

From the Microplex to Neural Systems

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Parker Symposium
31 March 2012

It All Started with Sherwood!!

DEVELOPMENT OF HIGH DENSITY READOUT FOR SILICON STRIP DETECTORS

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University of Hawaii, Hawaii, USA

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CERN, Geneva, Switzerland

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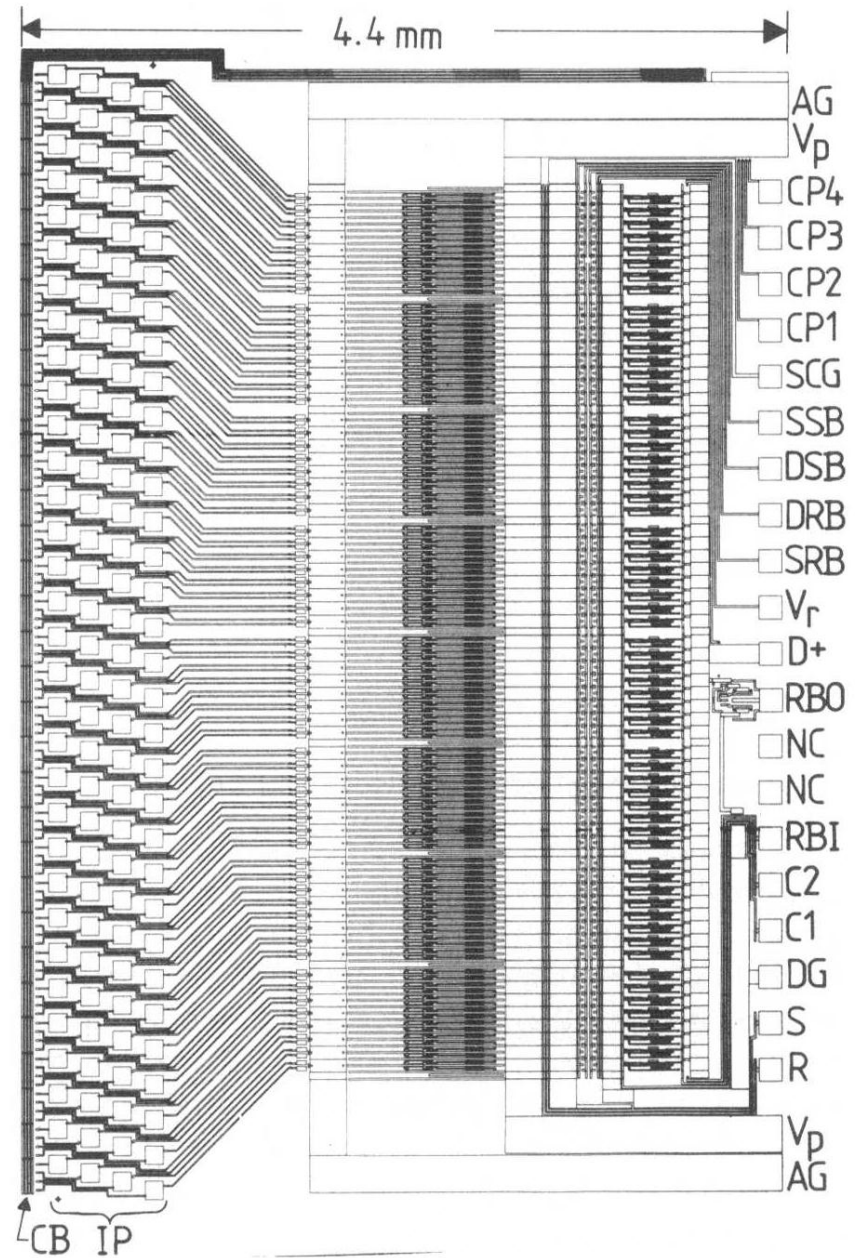
Microplex Chip

NIM 226 (1984) 200-203.

A compact readout for silicon strip detectors is being developed. It employs an nMOS circuit with 128 channels of charge sensitive amplifiers and multiplexed output.

Microplex Layout

- 128 low-noise amplifier channels
- Multiplexed output
- High Density Wire Bonding



Application to Other Scientific Fields

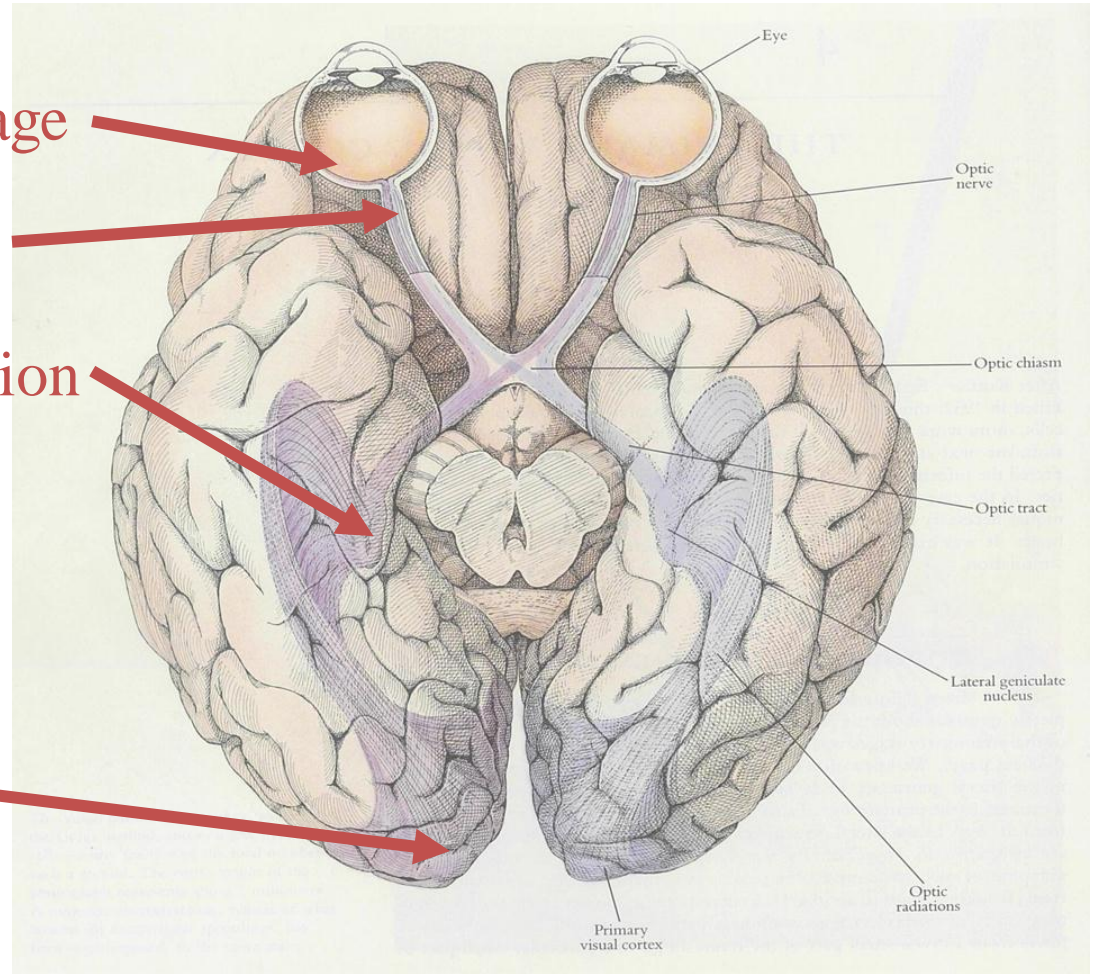
Pixel detector +
first processing stage

cable

relay station

The Visual System

more
processing



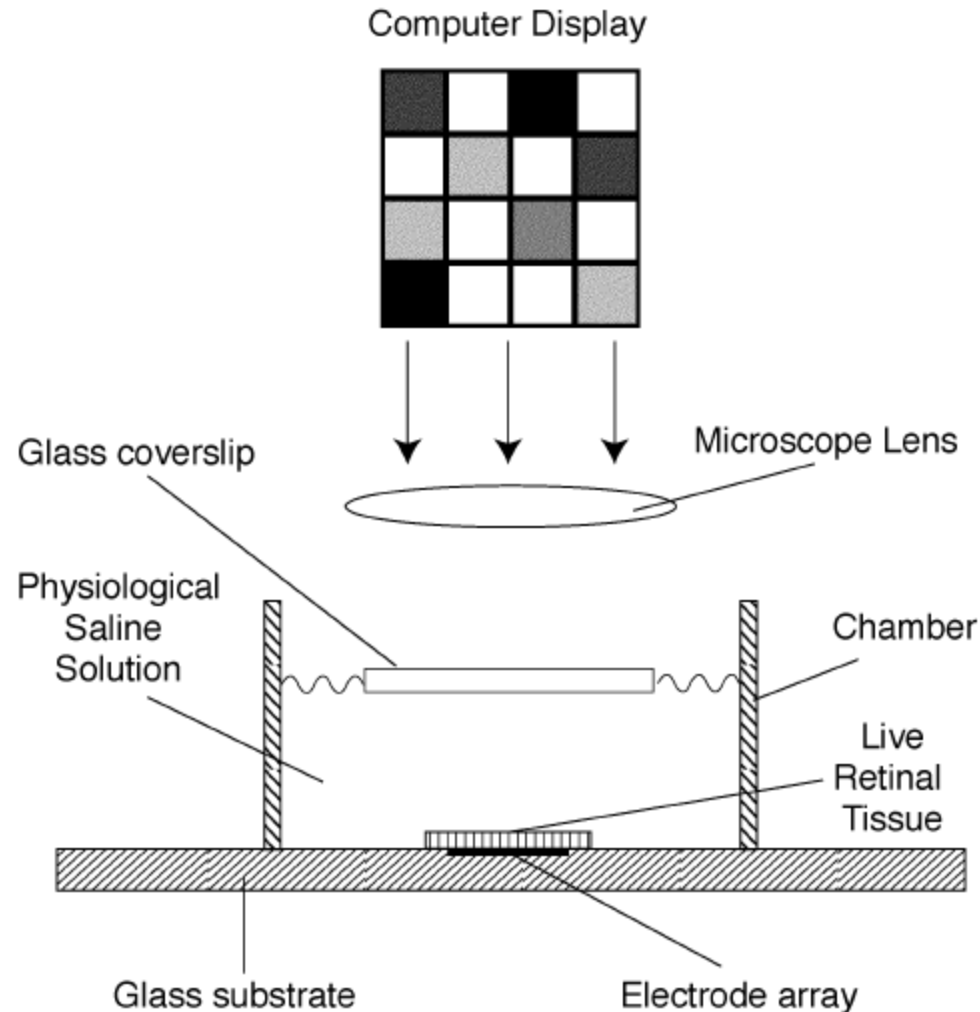
How do neural systems develop (get wired up)?

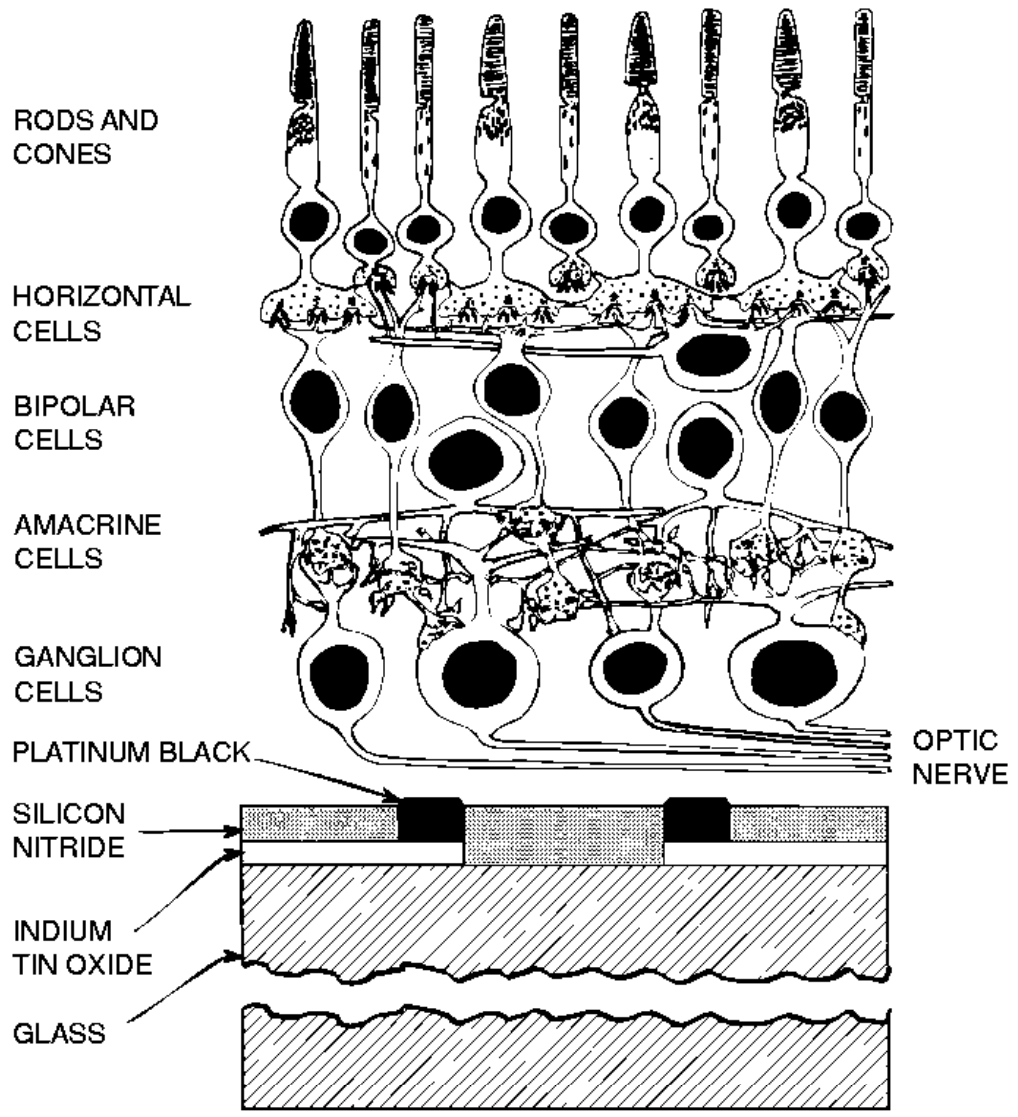
How do neural systems process information?

How can neural systems be repaired when something goes wrong (due to injury or disease)?

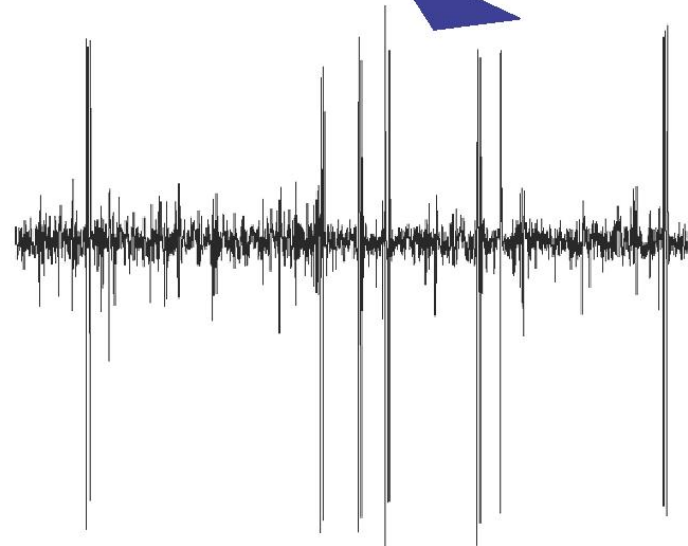
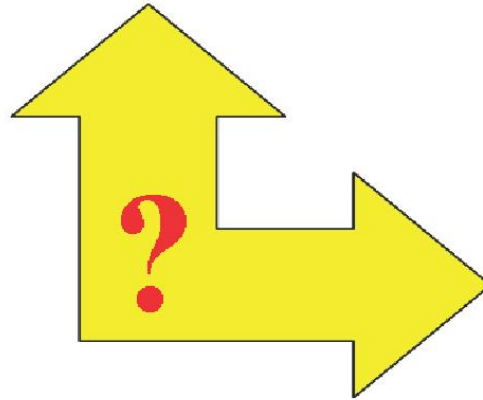
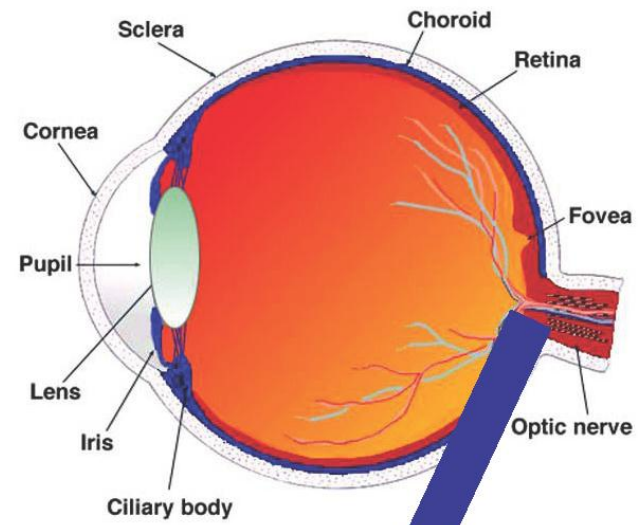
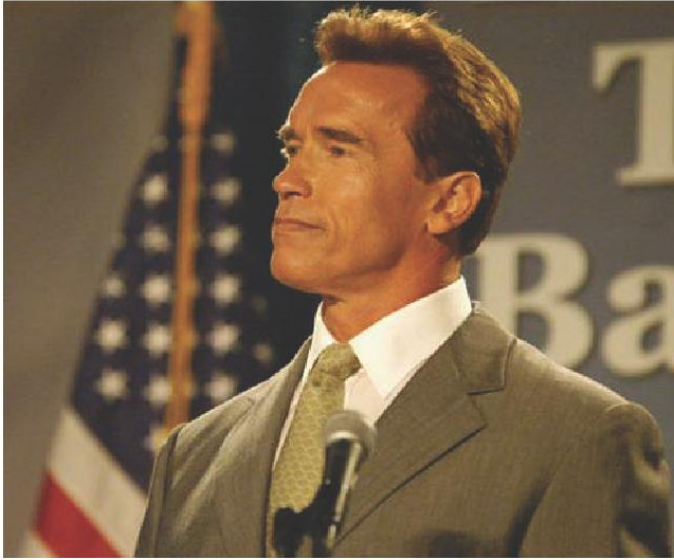
Functional Properties of the Retina

M. Meister, J. Pine, D. A. Baylor,
J. Neuroscience Meth. 51 (1994) 95.

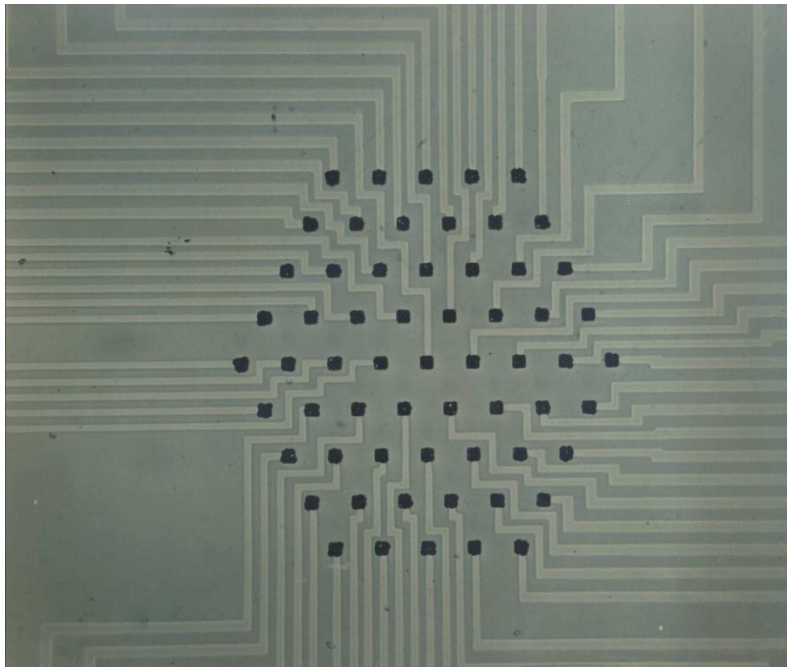




Recording Neural Activity: Retina



State-of-the-Art Multielectrode Array (1990)



- 61 electrodes,
- 60 μm electrode spacing,
- conventional electronics
- Mechanical “zebra” interconnect,
- tens of retinal ganglion cells simultaneously detected

How do we increase, by an order of magnitude, the number of neurons simultaneously detected – from tens of neurons to hundreds of neurons?

Number of Neurons: one→tens→hundreds→???

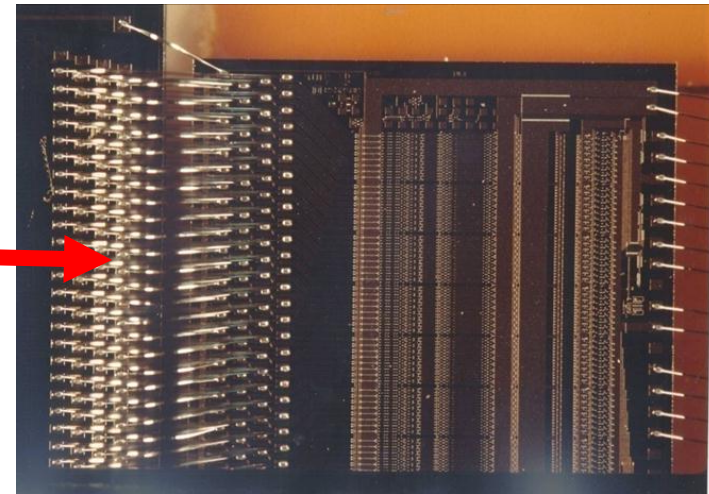
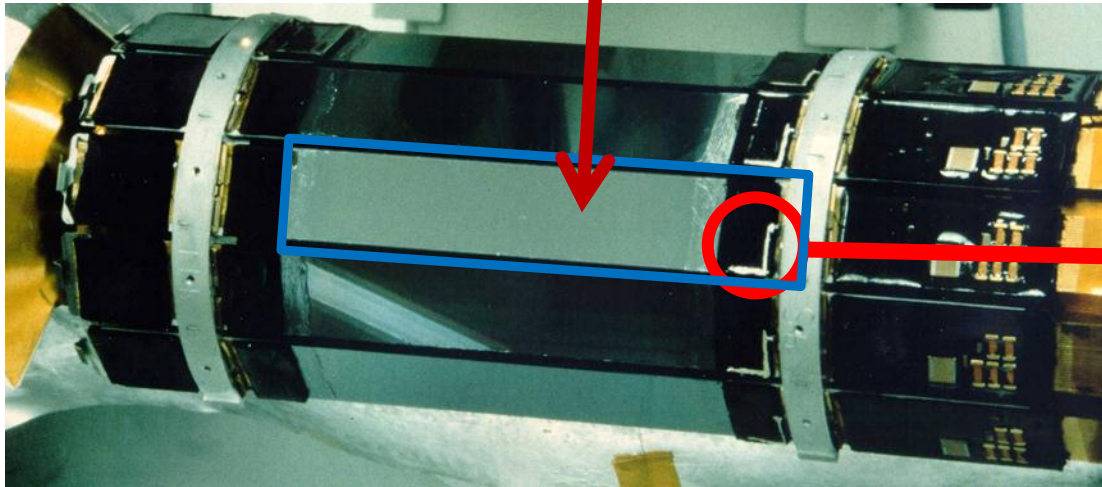
Number of Electrodes: 1→61→512→???

Sherwood, with the SSVD project, had shown the way:

- Silicon nanofabrication technology
- Multichannel low-noise analog amplifier integrated circuits
- Time multiplexing
- High-density wire bonding for interconnect

→ Make a Neural Recording (&Stimulation) Module
look like an SSVD Module

Turn a 61-electrode multielectrode array
into something like a
512-channel Silicon Strip Detector Module



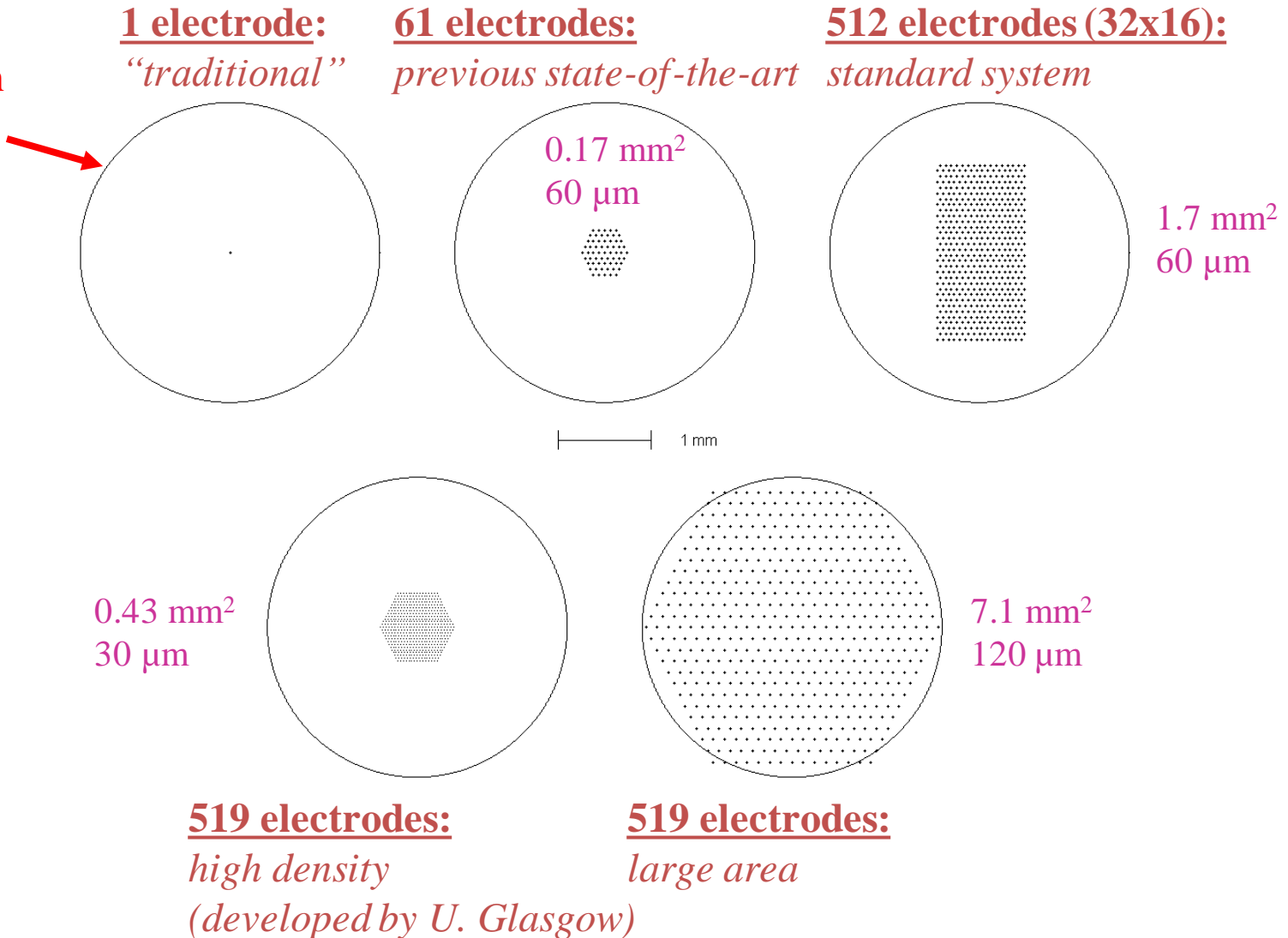
Silicon Strip Vertex Detector:
512 channels/module (18K channels total)

Microplex readout chip
(128 channels, 47.5 μm pitch)

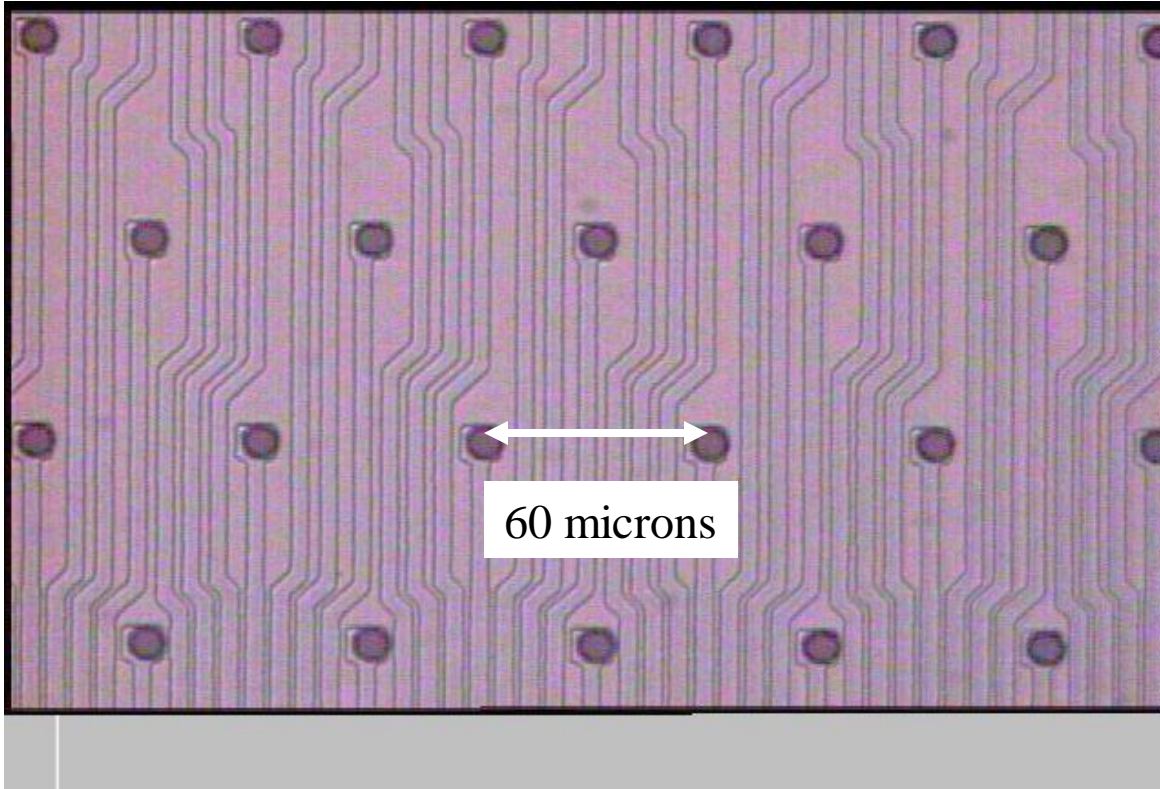
Electrode Array Geometries

(Electrode diameters = 5 μm ; area and electrode spacing given below.)

Input region
for monkey
MT neuron



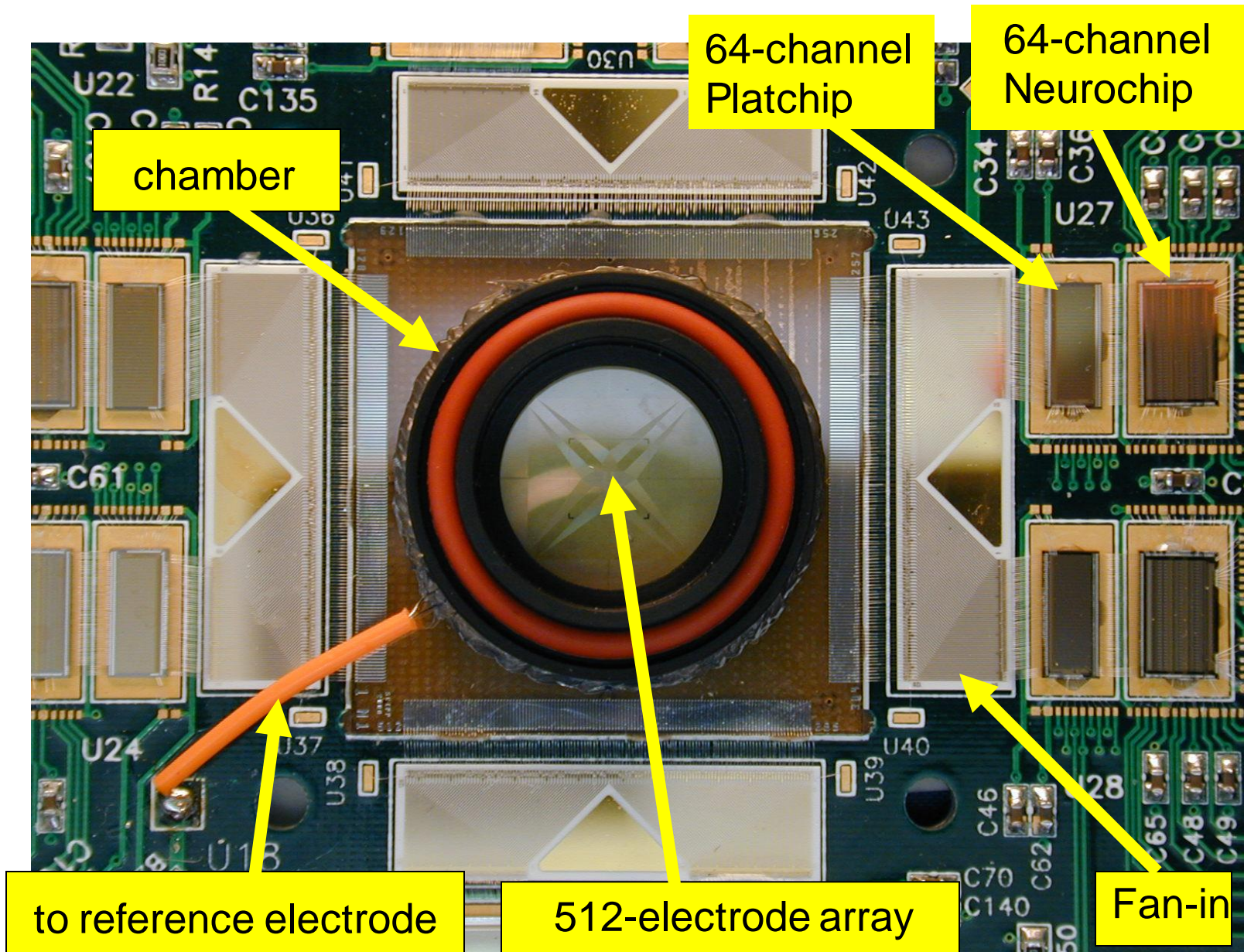
Section of 512-electrode Array (32x16)



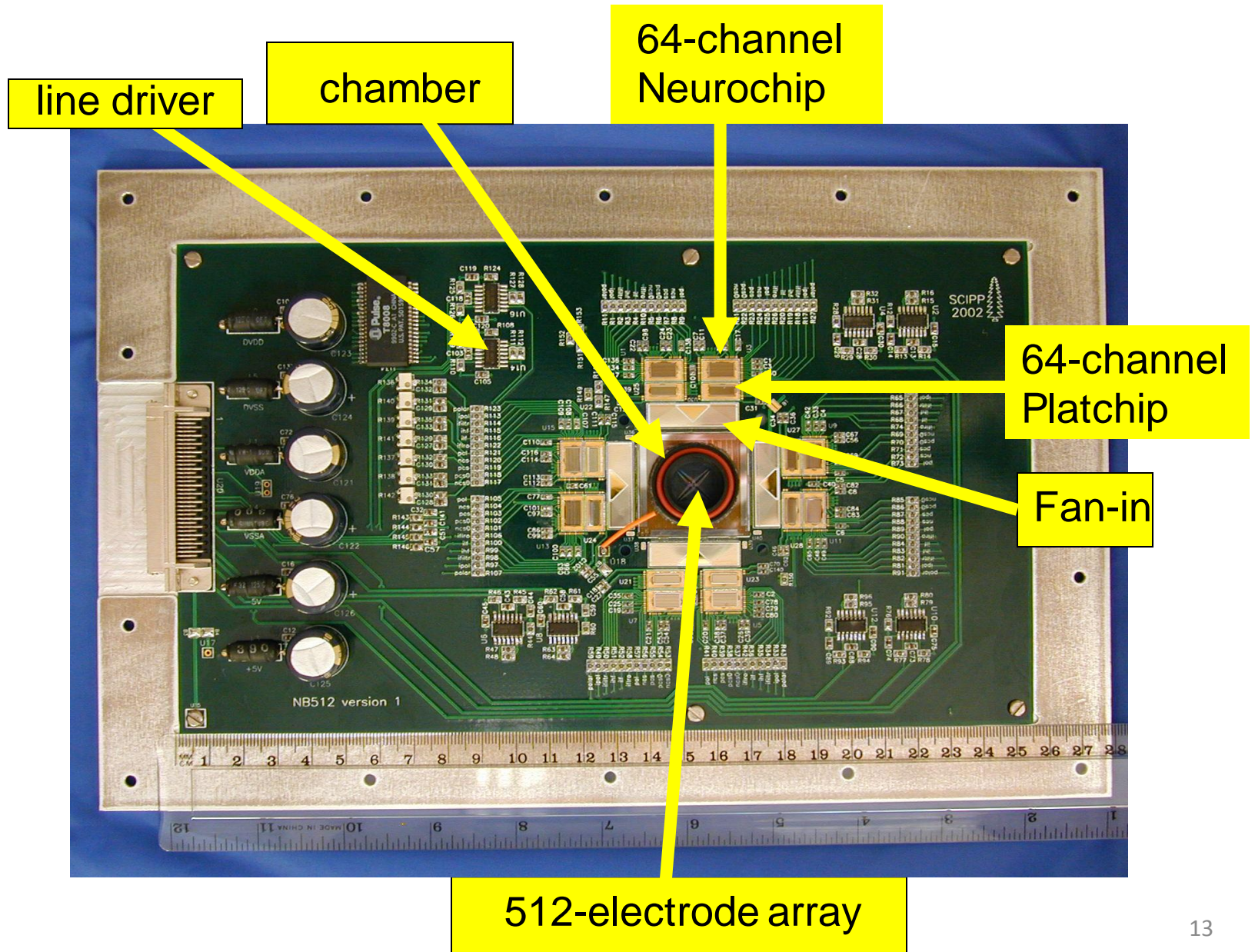
Electrode diameter = $5 \mu\text{m}$

(Litke et al., IEEE Trans. Nucl. Sci. (2004) 1434)

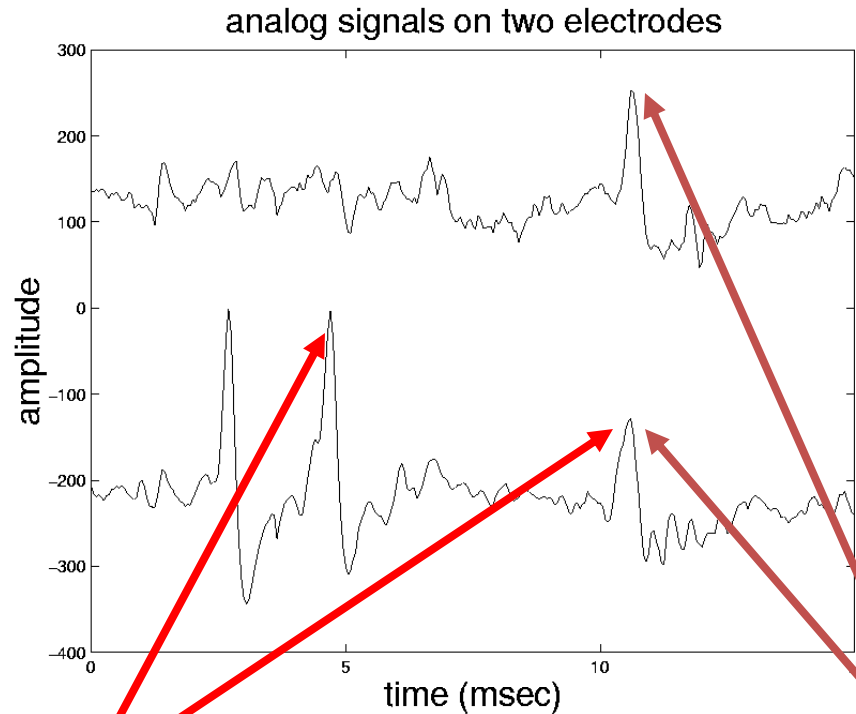
Section of 512-electrode “Neuroboard”



512-electrode "Neuroboard"



Spikes on electrodes \Rightarrow spikes from identified neurons



2 separate cells
recorded on same
electrode

Same cell
recorded on
2 electrodes

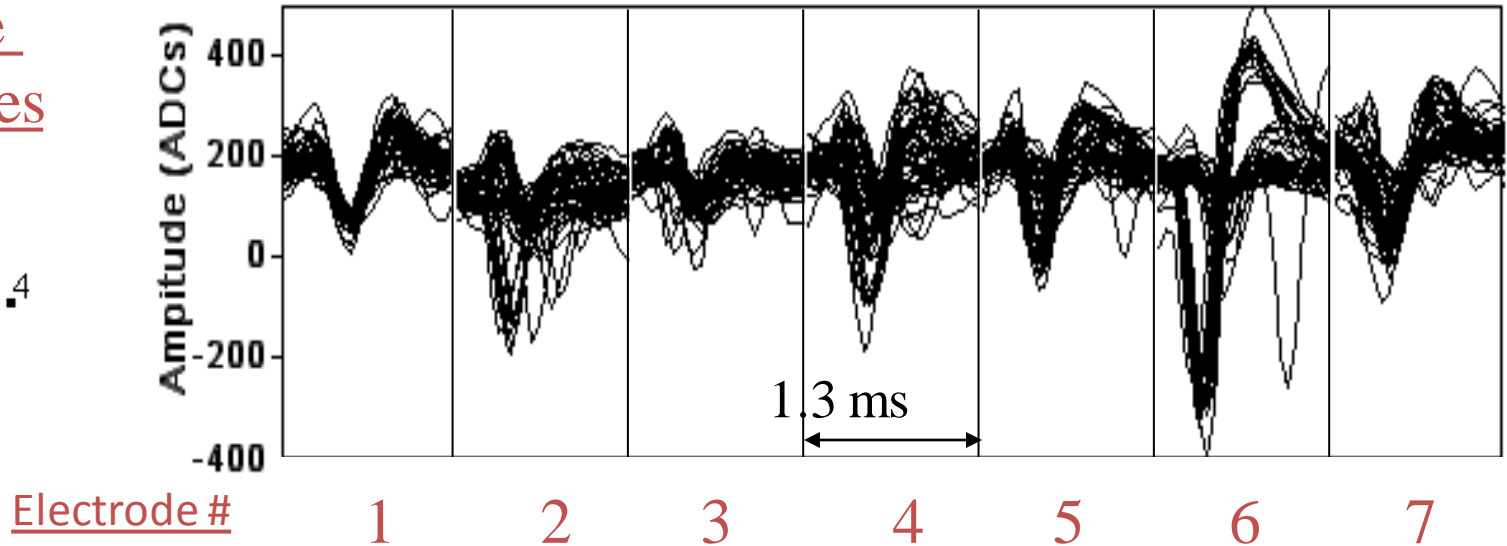
Neuron Identification

(signals on electrodes \Rightarrow spikes from identified neurons)

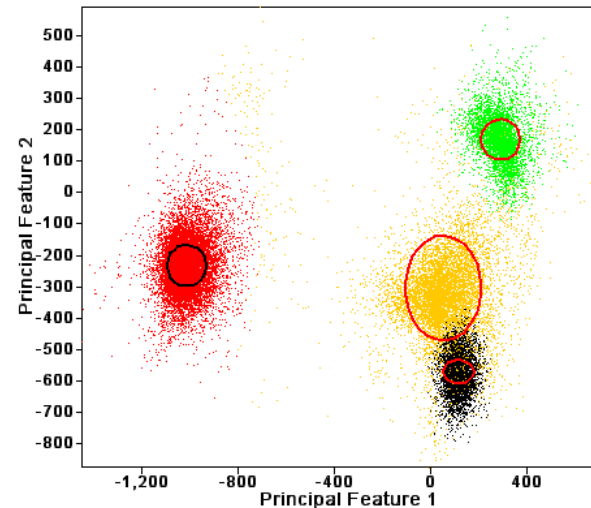
7x26=182 measurements

Multiple electrodes

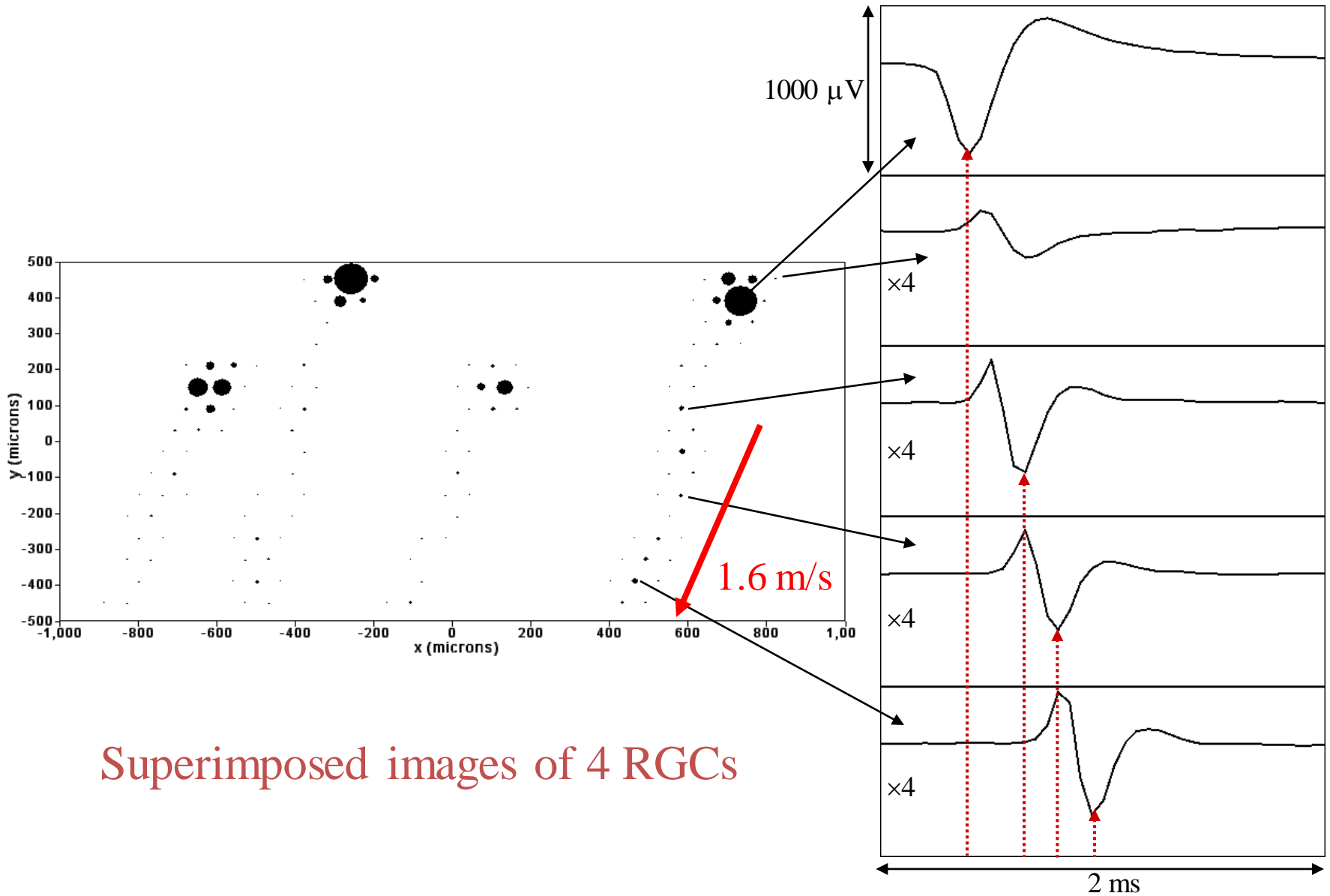
.3 .2
.5 .1 .4
.6 .7



- Principal Components Analysis
Find ~5 most significant variables that are linear combinations of the 182 measurements
- Multidimensional Clustering
 \Rightarrow Identified Neurons



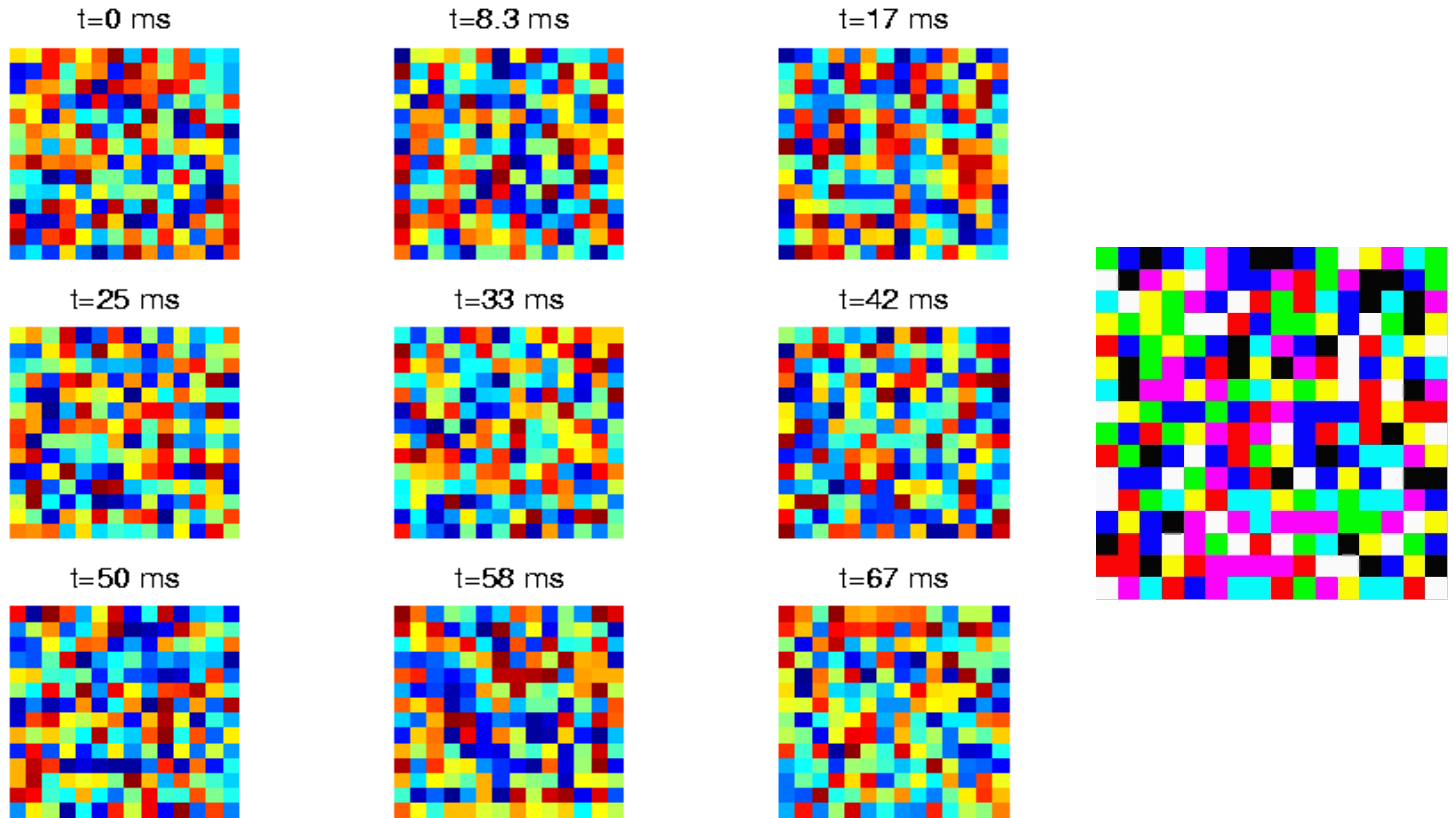
Electrophysiological Imaging



Superimposed images of 4 RGCs

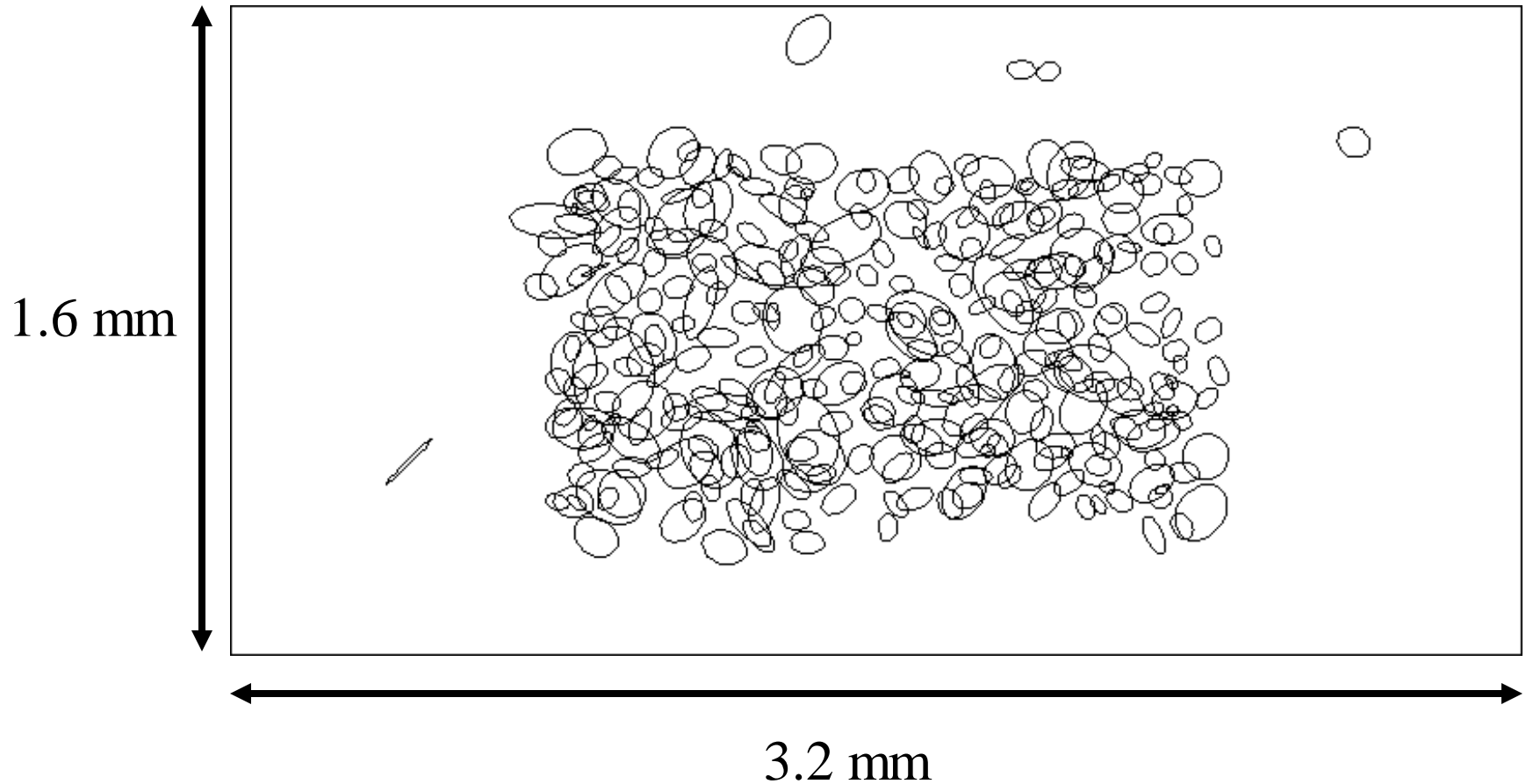
Measure the visual response properties of identified neurons

⇒ white noise analysis: use time sequence of random checkerboard images

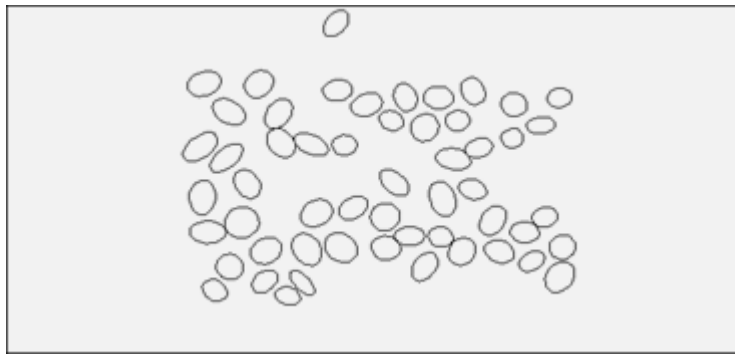


⇒ measure the “spike-triggered average” (sta) response for each neuron

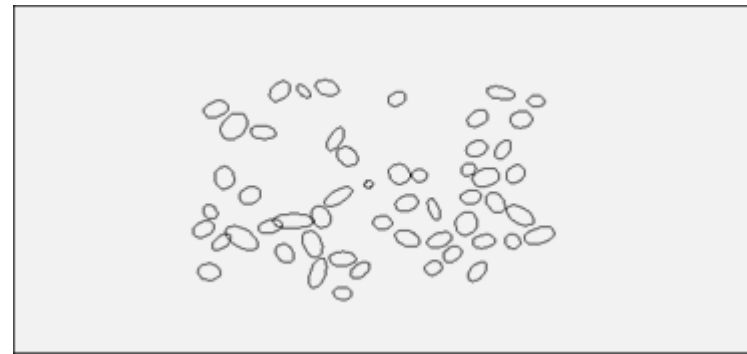
Light-sensitive regions (“receptive fields”) on the retina
for 338 identified retinal ganglion cells (RGCs)



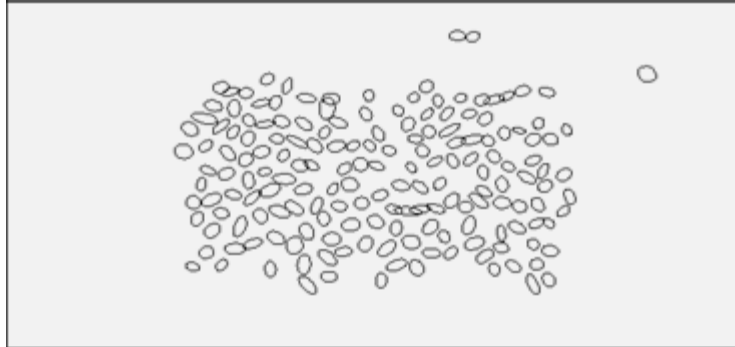
ON-
parasol



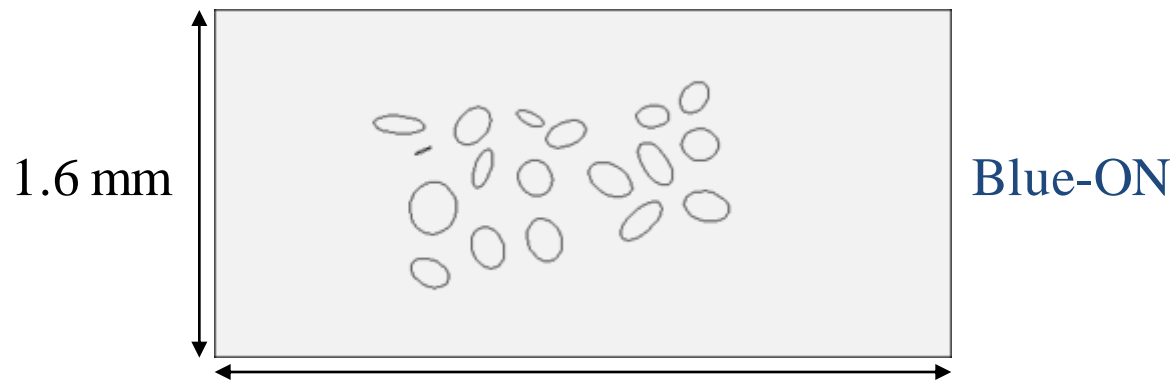
OFF-
parasol



ON-
midget

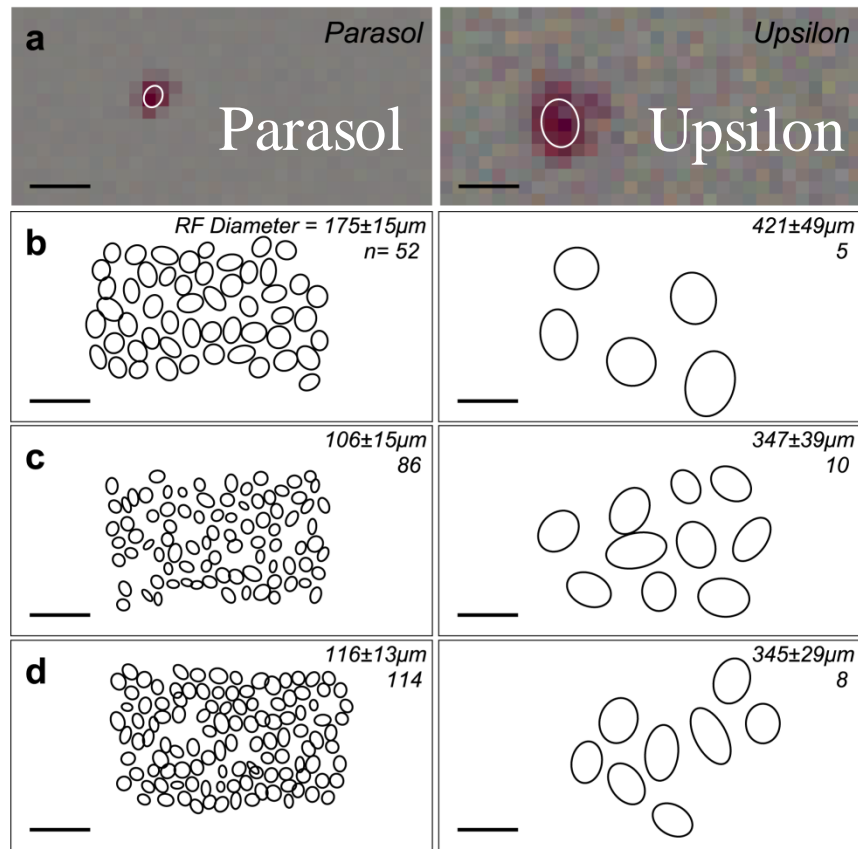


OFF-
midget



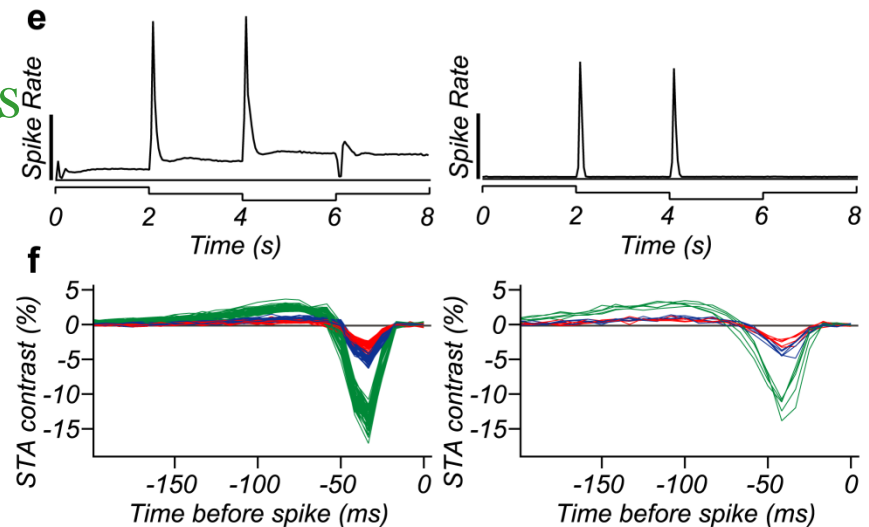
Blue-ON

Receptive field



Mosaics in 3 preparations

Response to diffuse light steps



Sta time filter

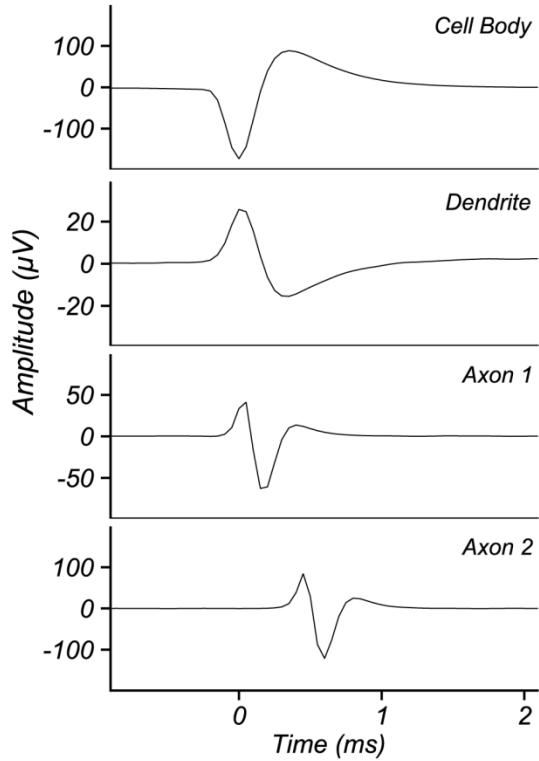
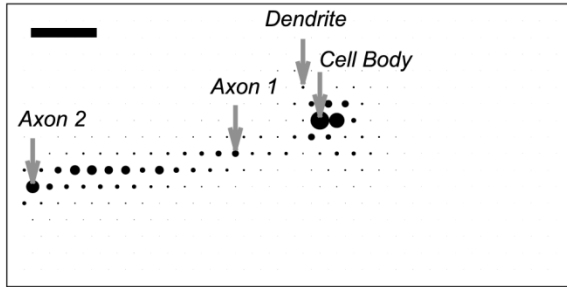
Parasol

Upsilon

Amacrine

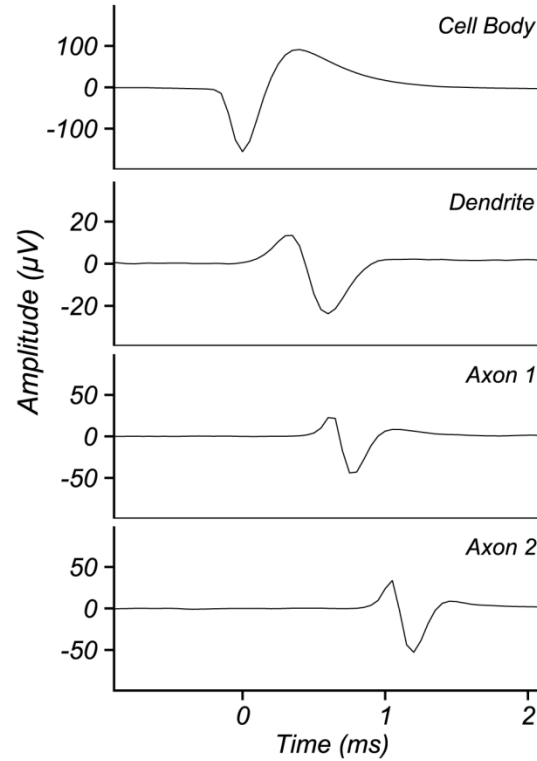
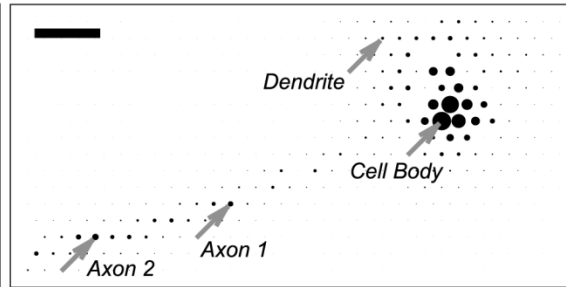
a

Parasol



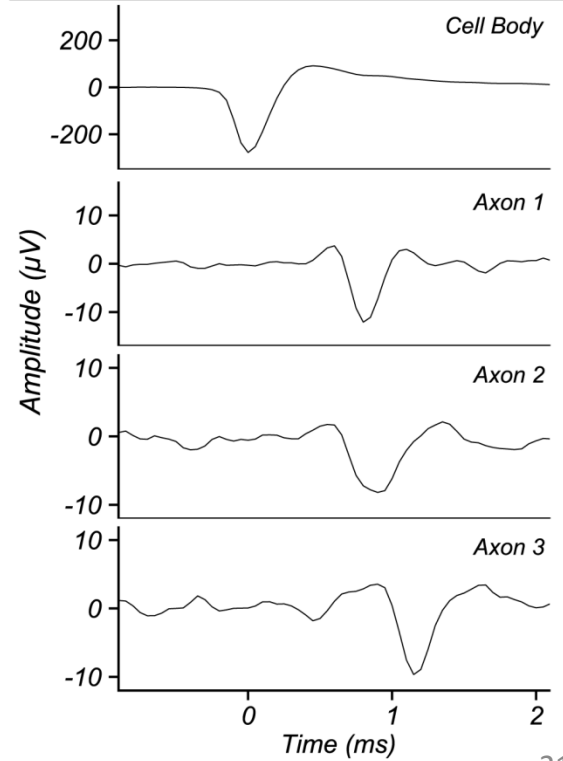
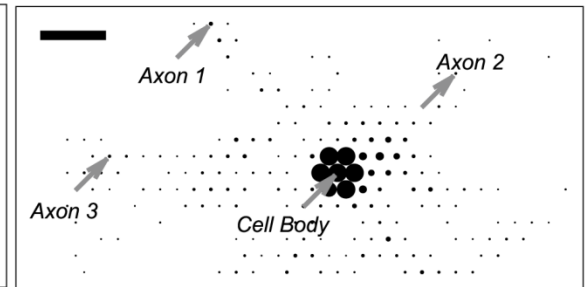
b

Upsilon

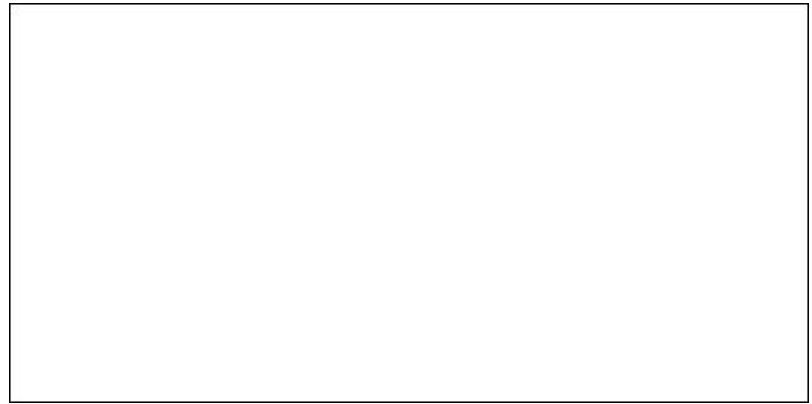


c

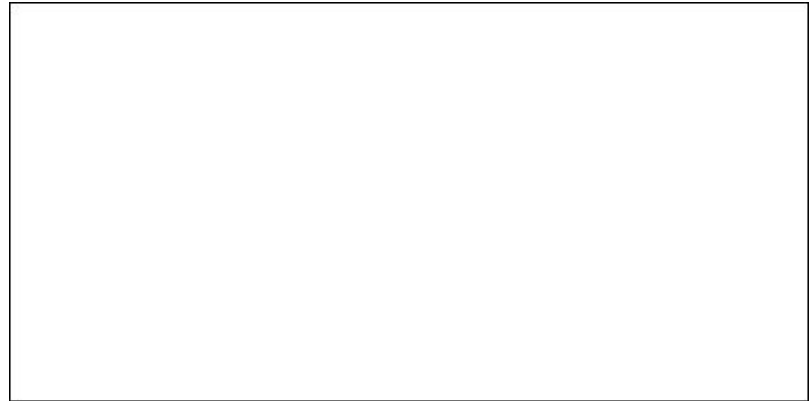
Amacrine



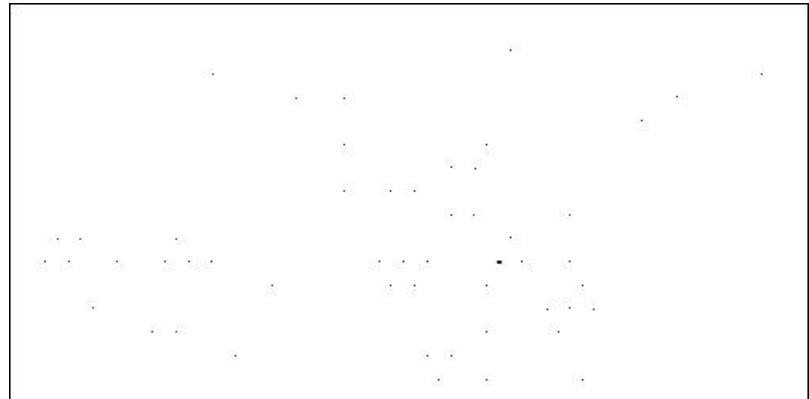
OFF Parasol



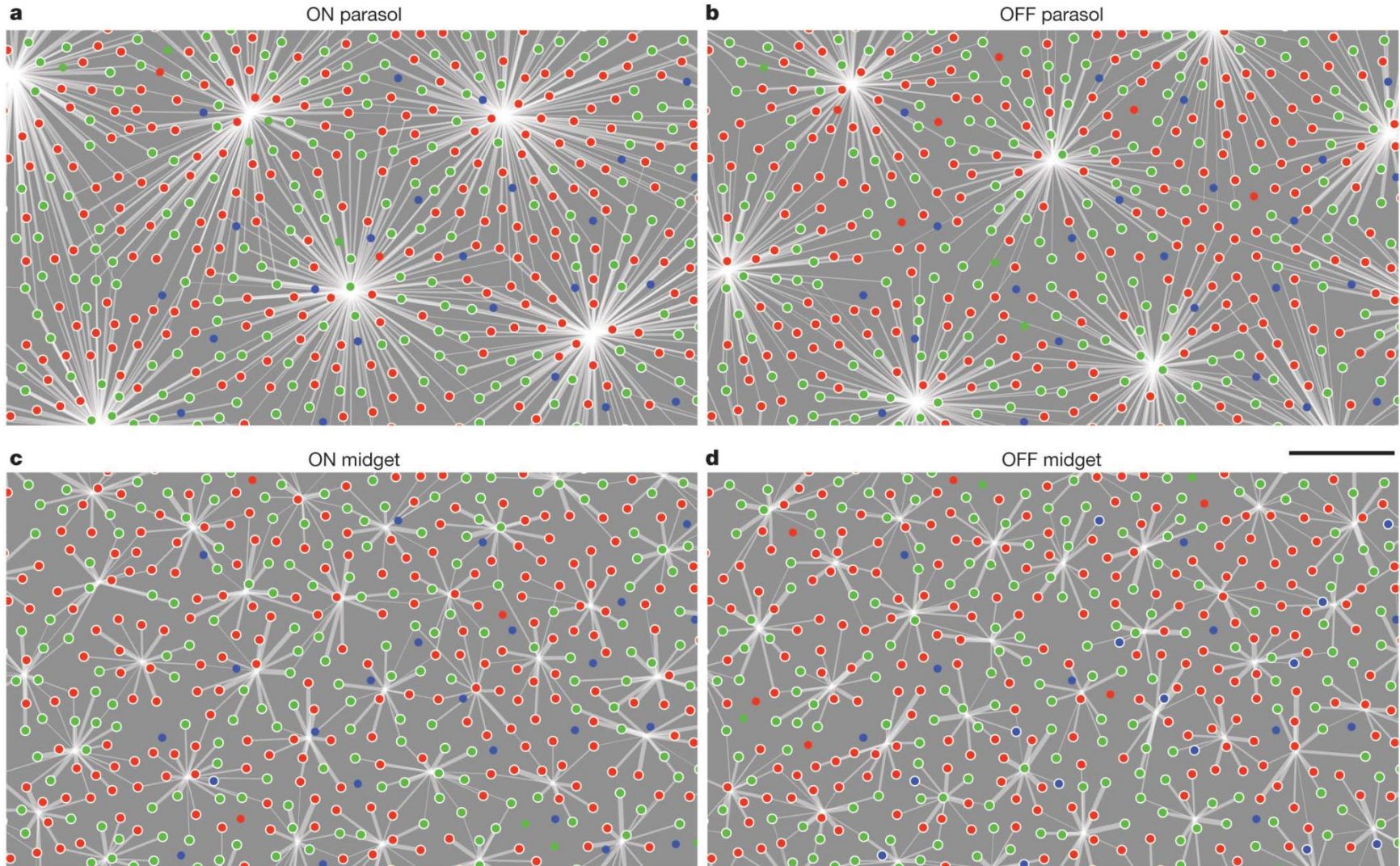
Upsilon



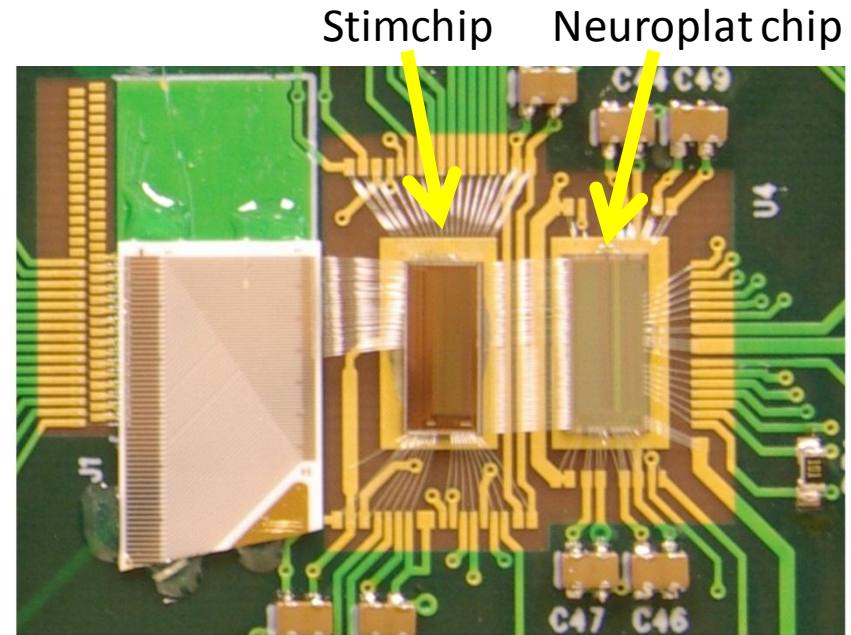
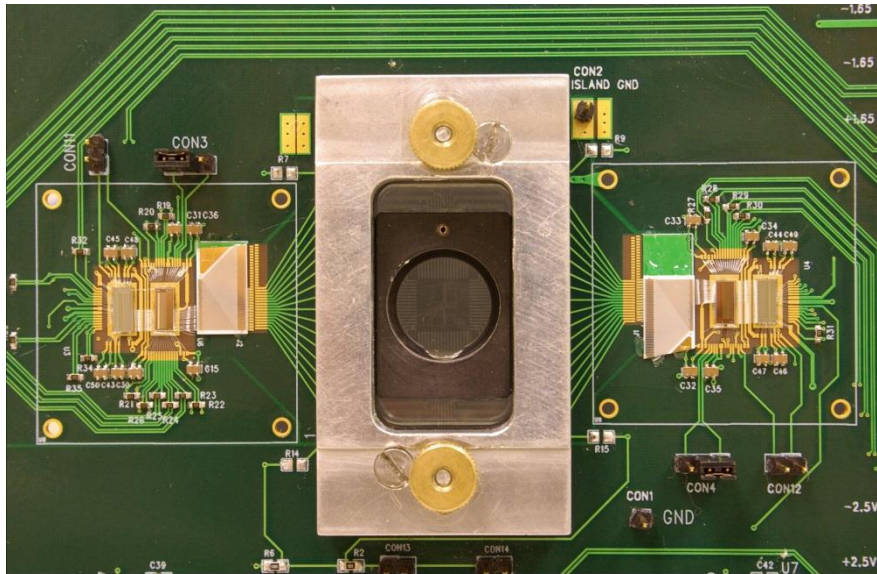
Polyaxonal Spiking Amacrine



Functional connectivity in the retina at the resolution of individual photoreceptors and individual retinal ganglion cells (Field et al., Nature 2010)



61 Electrode Neural Stimulation/Recording System

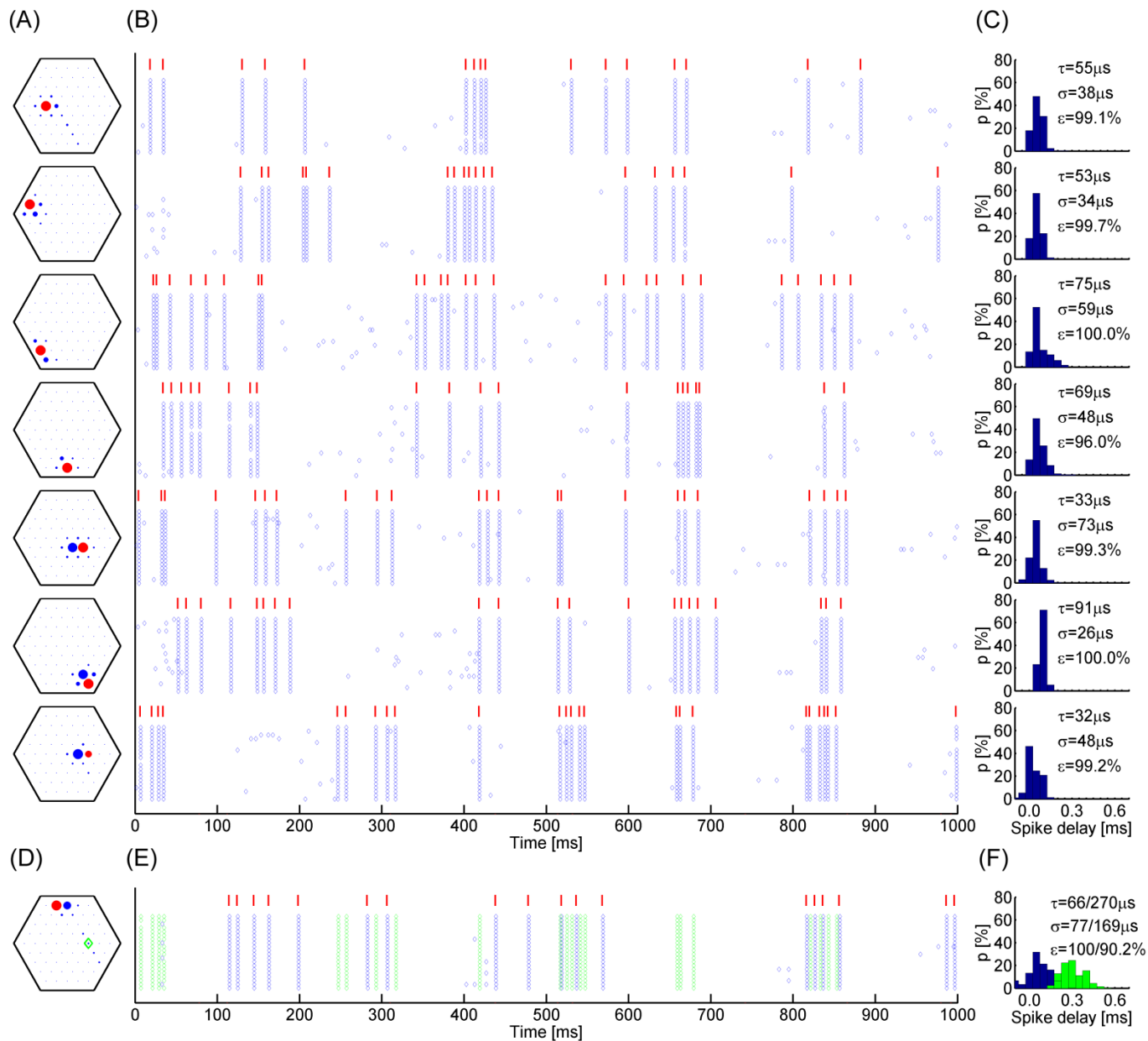


Stimchip

- 64 channels
- ability to generate arbitrary, independent stimulation pulses on each channel, under software control
- → spatio-temporal patterned stimulation

(P. Hottowy et al., Analog ICs and Sig. Processing **55** (2008) 239)

Spatio-Temporal Patterned Stimulation of Multiple Neurons with Multiple Electrodes (Application to Retinal Prosthesis Studies)



Technology Development and Applications to Other Neural Systems:

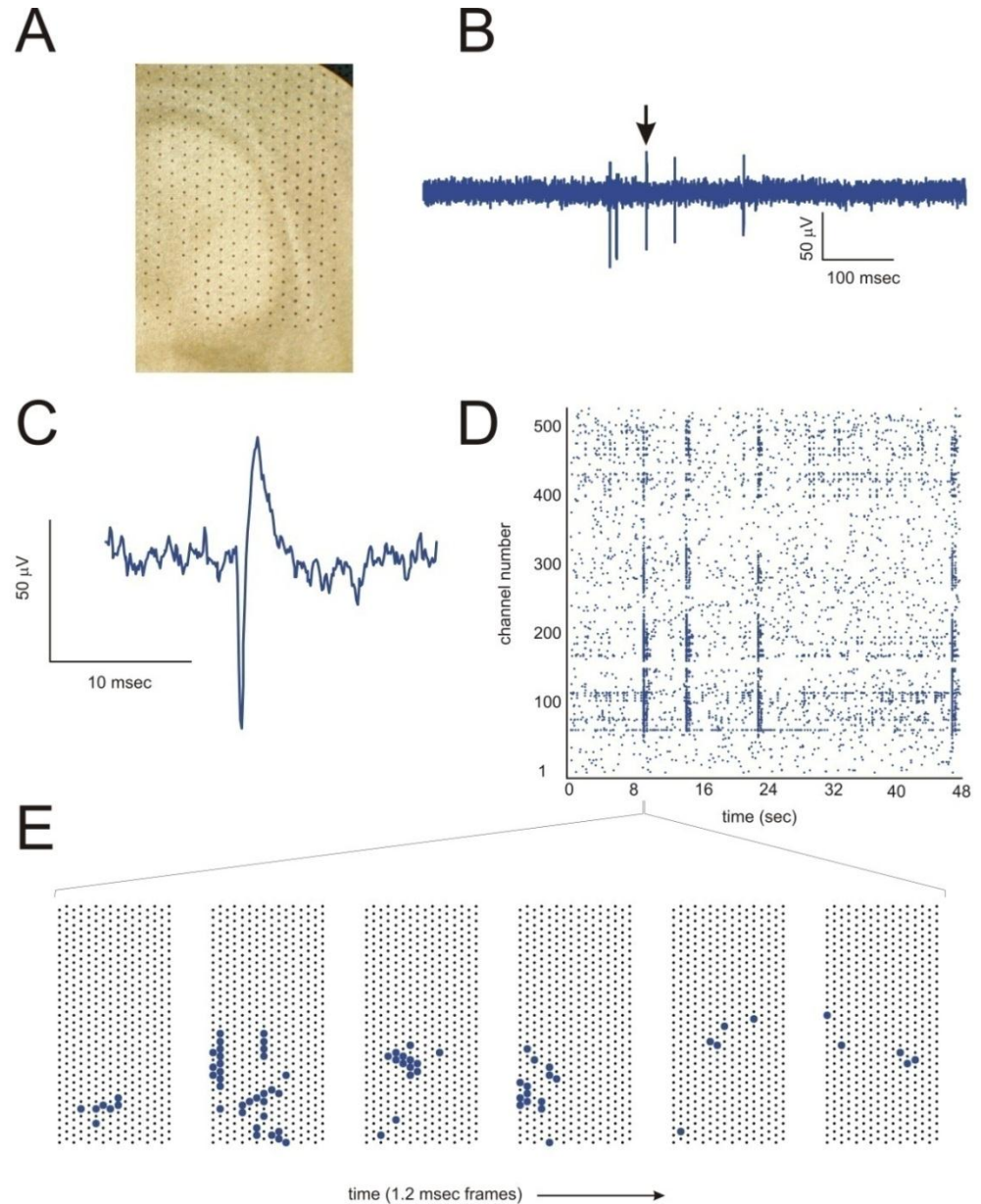
Cultured Brain Slices

(with John Beggs, Shinya Ito, Frances Yeh)

- Neural Connectivity and Neural Circuitry
- Neurological Disorders (with transgenic mice)
- Neurotoxicity

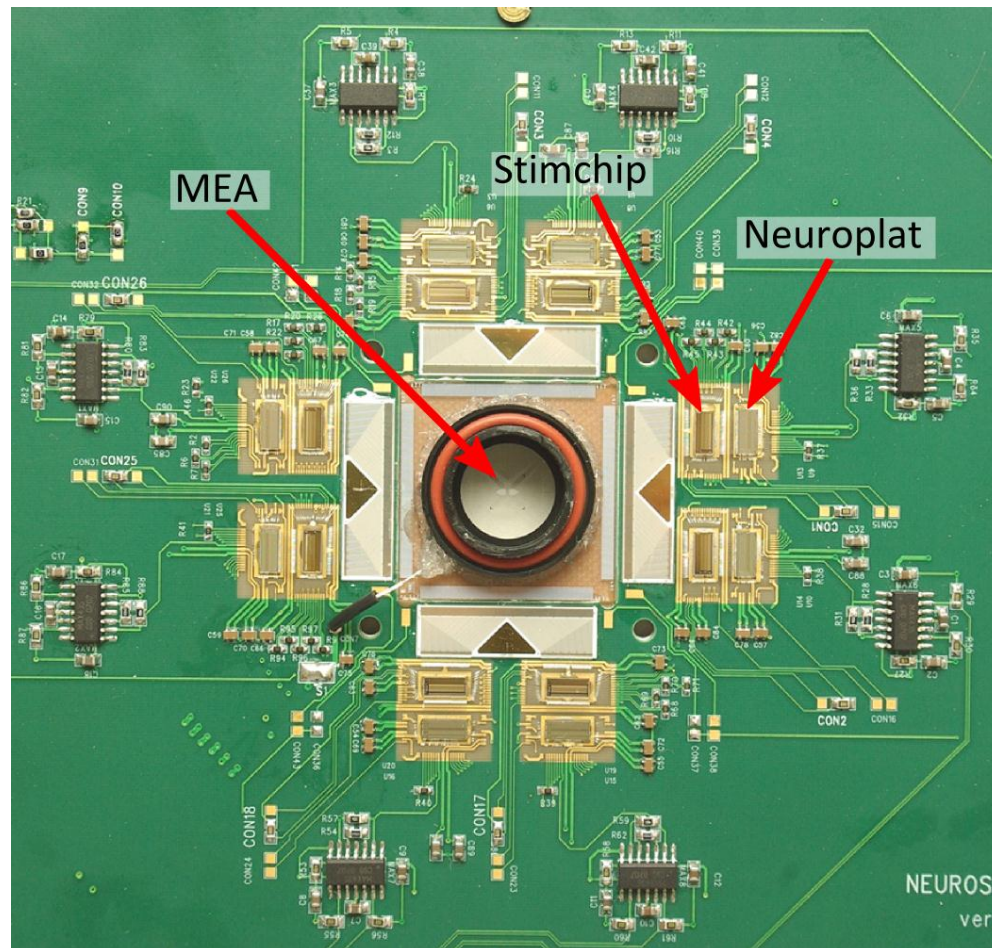
Stimulating & Recording Neural Activity: Cultured Brain Slices

Rat cultured cortical slice on 512-electrode array



512 Electrode System for Stimulating and Recording Neural Activity

write information into a cortical network and then record its response
⇒ use multielectrode electrical stimulation combined with multielectrode recording (as with retinal prosthesis studies)



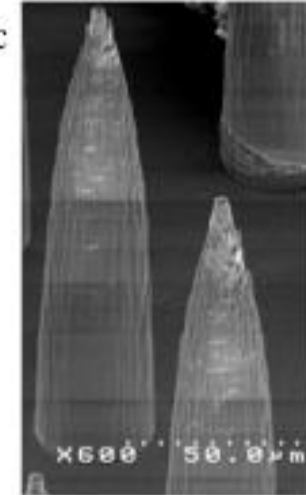
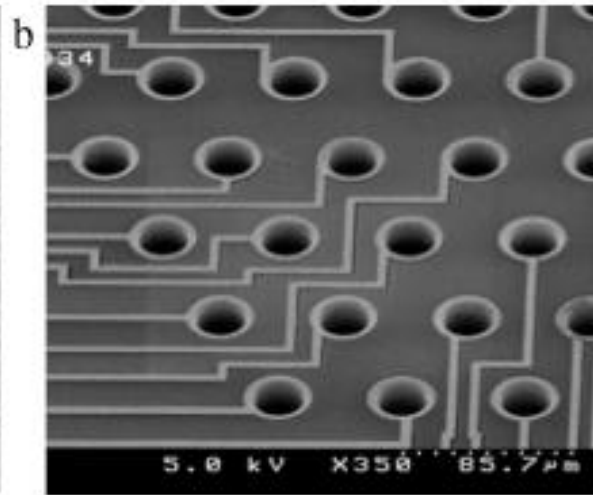
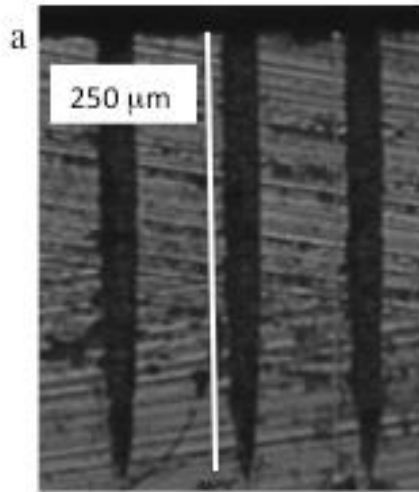
Acute Brain Slices

Bed-of-Nails electrode arrays:

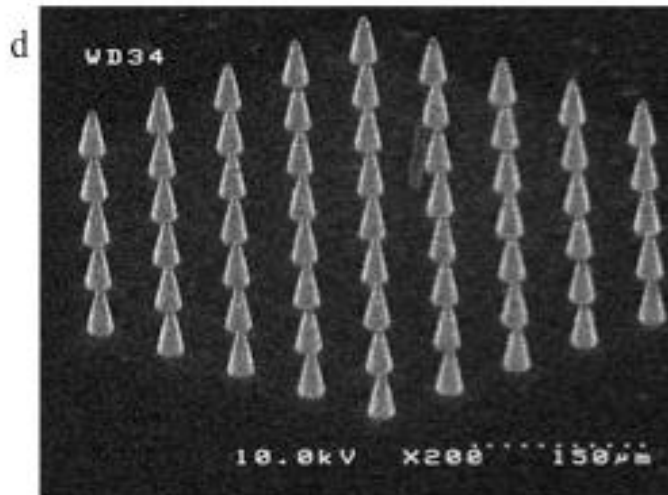
can penetrate through ~50 microns of dead tissue due to cut

- Based on 3D Detector Technology from Sherwood
- First Mechanical prototypes fabricated by Chris Kenney
- Electrically Active Bed-of-Nails Arrays Implemented by Debbie Gunning

Bed-of-Nails Electrode Array: (Fabricated by Debbie Gunning)

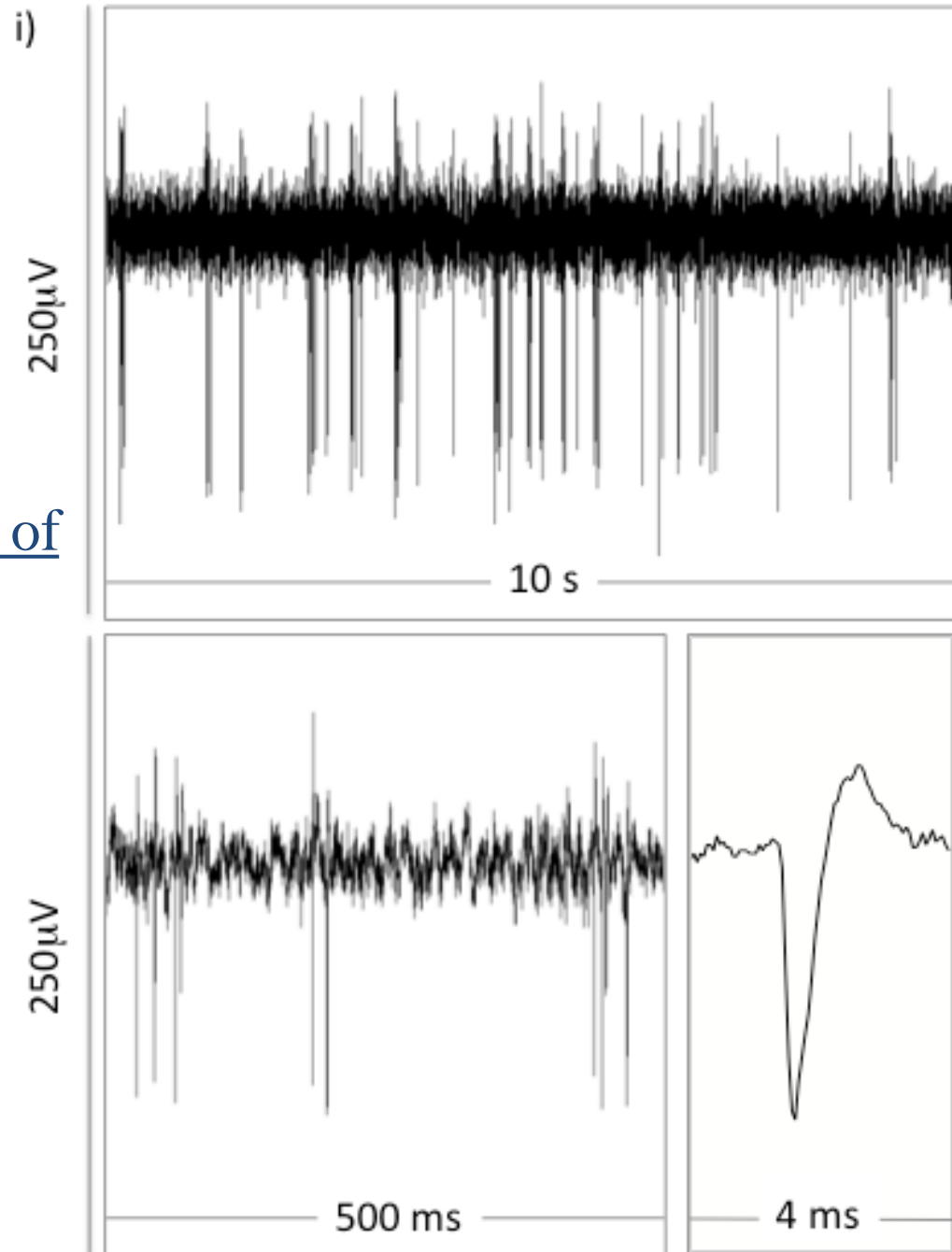


Height=150
microns;
Diameter at
base=30
microns

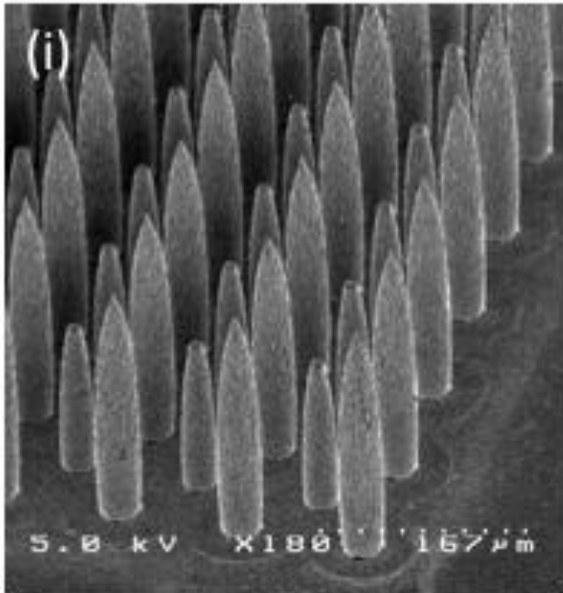


61
electrodes;
60 micron
spacing;
height=50
microns

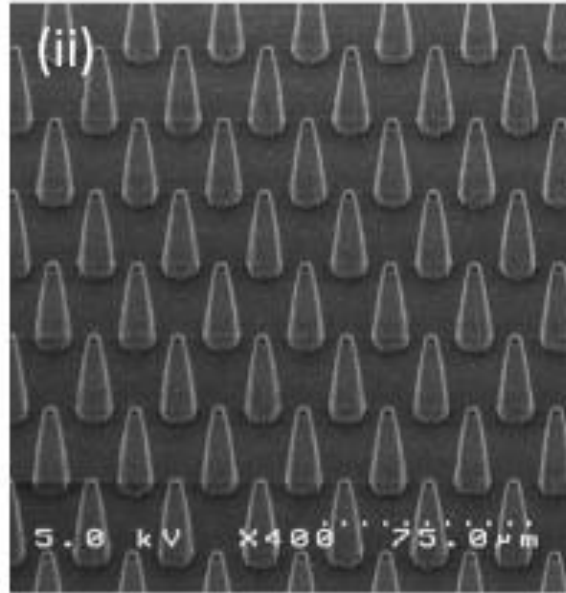
Bed-of-Nails Recording of
Neural Activity in a
Rat Acute Brain Slice



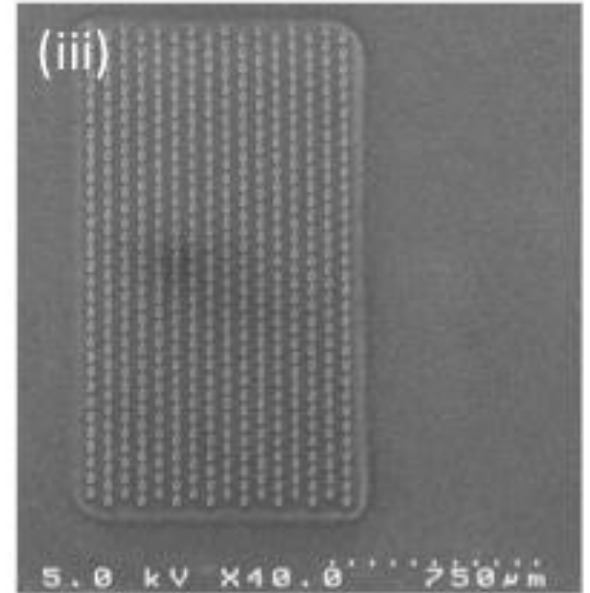
Mechanical prototypes
(fabricated by Debbie Gunning)



Multi-length needles



High density :
30 micron spacing



512 electrodes;
60 micron spacing

Wireless In-Vivo Brain Activity Recording System

Study the brain activity of awake, naturally-behaving animals
(e.g. burrowing, climbing, flying)

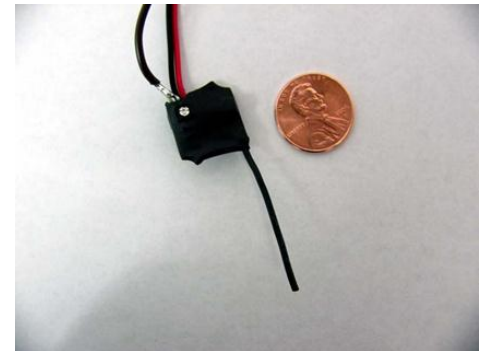
- Record brain activity on multiple (~64) electrodes (or ~16 tetrodes) with portable, battery-operated system that can be carried by a freely moving rat or a flying barn owl
- Neurochip-based system with wireless telemetry



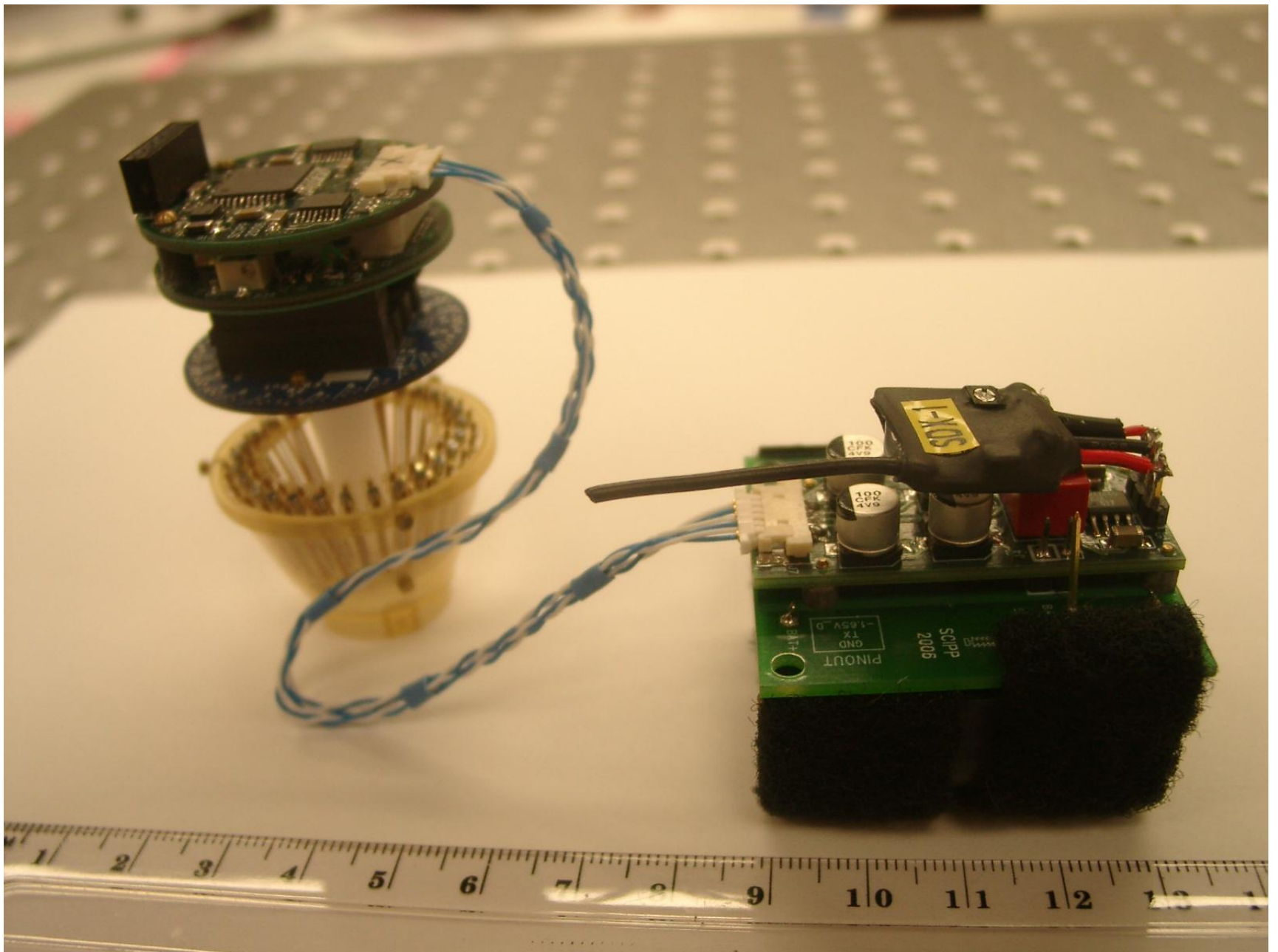
28 tetrodes
with microdrives
(Neuralynx)

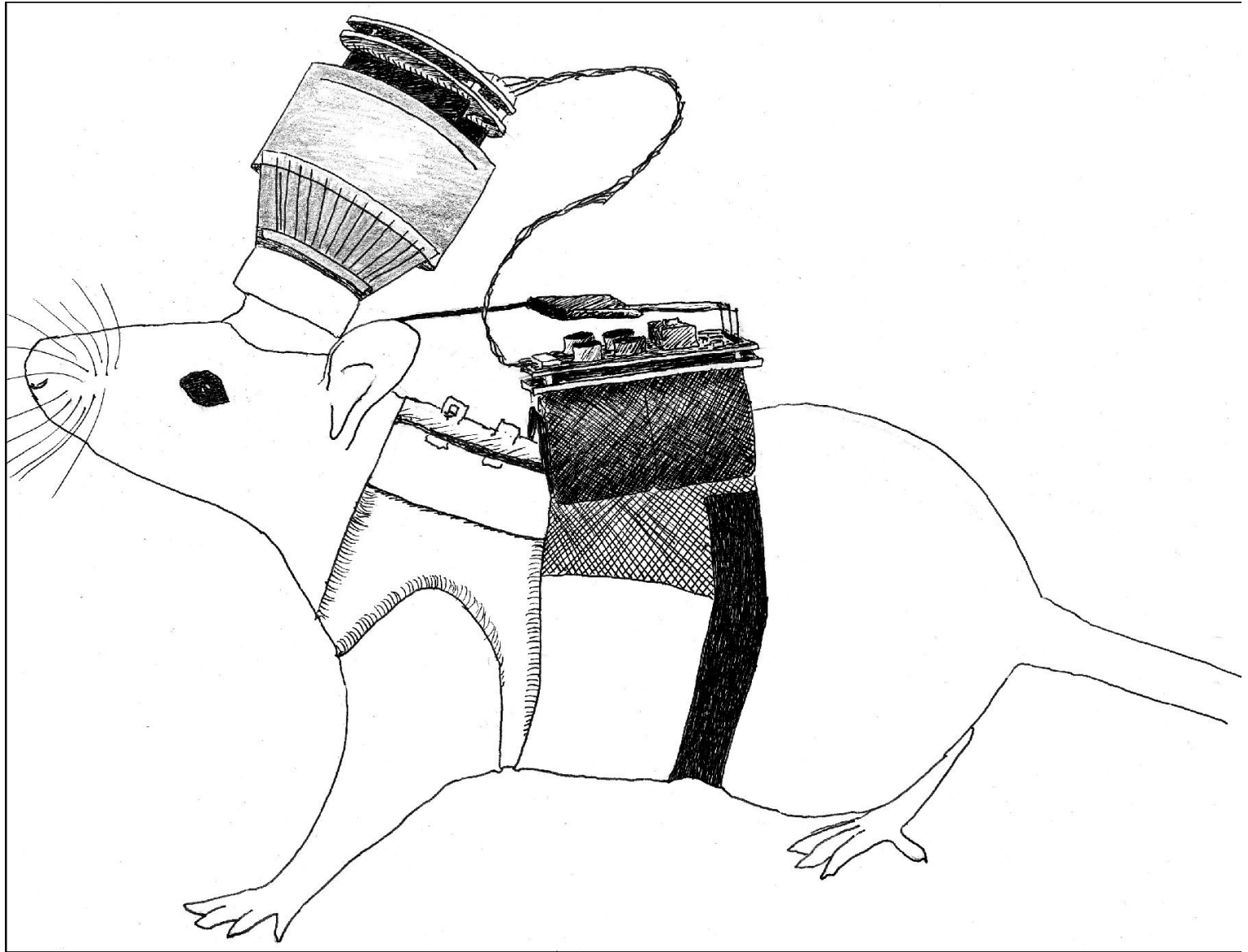


Neurochip
(Dabrowski)



Spy Transmitter
(Meister)

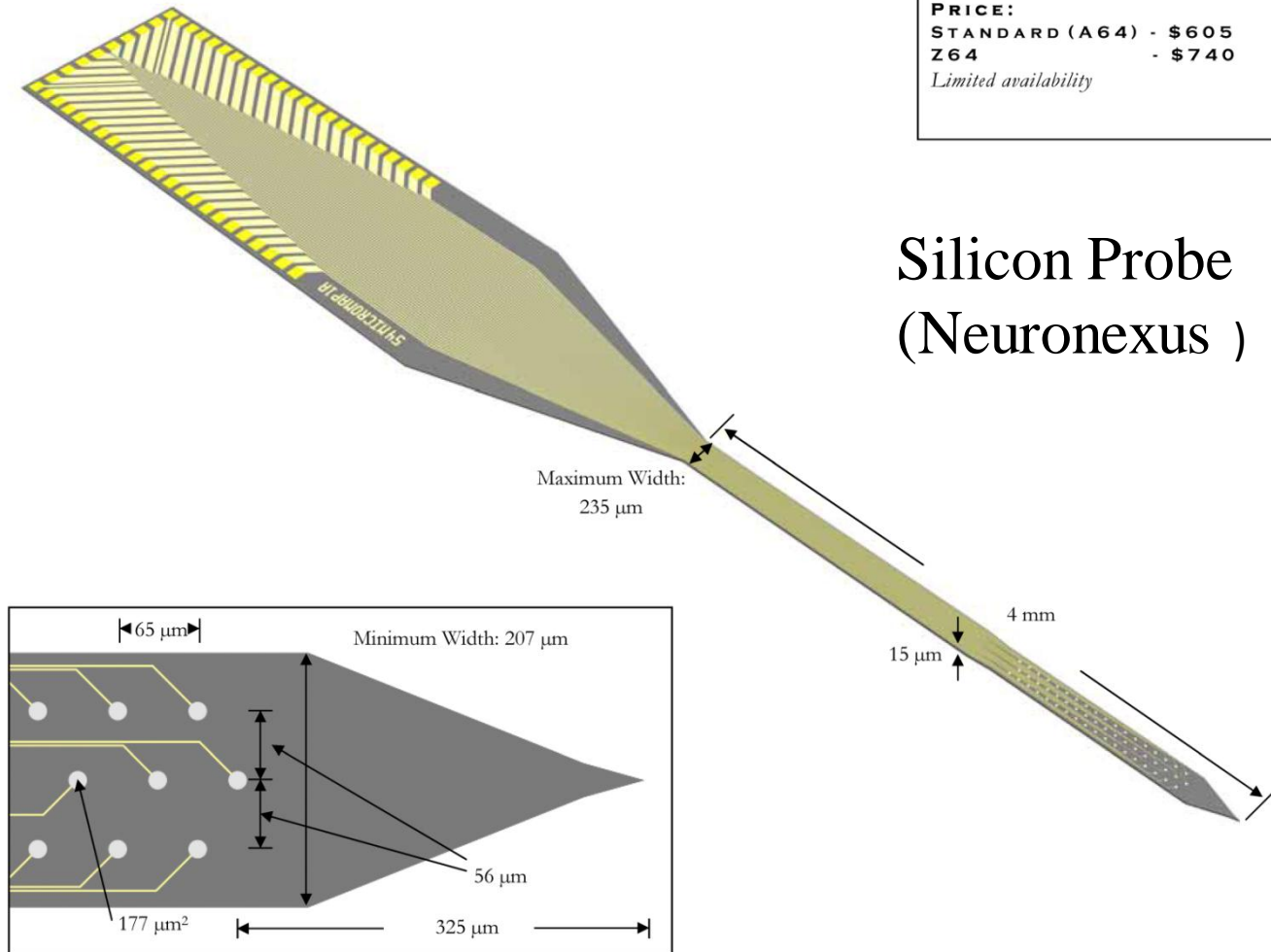


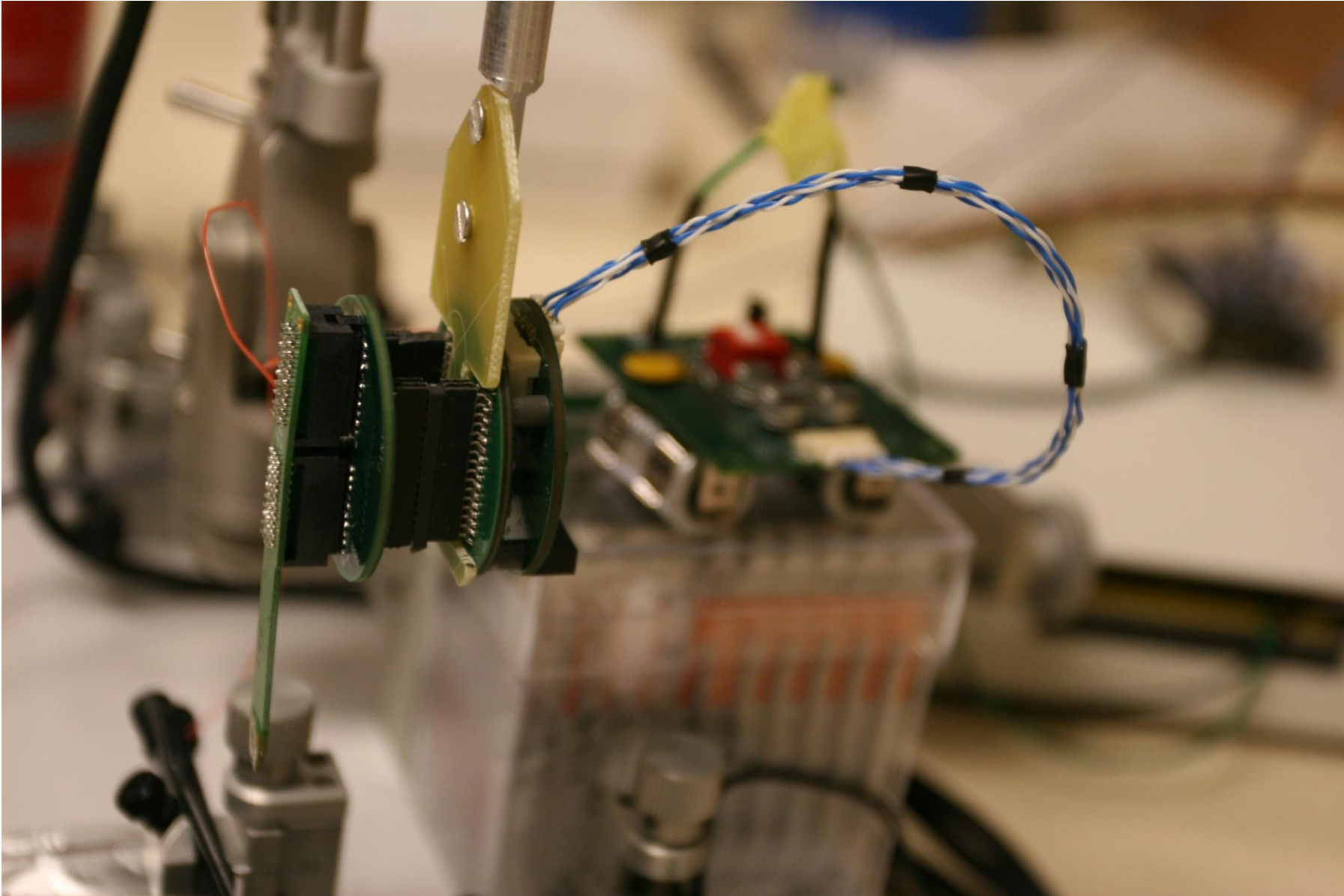


Wired System for Recording Brain Activity In Vivo with Silicon Probes (with Yi Zuo)

POLYTRODE 1A	
PRICE:	
STANDARD (A64)	- \$605
Z64	- \$740
<i>Limited availability</i>	

Silicon Probe (Neuronexus)





Thank you, Sherwood, for starting me off on this amazing journey.

You are one of my heroes. I have learned so much from you, sometimes much more than I realized I needed to know.

Decades before Google or Wikipedia or Scholarpedia, when I desperately needed information, I could depend on you for accurate and patient answers.

You are a National, even International, Resource. Keep it up!!

My Very Best wishes to you on your 80th birthday.