

The SLS 2.0 upgrade & its impact on structural biology and drug discovery

SPG Jahrestagung ETHZ 10.09.2024



Philip Willmott, Swiss Light Source, PSI

Contents

- What is a synchrotron?
- What defines a fourth-generation synchrotron?
- SLS 2.0 the upgrade
- MX in today's milieu
- SLS 2.0 the impact on MX and structural biology



Synchrotrons Key features



What is a synchrotron?

- Large-scale facility for generating high-intensity electromagnetic radiation
 - Most commonly in range of VUV to hard x-rays; also down to IR in some cases
 - "Synchrotron radiation"
- Key features of SR
 - Brightnesses many orders of magnitude greater than can be provided by lab-based x-ray sources
 - Extremely collimated beams
 - Extremely narrow beams
 - Tunability of the photon energy
 - e.g., SLS 2.0: ≃ 5 eV to > 80 keV
 - Multiple experiments ("beamlines") around the closed-loop structure of a synchrotron ~ 10 100

PS Center for Photon Scie

SR used extensively in macromolecular structural studies ("raison d'être" for SR!!)



PSI Center for Photon Science















PSI Center for Photon Science





PSI Center for Photon Science





Beamlines



ID (undulator) x-ray source



Brilliance – the synchrotron figure of merit



Units:	[ph/s]
	[mm ² mrad ²][0.1 % BW]



 \mathcal{E} , emittance = size x divergence (both x- and y-directions)



Brilliance (less whimsically)



- σ (source size) and σ' (source divergence) have contributions from both the electron beam and the photon beam
 - Photon part fundamental (diffraction limit)

$$\epsilon_{h\nu} = \sigma_{h\nu} \cdot \sigma'_{h\nu} = \frac{\lambda}{4\pi} \sim 10 \,\mathrm{pm} \cdot \mathrm{rad}$$

- Electron contribution determined by storage-ring performance
- 3rd generation (SLS)
 - Dominated by electron beam: $\epsilon_e \gg \epsilon_{h
 u}$
- 4th generation (DLSR, SLS 2.0)
 - Electron and photon contributions similar
 - \Rightarrow collimated AND small x-ray beams



Small-emittance DLSR-beams for MX



Fourth-generations synchrotrons Diffraction-limited storage rings – SLS 2.0



Double-bend achromats at synchrotrons

• Main limit to reducing emittance due to spread induced at bending-magnet achromats



What defines a DLSR (4th generation synchrotron)?

- Increase brilliance by decreasing emittance in electrons' orbital plane (ε_x)
- How?
 - For a given arc sector, use more bending magnets (M): "multibend achromat" (MBA)



Why only now?

- Using large 3rd-generation magnets would result in
 - An unacceptable increase in ring circumferences
 - Unavoidable alignment errors
- Reduce
 - Magnet sizes
 - More compact 4
 - Reduces B 👎
 - Distances between magnet poles
 - Increases again B description
- Small vacuum vessels
 - Difficult to pump
 - Require special "NEG" coating
 - Porous alloys of Al, Ti, Fe, V, Zr









SLS: Total # magnets = 388; ε_x = 5500 pm.rad SLS 2.0: Total # magnets = 1007; ε_x = 157 pm.rad (ca. 35 x smaller)

SLS v SLS 2.0



SLS v SLS 2.0



SLS

SLS 2.0



Further benefits of the small beams at DLSRs

X-ray source ("undulators")

- Smaller beam width
- ⇒ smaller magnet dimensions
- ⇒ reduced magnetic forces
 - "Force compensation" possible
- \Rightarrow smaller gap \Rightarrow more intensity & higher hv
- ⇒ more compact and stable designs



X-ray optics

- Smaller beam cross-sections
- ⇒ smaller dimensions of
 - x-ray mirrors
 - diffracting elements (crystals, gratings, multilayers)
- ⇒ more compact, lighter x-ray optics components
- \Rightarrow greater stability, less vibrations







SLS v SLS 2.0 performance enhancements in numbers





Macromolecular structure determination at DLSRs A bright future with complementary competition



The new kids on the block



- < 2012, MX enjoyed near total dominance</p>
- ~ 1980 2010: Developments in cryoEM
 - detectors
 - sample prep { Resolution
 - image analysis / breakthrough ~ 3 Å
- CASP14 (Nov. 2020): Alphafold2
 - Al program (DeepMind, Google)
 - Predicts structure from amino-acid sequence alone
 - Based on PDB database
 - 2022: structures uploaded of ~200 million proteins from 1 million species, covering nearly every known protein on the planet



< 2012: > 2012: "Blobology" hi-res cryoEM



Experiment Alphafold2



MX v cryoEM... so far (up to September 2024)



cryoEM



Real-space imaging technique

Single particles, no crystals needed

See also SPG plenary talk, Henning Stahlberg 17:15, 09.09.2024 and Luca Rima 17:00 today, this session

Alphafold2 *et al.*



Extracted from https://www.youtube.com/watch?v=gg7WjuFs8F4&t=149s

- Critical Assessment of Protein Structure Prediction CASP14, Nov. 2020
 - Metric: Global distance test global score (GDT-TS)
 - Percentage of well-modelled residues w.r.t. target
 - 90% is (was!) the holy grail





Alphafold2 in a nutshell



Alphafold2 in summary



1 million species

Images: creative commons and PRW

- Phase problem essentially solved!
- MIR, MAD, SAD, etc. no longer needed
- Molecular replacement (MR) still workhorse
- Refinement of Alphafold2 predictions
 - Is crystal structure = in-vivo structure?
- July 2024: Number of proteins solved by Alphafold2 since November 2020...

> 200'000'000!!

- 35% "highly accurate"
- 45% "sufficiently accurate for many applications"



Atomic resolution in structure determination

- A spatial resolution of approximately 2 Å or better is required to resolve individual atoms within a protein structure
- At this resolution:
 - Individual atoms and their positions can be distinguished
 - The electron density map is detailed enough to identify the atomic structure, including side chains of amino acids
 - Bond lengths and angles can be accurately measured
- For very high-resolution structures, resolutions better than ca. 1.5 Å are needed
 - Provides even more precise details about the atomic arrangement
 - ca. 1 Å resolution allows identification of hydrogen atoms, which are typically challenging to resolve at lower resolutions



Image from: https://pdb101.rcsb.org/learn/guide-to-understanding-pdb-data/resolution



Membrane proteins and GPCRs

- Membrane proteins
 - Relay signals between cell's internal and external environments
 - Transfer chemicals across cell membrane
 - Molecular weights ~ 10 to over 200 kDa
- G-protein-coupled receptors (GPCRs)
 - Recognize a wide variety of stimuli
 - Photons, ions, proteins, neurotransmitters, hormones...
 - Activate cellular responses
 - Molecular weights ~ 40 to 100 kDa



Photosynthetic Reaction Centre



Serial Synchrotron Crystallography (SSX) @ DLSRs

Membrane proteins

- 1/3 of all proteins
- 2/3 of medicinal drug targets
- 1 2% of all MX-solved structures!
- Why so under-represented?
 - Hydrophobic, hard to crystallize
 - Often micron-sized, poor quality
 - Improve using lipid cubic phase (LCP)
- Serial synchrotron crystallography (SSX)
 - RT or cryo
 - Conformational landscapes (3D shape)
 - Dynamics down to µs
 - Uses much less material than SFX @ XFELs



RT-SSX



Requirements for fragment screening

- Precise location and orientation of SMALL fragment on LARGE biological target
 - High resolution ~ 1.8 Å or better
- Fast throughput (100's of samples)
 - @ SLS 2.0
 - ca. 30+ fragment samples/hr
 - ca. 10 minutes/structural solution (local)
 - Bottleneck use off-site supercomputers
- Requires automation!!
 - 2025 onwards: ~ 10 100 TB/day!!
- Resolution and time-consuming structural solutions make cryoEM unsuited to fragment screening



α-ketoamide inhibitor with SARS-CoV-2 main protease 1.95 Å, PDB 6Y2F Zhang *et al.*, Science 368, 409–412 (2020)



Requirements for fragment screening



MX@SLS and SLS 2.0

- Three beamlines
 - 2001: PXI ID beamline: mainly scientific research, cutting-edge developments; some industry
 - 2004: PXII ID beamline: exclusively proprietary and drug discovery beamtime. Funded by industry
 - 2007: PXIII SB beamline: partly research, diffraction screening, phasing, industry. Upgraded BL completed in 2023, first users!!
- July 31st 2024: the 10'001st PDB entry from SLS registered!
 - Most # PDB entries/year/BL worldwide





PS Center for Photon Science

MX@SLS and SLS 2.0

SLS 2.0 – hi-speed, automated, intelligent learning



"Digital beamline scientist & digital user"





MX @ SLS 2.0 - "1³"



MX @ SLS 2.0 and machine learning for drug design

- Predictive vs. experimental validation:
 - Al/machine learning:
 - Predicts static structures with high accuracy
 - No physiological information about the dynamic nature or how proteins interact with ligands, etc.
 - Not (yet) reliable re. details of potential binding sites or how different ligands interact with these sites
 - Generates initial structure models and potential binding sites ⇒ speeds up preliminary stages of drug discovery
 - MX fragment Screening:
 - Directly observes how and where small chemical fragments bind to a target protein crucial for drug discovery!
 - Helps identify binding sites, understand binding affinities
 - Guides the design of more potent and selective compounds
 - Can identify conformational flexibility
 - Provides insights into how binding events can induce structural changes/dynamics
 - Important for understanding the full range of a protein's functional states
 - Essential experimental validation of predictive AI models
- Summary
 - Al algorithms are transformative tools for predicting protein structures
 - Fragment screening in MX remains a vital experimental tool
 - These two methods are complementary: Al-driven predictions provide valuable initial insights that guide experimental validation and optimization in drug discovery



Summary











Synchrotron/XFEL massive open online courses (MOOCs)



EPFL: two six-week Massive Open Online Courses (MOOCs)

- Introduction to synchrotron and XFEL radiation Part 1
- Introduction to synchrotron and XFEL radiation Part 2





Brilliance since Röntgen

