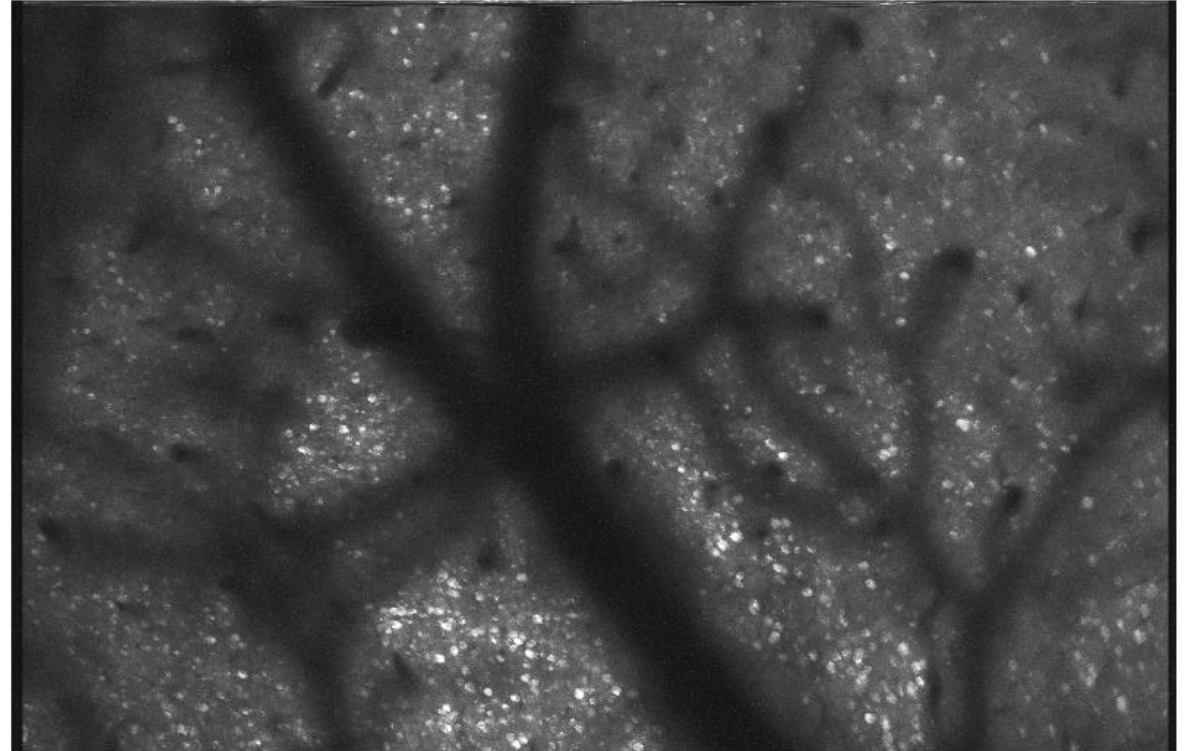
Animal 2



Animal 1



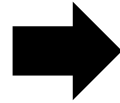
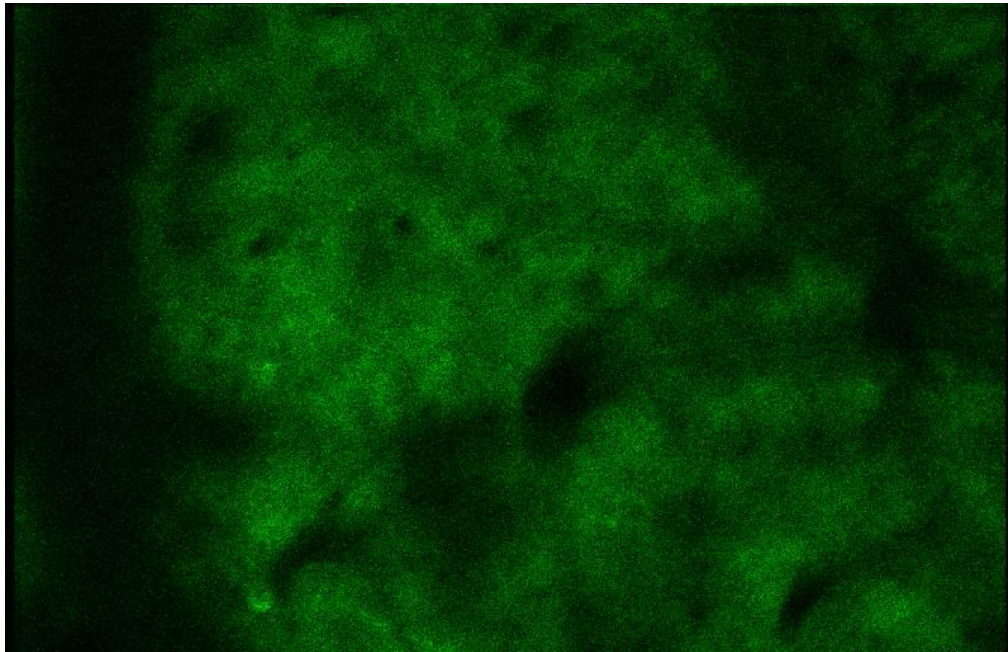
Animal 2



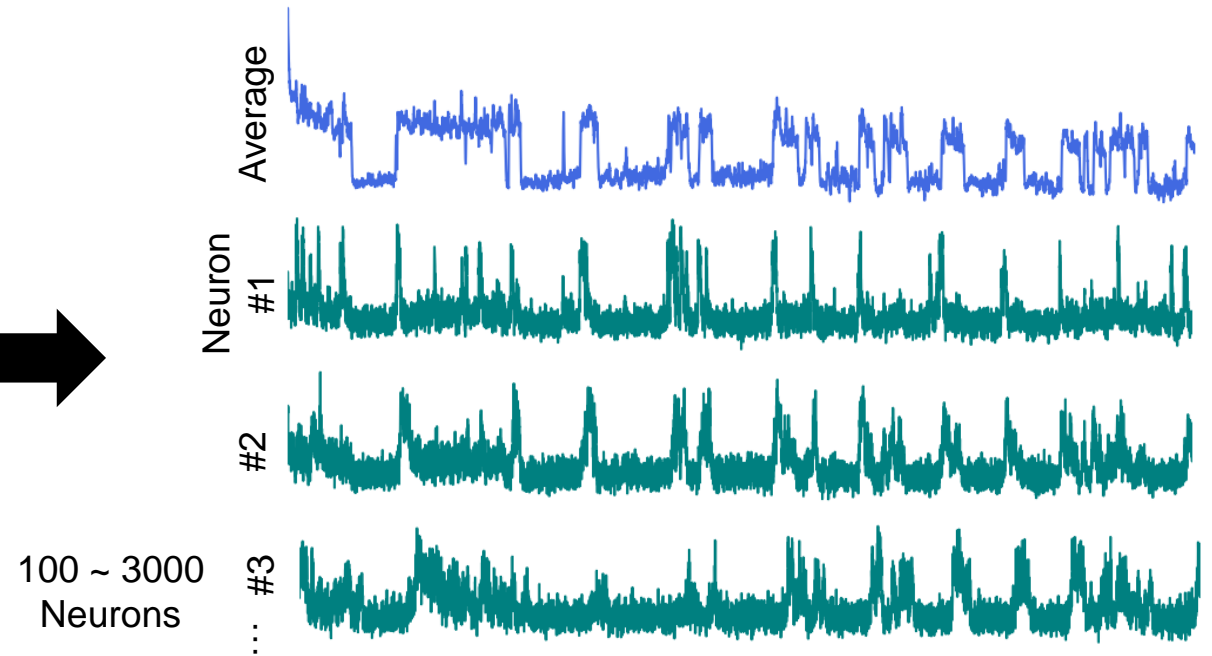


# Fluorescence Traces

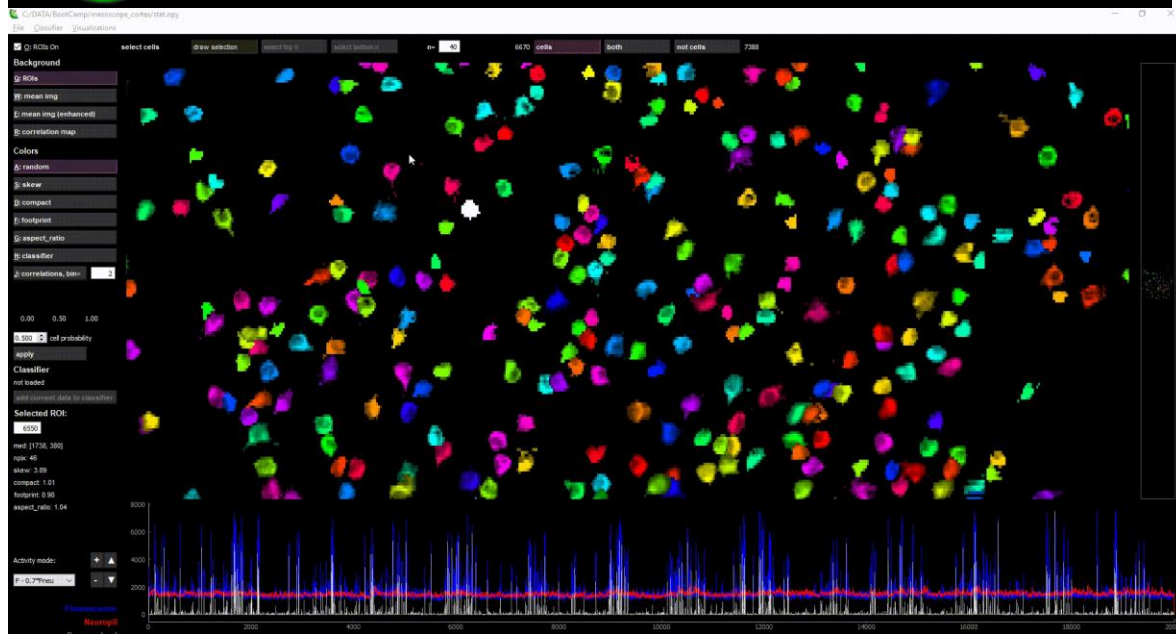
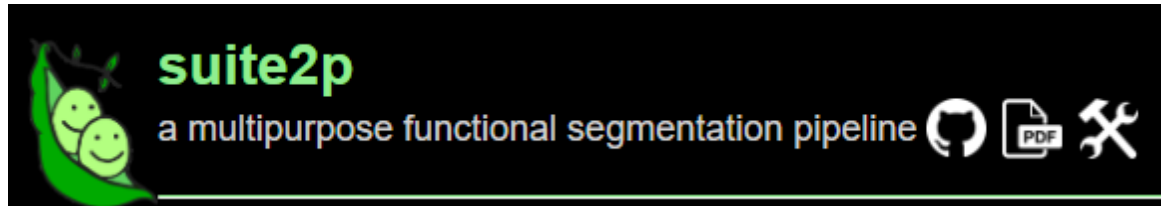
## 2P Calcium Imaging



## Fluorescence Traces



- Neurolabware
- Layers: L2/3
- Field of view: 1.9 mm x 1.2 mm; Laser: 920 nm
- Frame rate: 7.8 Hz
- Preprocessing: suite2p, Normalization



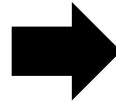
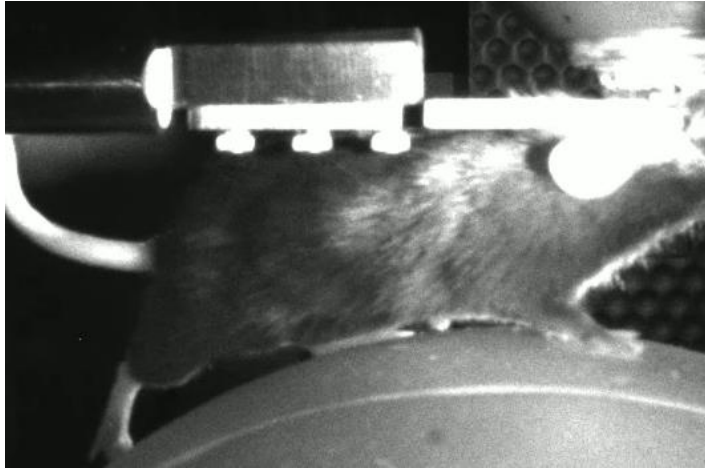
Registration  
Cell Detection  
Signal extraction  
Spike deconvolution

<https://www.suite2p.org/>

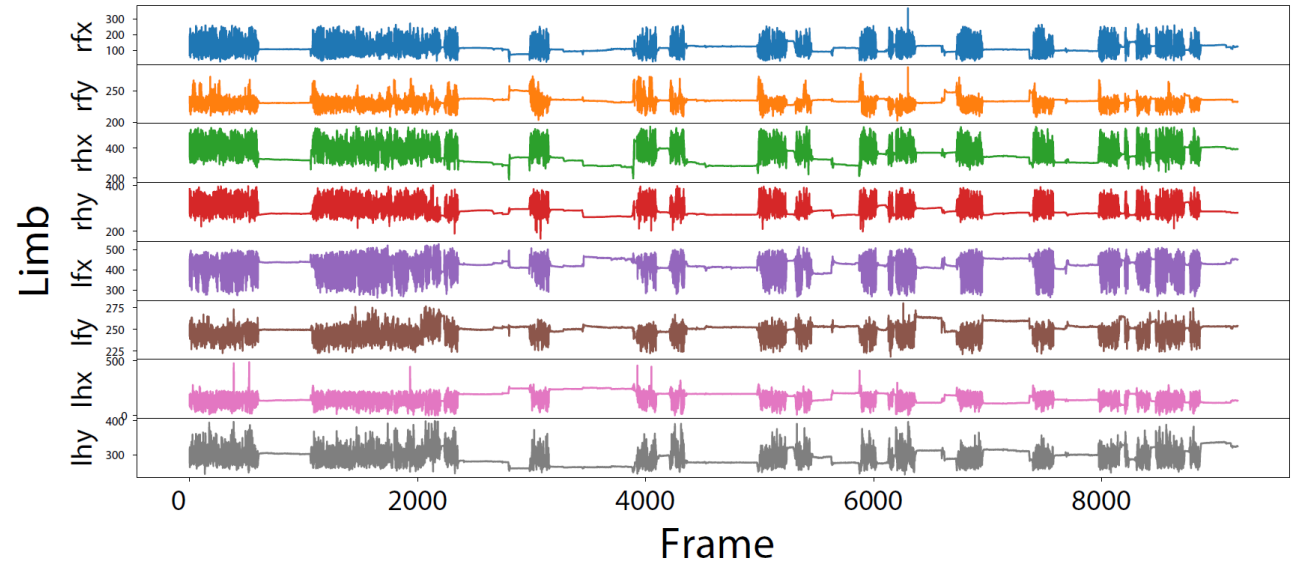
Doc on Outputs:<https://suite2p.readthedocs.io/en/latest/outputs.html>

# Limb Coordinates from a Running Mouse

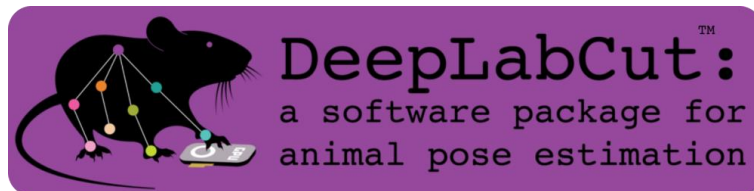
## Behavior Video



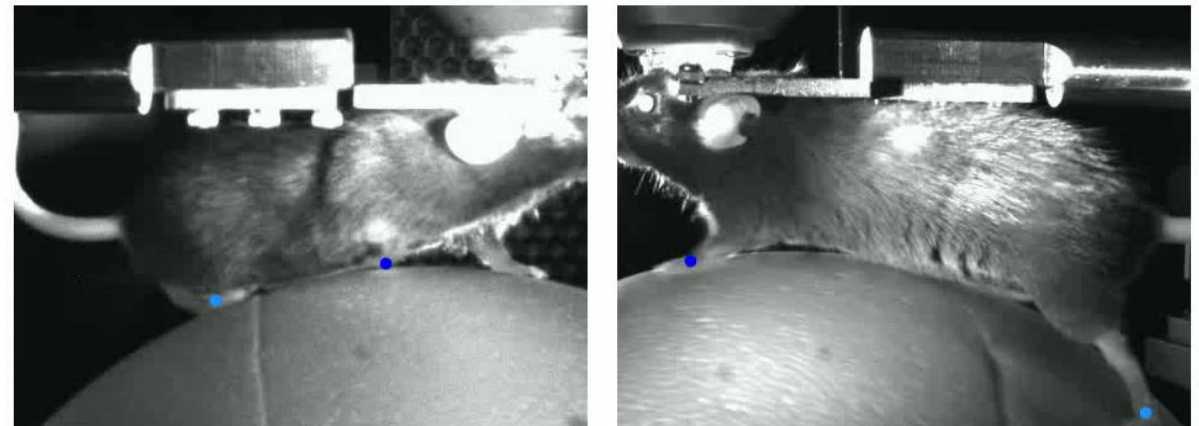
## Limb Coordinates



- Freely running on the ball.
- Trained for 4 days, 1 hour per day.
- Frame rate: 30 Hz
- Preprocessing: Deeplabcut, Normalization

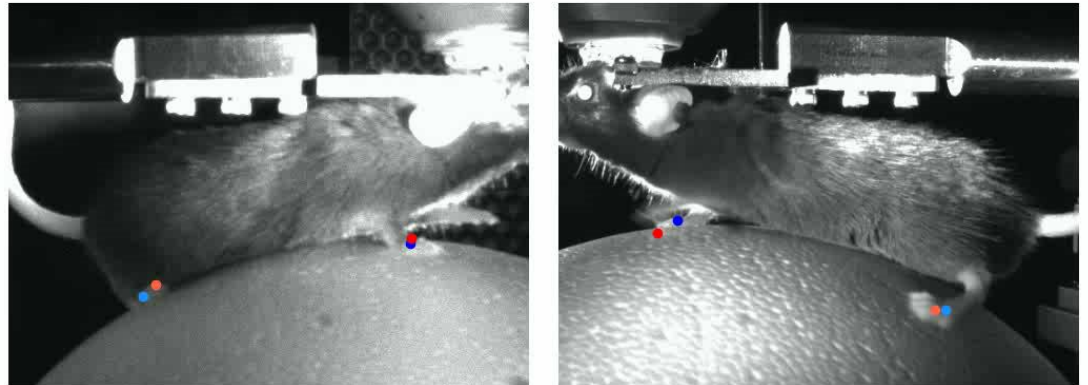
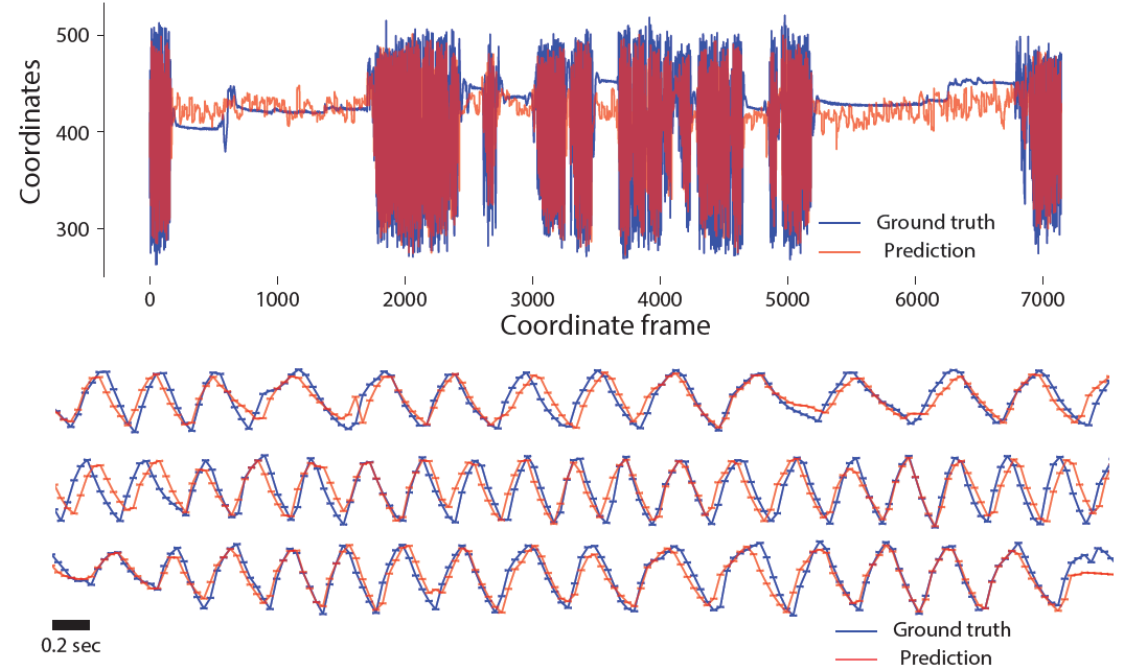
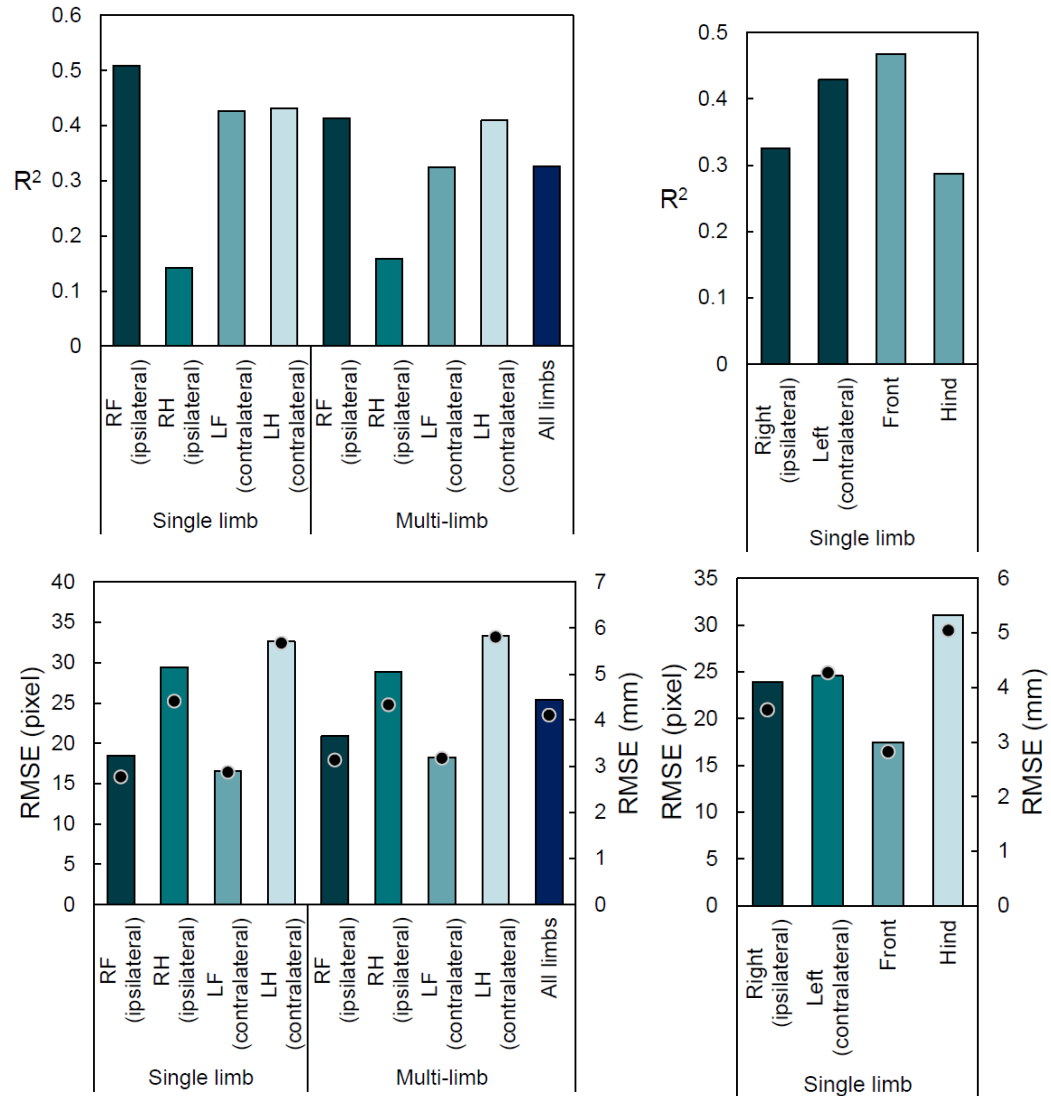


<http://www.mackenziemathislab.org/deeplabcut>





# All limbs(contralateral/ipsilateral, front/hind) could be decoded both simultaneously and separately from a single cortical hemisphere.



Animal 1, Multi-limb:  $R^2=0.326$ , RMSE=25.35 (4.11 mm)

# Examples of Open Datasets

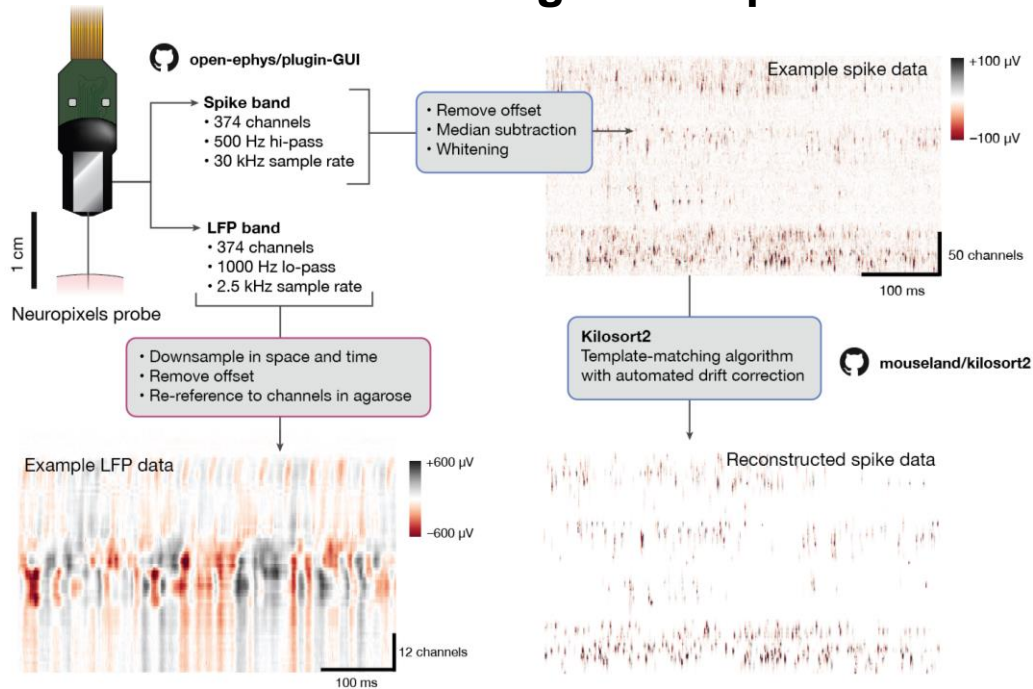
Allen Brain Map

<https://portal.brain-map.org/>

Neurodata Without Borders

<https://www.nwb.org/example-datasets/>

## Visual Coding - Neuropixels

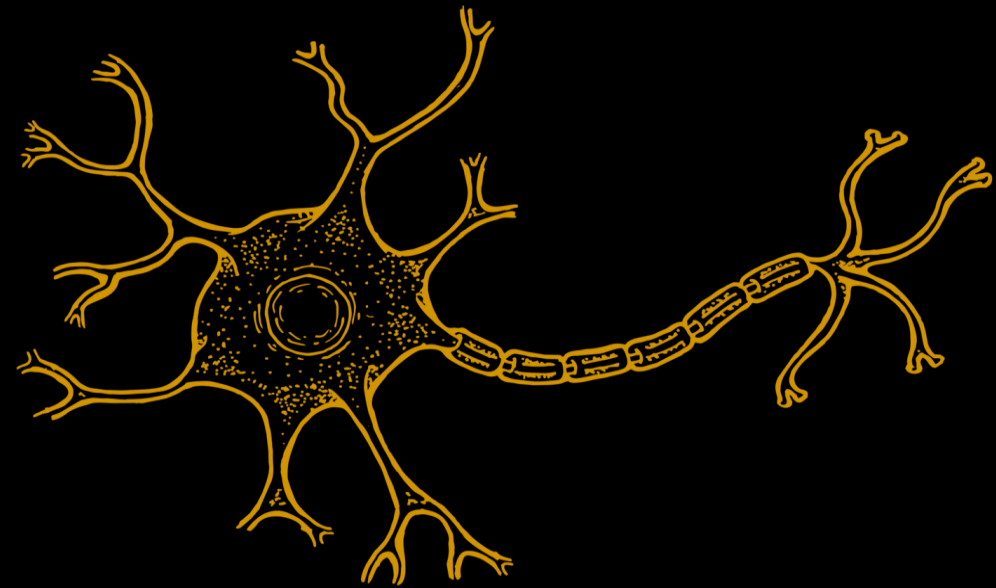


[https://allensdk.readthedocs.io/en/latest/visual\\_coding\\_neuropixels.html](https://allensdk.readthedocs.io/en/latest/visual_coding_neuropixels.html)

## Visual Behavior – 2P



<https://portal.brain-map.org/explore/circuits/visual-behavior-2p>



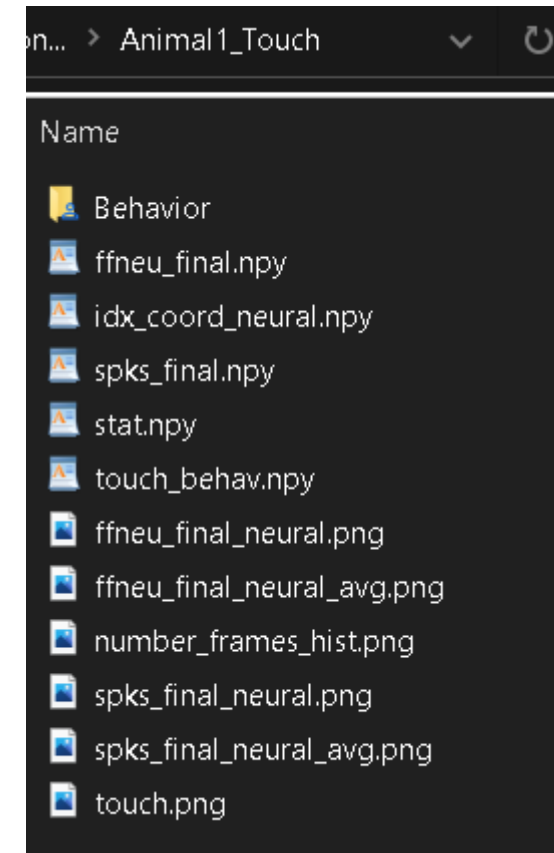
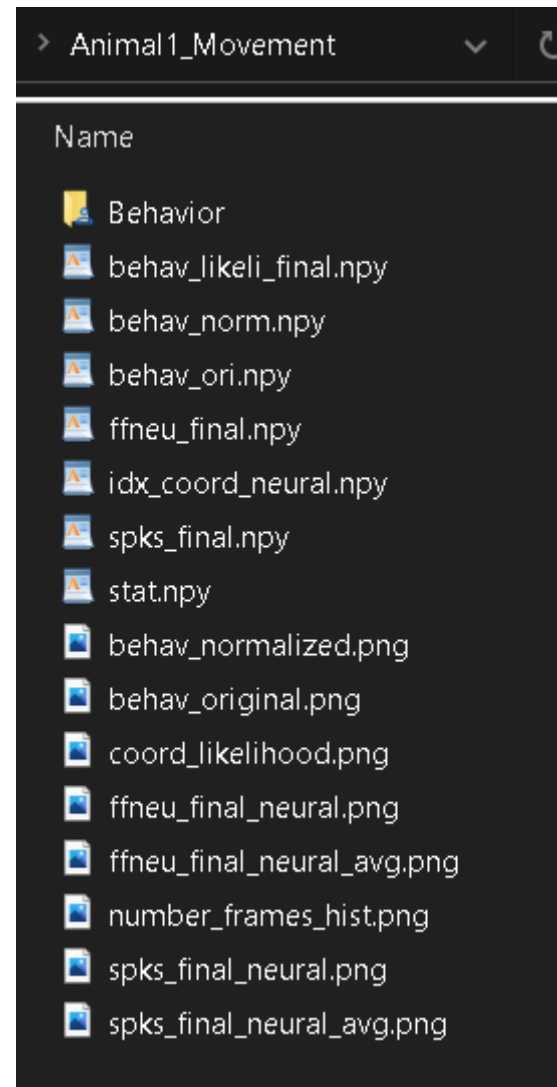
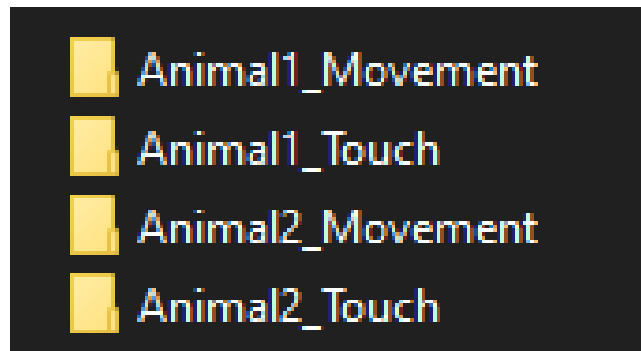
Documentation



These datasets are not validated yet.  
We are currently working on them.

# Guidelines:

- All raw data are available.
- Considering the limited time, preprocessed data is being provided.  
It was preprocessed and synchronized, so you can just start to use them right away.
- Choose one behavior between touch (classification) and movement (regression).  
Recommend trying classification first because it is simpler.
- Try Animal 1 first. And validate more with Animal 2 if Animal 1 worked.
  
- Research ideas
  - It is usual to use deconvolved fluorescence traces.  
But related studies suggest using raw 2P microscope images or fluorescence traces that are not deconvolved as future plans, because they might include more information.
  - If you want to try to use raw 2P microscope images or unprocessed original fluorescence traces, please feel free to go explore and have fun.
  - Consider the characteristics of the datasets – 2P images and behaviors.  
ex) Latency, GNN using spatial information, Denoising ...
  - LFADs on 2P: <https://www.nature.com/articles/s41593-022-01189-0>
  
- Even though you don't get the final results, focus on exploring data & we would love to hear about research ideas on the datasets or collaborations.
  
- Data: [https://drive.google.com/drive/folders/1kFaLNqckbXtQ8RgJwPDjKW\\_PoX5e\\_Kyi?usp=sharing](https://drive.google.com/drive/folders/1kFaLNqckbXtQ8RgJwPDjKW_PoX5e_Kyi?usp=sharing)



## Preprocessed

File name	Description	Shape
<b>Neural Signal</b>		
ffneu_final.npy	ffneu=F-0.7*Fneu(Neuropil) Normalized, Synchronized	(n_neuron, seq_neuron)
spks_final.npy	Deconvolved Normalized, Synchronized	(n_neuron, seq_neuron)
idx_coord_neural.npy	Array that tells neural frame number for each behavior frame.	(seq_behav, )
calcium_final.npy	Raw calcium images	(512, 796, seq_neuron)
stat.npy	Information about neuron positions Docs on suite2p website	(n_neuron, )
<b>Movement</b>		
behav_ori.npy	(Movement) Original limb coordinates	(8, seq_behav)
behav_norm.npy	(Movement) Processed limb coordinates. Normalized, Synchronized	(8, seq_behav)
behav_likely_final.npy	(Movement) Likelihood of predicted pixel from Deeplabcut	(4, seq_behav)
<b>Touch</b>		
touch_behave.npy	(Touch) Original touch stimulus loaded from cvs Start behavior frame, End behavior frame, limb(1=front right, 2=Hind right, 3=Front left, 4=Hind left)	(n_stimuli, 3)



```
im = np.zeros((ops_data['Ly'], ops_data['Lx']))
for n in range(stat.shape[0]):
    ypix = stat[n]['ypix'][~stat[n]['overlap']]
    xpix = stat[n]['xpix'][~stat[n]['overlap']]
    im[ypix, xpix] = importance[n]#n+1
```

## Behavior, Movement

File name	Description
limb1.avi limb2.avi	Behavior videos. Limb1: Right, Limb2: Left
limb1.csv limb2.csv	FL: Front Left, HL: Hind Left x,y: x and y coordinates of a limb likelihood: Likelihood of pixel prediction from Deeplabcut

## Behavior, Touch

File name	Description
limb1.avi limb2.avi	Behavior videos. Limb1: Right, Limb2: Left
touch_timestamps.csv	start, end: behavior frame of start/end of stimulus limb: 1=front right, 2=Hind right, 3=Front left, 4=Hind left

# Synchronization

